

## Thysanoptera

5800

, 89,

### Order Thysanoptera in Alfalfa Agroocenosis

Ivelina Nikolova

*Institute of Forage Crops, General Vladimir Vazov 89, 5800 Pleven, Bulgaria*

*E-mail: imnikolova@abv.bg*

Review paper

#### SUMMARY

Thrips are serious insect pests in various crops. Generally, they are highly adaptable. This adaptability is reflected in their ability to feed on different plant hosts, including alfalfa, and to cause substantial damage.

The literature review of Thysanoptera species in alfalfa agroocenosis included economically important thrips in the world and in Bulgaria. As a result of their nutritional activity, thrips damage vegetative and reproductive organs, causing economic losses and reduced forage and seed yields.

It indicated the harmful stages of the thrips development as well as sensitive alfalfa phases, through which the insect pest causes considerable damage.

**Key words:** Thysanoptera, species, mechanism of damage, alfalfa agroocenosis

#### STUDIES ABROAD

Thysanoptera order is known for about 6.000 species in the world (Bhatti, 2006; Thrips Wiki, 2015). They are classified in two suborders Terebrantia

Thysanoptera

: Thysanoptera,

Thysanoptera

6 000 (Bhatti,

2006; Thrips Wiki, 2015).

Terebrantia

Tubulifera,		and Tubulifera, containing six families, five of which belong to Terebrantia
(Adiheterothripidae,	Terebrantia	(Adiheterothripidae,
Fauriellidae, Merothripidae	Aeolothripidae,	Aeolothripidae,
- Tubulifera (Phlaeothripidae)	Thripidae)	Fauriellidae, Merothripidae Thripidae)
(Fauna Europaea Secretariat),	93%	(Fauna Europaea Secretariat).
	-	To the families and Thripidae Phlaeripididae
Thripidae Phlaeripididae (Mound,	1%	belongs 93% of thrips (Mound, 1997).
1997).		Globally, only 1% of the species
	-	described are economically harmful
(Mound and Teulon, 1995),		(Mound and Teulon, 1995) while the
	-	species numbers very rarely approached
Thysanoptera		10 in many continents (Gahukar, 2004).
10		
(Gahukar, 2004).		In that sense, one of the most harmful in
	-	Europe were considered <i>Frankliniella</i>
<i>Frankliniella occidentalis</i> Pergande,		<i>occidentalis</i> Pergande, <i>F. intonsa</i> Trybon
<i>F. intonsa</i> Trybon (Kirk and Terry, 2003;		(Kirk and Terry, 2003; Raspudi et al.,
Raspudi et al., 2009) <i>Thrips tabaci</i>		2009), and <i>Thrips tabaci</i> Lindeman (De
Lindeman (De Jager et al., 1997).	-	Jager et al., 1997). Serious harmful thrips
		in various crops usually were highly
		adaptable. That adaptability was reflected
	-	in their ability to feed on different host
		species and caused considerable damage
	-	(Morse and Hoddle, 2006).
(Morse and Hoddle, 2006).		Thrips rapidly increased their population
		density under favorable conditions using
		high reproductive potential and short
		breeding times (Mound, 1997).
(Mound, 1997).		Parthenogenesis, in combination with F1
		cross-over, can cause a female adult to
	F1,	form a whole new generation (Mound and
		Tuelon, 1995).
(Mound and		Also, parthenogenesis and crossover in
Tuelon, 1995).		insecticide-resistant female individuals
		can lead to rapid insecticide-resistant
		population growth. That further aided their
		adaptation.
		The relatively broad food tolerance
		increased thrips flexibility that allows
		invasive thrips to survived in a new
		environment. There was a limited diversity
		of hosts in this environment, which
		increased the stability of the population,
		even in the absence of the preferred plant
		species (Mound, 1997).

	(Mound, 1997).				
					The harmful thrip number on alfalfa strongly depended on the duration of the flowering stage and the quality of pollen (Trdan et al., 2005).
2005).		(Trdan et al.,			
	Thripidae	-	-		The Thripidae family is the largest family, including the most economically important thrips species of crop plants worldwide (Mound, 2002; Bhatti, 2006).
	,				
	-				
	Thysanoptera				
		(Mound, 2002;			
Bhatti, 2006).		1970			It contained 1970 species and 287 genera. On the leaves and/or the plant flowers the main thrips group fed, damaged and reproduced and only a few species were obligate predators (Mirab-balou et al., 2011b; ThripsWiki, 2015). Reitz et al. (2011) reported that the most important pests spreading not only in the US but also in Europe were <i>F. occidentalis</i> , <i>T. tabaci</i> , <i>T. palmi</i> Karny <i>Stenchaetothrips biformis</i> Bagnall. Several species of the <i>Scolothrips</i> genus are known as obligate predators of tetranychus mites (Mirab-balou et al., 2011b).
287	,				
	,	/			
	,				
		(Mirab-balou			
et al., 2011b;	ThripsWiki,	2015).			
-	,				
	,				
	, Reitz				
	(2011)		<i>F.</i>		
<i>occidentalis</i> ,	<i>T. tabaci</i> ,	<i>T. palmi</i>	Karny		
<i>Stenchaetothrips biformis</i>	Bagnall.				
<i>Scolothrips</i>					
	(Mirab-balou et al., 2011b).				
	Fauriellidae	3			The Fauriellidae family included 3 genera (Mound et al., 1980) and Adiheterothripidae - one genus and five species that were rather poorly known (Mound, 2002). Merothripidae Hood family included 15 species that usually inhabited dead twigs or leaf litter where they probably fed on fungal hyphae (Mound, 2011).
	(Mound et al., 1980),	Adiheterothripidae			
	-				
2002),		(Mound,			
15	Merothripidae	Hood			
	,				
		(Mound, 2011).			
	Aeolothripidae				
190	23				Aeolothripidae family included 190 species and 23 genera worldwide that occurred in the flowers of their hosts and fed mainly on the floral tissues of other Thysanoptera larvae species, and small arthropods (mites, psyllids, white wings, etc.) inhabiting the plant flowers (Tyagi et al., 2008; Mound, 2011; Thrips Wiki, 2015). Adult individuals and larvae of many species of that family were optional predators, although some were herbivorous (Tyagi et al., 2008).
,					
	Thysanoptera				
	(	,			
),					
	(Tyagi et al., 2008; Mound, 2011;	Thrips Wiki,	2015).		
Thrips Wiki,	2015).				
,					
	(Tyagi et al., 2008).				
Hoddle (2003),	-				According to Hoddle (2003), in warmer regions of the world, a considerable species

80%  
*Aeolothrips*

(Loomans et al., 1995).  
 Tubulifera  
 3500 450  
 – Phlaeothripidae  
 – Idolothripinae  
 Phlaeothripinae (Thrips Wiki, 2015).

(Mound, 2011).  
*Haplothrips*,  
 Asteraceae, Poaceae, Cyperaceae  
 Fabaceae (Mound and Minaei, 2007).  
 Thysanoptera

(Mirab-balou et al., 2011a).

Thysanoptera

number of that family were obligate predators. It was reported that 80% of the species belonging to the *Aeolothrips* genus were predators that played an important role, along with other beneficial agents, in the biological control of phytophagic thrips (Loomans et al., 1995).

The Tubulifera suborder consisted of about 3500 species and 450 genera included in one family - Phlaeothripidae and two subfamilies - Idolothripinae and Phlaeothripinae (Thrips Wiki, 2015). Some species fed on fungi, but most were pests and preferred to feed on leaves or flowers (Mound, 2011). One of the main pests found in different parts of the world were mainly species of the Haplothrips genus, and a large number of them living in the plant flowers of the family Asteraceae, Poaceae, Cyperaceae, and Fabaceae (Mound and Minaei, 2007).

Therefore, Thysanoptera is characterized by a diversity of biological species - predators, plant-eating and fungus-eating species.

Economically important alfalfa and other crop pests directly damaged plant tissue when fed or caused indirect damage through the transmission of pathogenic infections, in particular viruses (Mirab-balou et al., 2011a).

Thysanoptera species are very often meet in the agroecosystem of perennial legumes, including alfalfa.

The short development cycle under favorable conditions and their ability to spread rapidly (through natural flight, air masses, etc.) support their harmful activity.

They have a pierce-sucking oral apparatus through which they inject enzymes into the plant parts, causing biochemical changes. As a result, thrips suck out plant juice. Result of the inflicted damage is a disruption of plant growth and physiology, deformation and wrinkling

<p>(Summers et al., 2006).</p>	<p>- of the leaves due to unequal growth around the damaged area. When feeding is particularly close to the central fiber, the leaf curves and resembles a half-open funnel (Summers et al., 2006).</p>
<p>20% (Yongfu, 1991),</p>	<p>Thrips damage is not only a reduction in forage yield above 20% (Yongfu, 1991) but species are carriers of viral and other pathogenic infections, which cause various plant diseases.</p>
<p>(Mound, 2002; Razi et al., 2013).</p>	<p>Thrips can infect and transmit viral pathogens when fed through saliva as other pierce-sucking insects (Mound, 2002; Razi et al., 2013).</p>
<p>(2015) Thysanoptera, <i>Frankliniella occidentalis</i></p>	<p>In Central Greece, Badieritakis and Collective (2015) reported eight species of Thysanoptera, among which <i>F. occidentalis</i> was the most abundant in alfalfa with a tendency to integrate with other species.</p>
<p>Raspudi (2009) Thysanoptera Thripidae, <i>T. tabaci</i> Asteraceae Fabaceae,</p>	<p>Raspudi et al. (2009) studied Thysanoptera in Croatia on different hosts and found that the most numerous and species-rich was the Thripidae family. In that family, <i>Thrips tabaci</i> was a major dominant species in plant species of Asteraceae and Fabaceae, with a preference for alfalfa in legumes. The second most abundant species, according to the authors, was <i>Frankliniella intonsa</i>, found primarily in Asteraceae and Fabaceae, and again with a preference for <i>M. sativa</i>. Both species are known as widespread insect pests in Slovenia (Trdan, 2003), Serbia (Andjus and Trdan, 2005 a, b), Hungary (Jenser and Czencz, 1988) and in many other European countries with a continental climate.</p>
<p><i>intonsa</i>, Asteraceae Fabaceae, <i>M. sativa</i>.</p>	<p><i>F. intonsa</i>, found primarily in Asteraceae and Fabaceae, and again with a preference for <i>M. sativa</i>. Both species are known as widespread insect pests in Slovenia (Trdan, 2003), Serbia (Andjus and Trdan, 2005 a, b), Hungary (Jenser and Czencz, 1988) and in many other European countries with a continental climate.</p>
<p>(Trdan, 2003), (Andjus and Trdan, 2005 a,b), (Jenser and Czencz, 1988) <i>Frankliniella</i> <i>F. occidentalis</i>, <i>F. schultzei</i> Trybom <i>F. intonsa</i>, <i>Odontothrips</i> - <i>confusus</i> Priesner,</p>	<p>Common and major pests of alfalfa of the <i>Frankliniella</i> genus in Iran were <i>F. occidentalis</i>, <i>F. schultzei</i> Trybom and <i>F. intonsa</i>, and of the <i>Odontothrips</i> genus - <i>O. confusus</i> Priesner, which was widespread in most of Iran and inhabited</p>

(Mirab-balou and Chen, 2011 a, b).  
Summers  
(2006),

*Caliothrips fasciatus* Pergande /  
*T. tabaci*.

”  
*F. occidentalis*.

*O. confusus* (36.7%), *T. tabaci* (31%)  
*F. intonsa* (23%),

(Ábrahám, 2012).

*T. tabaci*, *T. angusticeps* Uzel, *F. occidentalis*,  
*F. intonsa*, *C. fasciatus*, *Haplothrips niger* Osborn,  
*O. confusus* (Funderburk, 2002).

Funderburk (2002),

*Frankliniella* Thrips.  
Lewis (1973),

( )  
Brodbeck  
(2002)

*Frankliniella occidentalis*

Thysanoptera

(Mirab-balou and Chen, 2011 a, b);

mainly alfalfa and clover flowers (Mirab-balou and Chen, 2011 a, b).

According to Summers et al. (2006), very high populations of *Caliothrips fasciatus* Pergande and / or *T. tabaci* were observed in alfalfa fields. Both species were considerably more "destructive" and harmful than, for example, *F. occidentalis*. Authors added that alfalfa grown no irrigation can be considerably more susceptible to damage than alfalfa when irrigated.

In Hungary, *O. confusus* (36.7%), *T. tabaci* (31%) and *F. intonsa* (23%) were reported as dominant phytophages which not only feed and damage but also reproduce on alfalfa plants (Ábrahám, 2012).

Several authors confirmed that one of the most important and common phytophagic thrips alfalfa were *T. tabaci*, *T. angusticeps* Uzel, *F. occidentalis*, *F. intonsa*, *C. fasciatus*, *Haplothrips niger* Osborn, *O. confusus* and others (Funderburk et al., 2002).

According to Funderburk (2002), the most common thrips species were those that damaged the reproductive organs - the flowers and mostly those of the *Frankliniella* and *Thrips* genus.

According to Lewis (1973), many species inhabited plant flowers as the latter not only provided a suitable microclimate for growth, development, and reproduction, but also protected thrips from exposure to external unfavorable factors through narrow slits in the plant flowers.

Also, Brodbeck et al. (2002) reported that insects preferred flowers to leaves because of their higher nitrogen content in pollen.

*Frankliniella occidentalis* is one of the main and dominant phytophages of the Thysanoptera order in alfalfa (Mirab-balou and Chen, 2011 a, b; Badieritakis et al.,

Badieritakis et al., 2015).	- 2015). The species has a cosmopolitan distribution and is a polyphage,
,	- characterized by a short reproductive period and a tendency for parthenogenesis (Kirk and Terry, 2003). Thrips feed on more than 500 plant species belonging to more than 50 families (Moritz, 2002).
(Kirk and Terry, 2003).	
500	
50	(Moritz, 2002).
Funderburk (2015)	, Reitz
<i>ccidental</i>	- <i>F.</i>
"	- Based on economic evaluations, Reitz and Funderburk (2015) found that <i>F. occidentalis</i> is one of the "most destructive" agricultural insect pests worldwide, and its appearance as a major pest occurred relatively recently. On the other hand, it was reported that <i>F. occidentalis</i> is a facultative predator and can eat with other phytophagous thrips, reducing their numbers and displacing them in interspecific competition (Faraji et al., 2002).
,	
,	
<i>F. occidentalis</i>	- The result of competition worldwide for the spread of <i>F. occidentalis</i> was that the species inhabited six continents.
,	
,	
(Faraji et al., 2002).	
<i>F. occidentalis</i>	
,	
,	
(Moritz et al., 2004, Kirk and Terry, 2003).	All of them had local thrips species inhabiting the local flora (Moritz et al., 2004, Kirk and Terry, 2003). If <i>F. occidentalis</i> can compete with the local species, then that will increase the invasive threat from the economically harmful Thysanoptera species on alfalfa and other crops (Northfield, 2005).
<i>F. occidentalis</i>	
,	
Thysanoptera	
(Northfield, 2005).	
Ripa	- Ripa et al. (2009), examining <i>F. occidentalis</i> preferences for plant organs, found that thrips predominated on the leaves compared with buttons, but there were considerably more adult individuals when the plants were in the flowering stage.
<i>F. ccidental</i>	
,	
,	
,	
,	
,	
,	
,	
,	
,	An important dominant pest in

*intonsa*, (Deligeorgidis et al., 2002).  
(Murai and Loomans, 2001).  
, Atakan Uygur (2005)  
*F. intonsa*  
*occidentalis*, – *F. gemina* (Bagnall) (de Borbon et al., 2006).  
(Hansen et al., 2003).  
(Brodbeck et al., 2002).  
*intonsa*  
*F. occidentalis*  
*F. intonsa* (Tao et al., 2011).  
*Thrips tabaci*  
(Boateng et al., 2014).  
41 (Ghabn, 1948),  
355 (Morison, 1957),  
140 (Ananthakrishnan, 1973).  
(Diaz-Montano et al., 2011).

*F.* alfalfa was *F. intonsa*, due to its high population density and polyphagia (Deligeorgidis et al., 2002). The species was a facultative phytophage and pollen was an important nutrient source for it (Murai and Loomans, 2001).  
In Turkey, Atakan and Uygur (2005) reported that *F. intonsa* displaced *F. occidentalis* in numbers and Argentina - *F. gemina* (Bagnall) (de Borbon et al., 2006). That could be due to competition between species or other factors that regulated population numbers (Hansen et al., 2003). Insect fecundity (including development, longevity, and oviposition) depended on the food quality (Brodbeck et al., 2002).  
The degree of rapid development and high fecundity were a sure sign for the choice of suitable hosts.  
Also, the survival rate, life duration, fecundity and vital parameters of *F. occidentalis* and *F. intonsa* were mainly influenced by temperature, as the life duration of both species decreased with increasing temperature (Tao et al., 2011). *Frankliniella intonsa* had a higher chance of survival at higher temperature values.  
*Thrips tabaci* was a cosmopolitan polyphage that was characterized by a wide host variety (Boateng et al., 2014). Some authors reported that the species was found in 141 plant species from 41 families (Ghabn, 1948), others informed for 355 host species (Morison, 1957), and the third - to 140 plant species from 40 families (Ananthakrishnan, 1973).  
Over the past two decades, the species became a global insect pest, due to development of insecticide resistance, its ability to transmit infectious pathogens, and its high reproductive capacity at high temperatures, especially in dry and hot weather (Diaz-Montano et al., 2011).



<p><i>T. tabaci</i>, (Parrella and Lewis, 1997).</p>	<ul style="list-style-type: none"> <li>- Damage caused by <i>T. tabaci</i> led to a decrease in the photosynthetic activity of the plants by destroying the chlorophyll pigments in the leaf mesophyll (Parrella and Lewis, 1997).</li> <li>- That may impede the transport of nutrients to the generating organs. Because individual plant cells were destroyed as a result of the nutrition, leaf symptoms were in the form of silver spots.</li> </ul>
<p>(Pushpendra et al., 2014).</p>	<ul style="list-style-type: none"> <li>- Stains had good visibility due to the suctioned cell voids. Although the individual leaf spots on feeding were small, the damaged area of the leaf was not refundable. The area became larger and more distinct as the plant grows. Due to the rapid reproduction and high mobility, thrips had become increasingly important over the years (Pushpendra et al., 2014).</li> </ul>
<p><i>T. tabaci</i>, (Lewis, 1973).</p>	<ul style="list-style-type: none"> <li>- Losses due to the attack of <i>T. tabaci</i> depended on many factors, including the thrip numbers, favorable weather conditions for their reproduction, plant development stage, the sensitivity of the varieties to nutrition and damage, the presence of plant infection by vector thrips (Lewis, 1973).</li> </ul>
<p>24- Pobo niak Koschier (2014) <i>tabaci</i></p>	<ul style="list-style-type: none"> <li>- Concerning the timing of the 24-hour daily cycle, Pobo niak and Koschier (2014) reported that the peak activity of <i>T. tabaci</i> was at noon and regardless of the plant development stage, most of the adult individuals were located at the top of the plants.</li> </ul>
<p><i>Frankliniella Thrips</i>,</p>	<ul style="list-style-type: none"> <li>- That should be considered in the monitoring, as well as developing methods for proper sampling and implementing appropriate control strategies as needed.</li> <li>- It is assumed that species of the <i>Frankliniella</i> and <i>Thrips</i> genus, due to its wide polyphagia, overtaking the capacity</li> </ul>

(Mound and Teulon, 1995).	-	of its natural bio-agents to regulate their population density (Mound and Teulon, 1995). Recent studies indicated that the species <i>Orius</i> (Heteroptera: Anthocoridae) and <i>Thripinema</i> (Tylenchida: Allantonematidae) played an important role as entomophagous in flower thrips, suppressed pest populations and caused local numerical reductions (Funderburk et al., 2000).
<i>Orius</i> (Heteroptera: Anthocoridae)	-	
<i>Thripinema</i> (Tylenchida: Allantonematidae)	-	
(Funderburk et al., 2000).	-	
<i>Odontothrips</i> (Thripidae)	-	The <i>Odontothrips</i> (Thripidae) genus was cited by many authors as a pest on legumes that fed on the plant flowers (Northfield, 2005). Several species of the genus, such as phytophages in alfalfa, were reported. For example, <i>O. confusus</i> was reported as a pest in alfalfa in France and the Czech Republic (Pitkin, 1972), as well as in other leguminous crops in Romania (Pustai et al., 2016). In China, one of the dominant species of Thysanoptera in alfalfa was <i>O. lotti</i> Haliday, such as LiLi et al. (2011), studying the spatial distribution of the species, through flowering, found that it had a homogeneous distribution or a randomly distributed structure on a horizontal level. The authors added to the decreasing tendency of its abundance from the top of the plants to the root at a vertical level.
(Northfield, 2005).	-	
<i>O. confusus</i>	-	
(Pitkin, 1972),	-	
(Pustai et al., 2016).	-	
Thysanoptera	-	
<i>O. lotti</i> Haliday, LiLi	-	
(2011),	-	
<i>Odontothrips melliloti</i> Priesner	-	<i>Odontothrips melliloti</i> Priesner was a recently reported pest in alfalfa crop for China's fauna (Mirab-balou et al., 2011a).
(Mirab-balou et al., 2011a).	-	
Aeolothripidae,	20	In Europe, from the Aeolothripidae family, more than 20 predatory species of the Aeolothrips genus was described, as most of them belong to facultative predators (Trdan et al., 2005). <i>Aeolothrips intermedius</i> Bagnall was the most common species of that family on the Old Continent (Deligeorgidis et al., 2002) and was often mentioned in the context of biological control (von Zegula et al., 2003). The species was usually not only the most numerous of the Aeolothripidae but it was also a predator with economic
Aeolothrips,	-	
(Trdan et al., 2005). <i>Aeolothrips intermedius</i> Bagnall	-	
(Deligeorgidis et al., 2002)	-	
(von Zegula et al., 2003).	-	
Aeolothripidae,	-	

(Trdan et al., 2005).  
*A. intermedius*  
 44 Thysanoptera (Loomans et al., 1995),

*T. tabaci* (Franco et al., 1999),  
*Heliothrips haemorrhoidalis* Bouche,  
*Odontothrips confusus* Priesner  
*Haplothrips* (Khosbayar, 2001).  
*Aeolothrips* spp.

(Sabelis and van Rijn, 1997). ( 100 )

(Von Zegula et al., 2003).

*A. intermedius*  
 , Trdan  
 (2005)  
 30  
 16

, *A. intermedius*

*Haplothrips*  
*(Haplothrips kurdjumovi* *H. subtilissimus*,  
 Phlaeothripidae), *Scolothrips*  
 (Thripidae), *Franklinothrips*  
*Karnyothrips* (Tyagi et al., 2008).

*Frankliniella*  
*intonsa*, *T. tabaci*, *Taeniothrips frici*,

importance for phytophagous thrips and mites (Trdan et al., 2005). *Aeolothrips intermedius* was considered a potentially very effective autochthonous facultative predator among the zoophagous thrips in Europe. Adults and larvae feed on 44 species of phytophagous of the Thysanoptera order (Loomans et al., 1995), and commonly reported as an effective predator of *T. tabaci* (orres-Vila et al., 1994; Franco et al., 1999), as well as *Heliothrips haemorrhoidalis* Bouche, *Odontothrips confusus* Priesner and species of the genus *Haplothrips* (Khosbayar, 2001). Some other species of *Aeolothrips* spp. were also considered potentially important autochthonous facultative predators (Sabelis and van Rijn, 1997). Most of the species in that genus (about 100 species) spend much of his life in plant flowers and feed on pollen and small arthropods, (Yee et al., 2001; Von Zegula et al., 2003).

As a result of an extensive study of *A. intermedius* on different plant species in Slovenia, Croatia and Serbia, Montenegro, Trdan et al. (2005) found it on 30 different crops, belonging to 16 botanical families. In alfalfa, as well as in other hosts, *A. intermedius* was always present in mixed populations with phytophagous or facultative phytophagous insects.

The authors reported that pollen, as an alternative food for the predator, played an extremely important role in its development and reproduction.

Other well-known predators in Europe were species of the *Haplothrips* genus (*Haplothrips kurdjumovi* and *H. subtilissimus*, Phlaeothripidae), *Scolothrips* (Thripidae), *Franklinothrips* and *Karnyothrips* (Tyagi et al., 2008).

#### **STUDIES IN BULGARIA**

In Bulgaria, as harmful thrips on alfalfa, were reported *Frankliniella intonsa*, *T. tabaci*, *Taeniothrips frici*,

*Haplothrips aculeatus*, *H. angusticornis*  
 . (Donchev, 1972, 1976). Karadjova  
 Krumov (2015)

26

Thripidae.  
 Tomov (2010)

*F. occidentalis*

Trenchev (1991),  
*F. occidentalis*,

Thysanoptera

*Haplothrips aculeatus*, *H. angusticornis*  
 and others (Donchev, 1972, 1976).

- Karadjova and Krumov (2015) summarize  
 - all published data on the occurrence and  
 - spread of thrips in Bulgaria, as well as  
 - information about their preference for  
 - eating. The authors reported that 26  
 - phytophagous and entomophagous  
 - species of thrips were found in alfalfa,  
 - with predominant species diversity of the  
 - Thripidae family.

- Tomov et al. (2010) reported that *F.*  
*occidentalis* was one of eleven invasive  
 - insect species imported into Bulgaria that  
 - were economically important crop pests.

- They posed a serious threat to the  
 - economy, biodiversity, human and animal  
 - health. According to Trenchev (1991), the  
 - biggest damage caused by *F. occidentalis*  
 - was the transmission of tospoviruses.

- Studies on the species composition  
 - and seasonal dynamics of alfalfa thrips in  
 - Bulgaria are unsatisfactory and scarce.  
 - That kind of research is important  
 - because it helps to identify the  
 - appropriate plant development stage for  
 - pest control when the economic threshold  
 - of damage is exceeded. They are also at  
 - the service of Bulgarian farmers for  
 - reducing losses from insect pests.

- Based on the literature review  
 - presented, it can be summarized that  
 - most of the Thysanoptera order are  
 - polyphages, allowing them to survive and  
 - develop in different agroecosystems. Some of  
 - them pose a constant threat to the  
 - cultivation of both alfalfa and other  
 - cultivated species.

- Therefore, knowledge of the  
 - abundance and species diversity of alfalfa  
 - thrips is extremely useful for both  
 - fundamental entomology and scientifically  
 - applicable.

## / REFERENCES

1. **Ábrahám, R.**, 2012. First investigation of species composition of thysanoptera inhabiting alfalfa based on their second stage larvae. *Acta Phytopathologica et Entomologica Hungarica*, 47 (1), 81-86.
2. **Ananthakrishnan, T.N.**, 1973. Thrips: biology and control. MacMillan, New Delhi, India.
3. **Andjus, Lj. and S. Trdan**, 2005a. Tobacco thrips (Thrips tabaci Lindeman), the most harmful type of thrips outside. *Herbal medicine*, 33, 395-400 (Croatia).
4. **Andjus, Lj. and S.Trdan**, 2005b. Thrips in alfalfa. *Herbal medicine*, 33, 538-542 (Croatia).
5. **Atakan, E. and S. Uygur**, 2005. Winter and spring abundance of Frankliniella spp. and Thrips tabaci Lindeman (Thysan., Thripidae) on weed host plants in Turkey. *Journal of Applied Entomology*, 129, 17-26.
6. **Badieritakis, E.G., R.C. Thanopoulos, A.A. Fantinou and N.G. Emmanouel**, 2015. A qualitative and quantitative study of thrips (Thysanoptera) on alfalfa and records of thrips species on cultivated and wild Medicago species of Greece. *Biologia*, 70 (4), 504-515.
7. **Bhatti, J.S.**, 2006. The classification of Terebrantia (Insecta) into families. *Oriental Insects*, 40, 339-375.
8. **Boateng, C.O., H.F. Schwartz, M.J. Havey and K. Otto**, 2014. Evaluation of Onion Germplasm for Resistance to Iris Yellow Spot (Iris yellow spot virus) and Onion Thrips, Thrips tabaci. *Southwestern Entomologist*, 39(2), 237-260.
9. **Brodbeck, B.V., J. Funderburk, J. Stavisky, PC. Andersen and J. Hulshof**, 2002. Recent advances in the nutritional ecology of thrips, or the lack thereof, pp. 145-153. In R. Marullo and L. Mound (eds.) Thrips and tospoviruses: proceedings of the 7th International Symposium on Thysanoptera. CSIRO Entomology, Canberra, Australia.
10. **De Borbon, C.M., O. Gracia and R. Piccolo**, 2006. Relationships between tospovirus incidence and thrips populations on tomato in Mendoza, Argentina. *Journal Of Phytopathology*, 154, 93-99.
11. **de Jager, C.M., R.P.T. Butot, M.E.C. Uiterdijk and E. van der Meijden**, 1997. Environmental influences on feeding damage caused by western flower thrips (Thysanoptera: Thripidae) to chrysanthemum. *Journal of Economic Entomology*, 90, 188-194.
12. **Deligeorgidis, P.N., C.G. Athanassiou and N.G. Kavallieratos**, 2002. Seasonal abundance, spatial distribution and sampling indices of thrips populations on cotton; a 4-year survey from central Greece. *Journal of Applied Entomology*, 126, 343-348.
13. **Diaz-Montano, J., M. Fuchs, B.A. Nault, J. Fail and A.M. Shelton**, 2011. Onion thrips (Thysanoptera: Thripidae): a global pest of increasing concern in onion. *Journal of Economic Entomology*, 104, 1-13.
14. **Donchev, K.**, 1976. Contribution to Thysanoptera Classification in Bulgaria III. *Bulgarian Journal of Crop Science*, XIII (1), 175-181 (Bg).
15. **Donchev, K.**, 1972. Contribution to Thysanoptera in Bulgaria II. *Bulgarian Journal of Crop Science*, IX (3), 131-135 (Bg).
16. **Faraji, F., A. Janssen and M.W. Sabelis**, 2002. Oviposition patterns in a predatory mite reduce the risk of egg predation caused by prey. *Ecological Entomology*, 27, 660-664.
17. **Franco, S., P. Beignet, E. Rat and E. Thibout**, 1999. The effects of thrips on wild and cultivated alliaceous plants in France. *Phytoma*, 514, 41-44.

18. **Funderburk, J.**, 2002. Ecology of Thrips. In Mound LA, Marullo R (Eds) Thrips and Tospoviruses: Proceedings of the 7th International Symposium on Thysanoptera. Reggio Calabria, Italy: *CSIRO Entomology*, pp.121-128.
19. **Funderburk, J., J. Stavisky and S. Olson**, 2000. Predation of *Frankliniella occidentalis* (Thysanoptera: Thripidae) in field peppers by *Orius insidiosus* (Hemiptera: Anthocoridae). *Environmental Entomology*, 29, 376-382.
20. **Gahukar, R.T.**, 2004. Bionomics and management of major thrips species on agricultural crops in Africa. *Outlook Agric* 33, 191-199.
21. **Ghabn, E.**, 1948. Contribution to the knowledge of the biology of *Thrips tabaci* Lind. *Bulletin of the Entomological Society of Egypt*, 32, 123-174.
22. **Hansen, E.A., J.E. Funderburk, S.R. Reitz, S. Ramachandran, J.E. Eger and H. MacAuslane**, 2003. Within-plant distribution of *Frankliniella* species (Thysanoptera: Thripidae) and *Orius insidiosus* (Heteroptera: Anthocoridae) in field pepper. *Environmental Entomology*, 32, 1035-1044.
23. **Hoddle, M.S.**, 2003. The effect of prey species and environmental complexity on the functional response of *Franklinothrips orizabensis*: a test of the fractal foraging model. *Ecological Entomology*, 28, 309-318.
24. **Jenser, G. and K.Czencz**, 1988. Thysanoptera species occurring frequently on cultivated plants in Hungary. *Acta Phytopathologica et Entomologica Hungarica*, 23, 285-289.
25. **Karadjova, O. and V. Krumov**, 2015. Thysanoptera of Bulgaria. *Zookeys*, (504), 93-131.
26. **Kirk, W.D.J. and L.I. Terry**, 2003. The spread of the western flower thrips *Frankliniella occidentalis* (Pergande). *Agricultural and Forest Entomology*, 5, 301-310.
27. **Lewis, T.**, 1973. Thrips - Their Biology, Ecology and Economic Importance. London, Academic Press. 349 pp.
28. **LiLi, Z., W. PuChang, L. YuLiang and C. YanXi**, 2011. Study on spatial distribution pattern of thrip (*Oditothrips liti* Haliday, 1852) on alfalfa (*Medicago sativa*). *Agricultural Science & Technology*, 12 (7), 990-993.
29. **Loomans, A.J.M., J.P. Van Lenteren, M.G. Tommasini, S. Maini and J. Riudavets**, 1995. Biological control of thrips pests. *Wageningen agricultural university papers*, 95-1, 1- 201.
30. **Mirab-balou, M. and X.-X. Chen**, 2011b. Iranian thripinae with ctenidia laterally on the abdominal tergites (Thysanoptera: Thripidae). *Natura Montenegrina, Podgorica*, 10 (4), 435-466.
31. **Mirab-balou, M. and X.-X. Chen**, 2011 . The Megalurothrips genus-group in Iran (Thysanoptera: Thripidae). *Munis Entomology & Zoology*, 6 (2), 944-952.
32. **Moritz, G.**, 2002. The biology of thrips is not the biology of their adults: a developmental view. Thrips and tospoviruses: Proceedings of the 7th International Symposium on Thysanoptera, pp. 259-267.
33. **Moritz, G., L.A. Mound, D.C. Morris and A. Goldarazena**, 2004. Pest thrips of the world: an identification and information system using molecular and microscopical methods (CD-ROM). Centre for Pest Information and Transfer, Brisbane, Australia.
34. **Morse, J.G. and M.S. Hoddle**, 2006. Invasion biology of thrips. *Annual Review of Entomology*, 51, 67-89.
35. **Mound, A.L.**, 1997. Thrips as crop pests. *Biological Diversity*, CAB Int., 198-215.
36. **Mound, L.A.**, 2011. Thysanoptera (Thrips) of the World-a checklist. Electronic Database accessible at <http://www.ento.csiro.au/thysanoptera/worldthrips.html>.
37. **Mound, L.A. and D.A.J. Teulon**, 1995. Thysanoptera as phytophagous opportunists, pp. 3-19. In B. L. Parker, M. Skinner, and T. Lewis (eds.), Thrips biology

and management. Plenum, New York.

38. **Mound, L.A. and K. Minaei**, 2007. Australian thrips of the Haplothrips lineage (Insecta: Thysanoptera). *Journal of Natural History*, 41, 2919-2978.
39. **Mound, L.A.**, 2002. So many thrips – so few tospoviruses? Proceedings of the 7th International Symposium on Thysanoptera, 15-18.
40. **Mound, L.A., B.S. Heming and J.M. Palmer**, 1980. Phylogenetic relationships between the families of recent Thysanoptera. *Zoological Journal of the Linnean Society of London*, 69, 111-141.
41. **Murai, T. and A.J.M. Loomans**, 2001. Evaluation of an improved method for mass-rearing of thrips and a thrips parasitoid. *Entomologia Experimentalis Et Applicata*, 101, 281-289.
42. **Northfield, T.D.**, 2005. Thrips competition and spatiotemporal dynamics on reproductive hosts. A Thesis for the degree of Master of Science, University of Florida, 69p.
43. **Parrella, M.P. and T. Lewis**, 1997. Integrated pest management in field crops, pp. 595- 614. In T. Lewis (ed.), *Thrips as crop pests*. CAB Inter., New York.
44. **Poboniak, M. and E. Koschier**, 2014. Effects of pea (*Pisum sativum* L.) cultivars on Thrips tabaci Lindeman preference and performance. *The Journal of Agricultural Science*, 152 (6), 885-893.
45. **Pushendra, E., A.S. Thakur, M. Thomas, A.K. Bhowmick and H.L. Sharma**, 2014. Screening of onion genotypes against Thrips tabaci Lind. in Central India. *Internal Journal of Biodiversity and Conservation*, 6 (12), 806- 813.
46. **Pustai, P.M., I. Oltean, V. Florian and T. Florian**, 2016. Assessment of soil entomofauna in crops of forage legumes with traps supply units in 2015. *ProEnvironment*, 9, 404-407.
47. **Raspudi, E., M. Ivezi, M. Brmež and S. Trdan**, 2009. Distribution of Thysanoptera species and their host plants in Croatia. *Acta Agriculturae Slovenica*, 93(3), 275-283.
48. **Razi, S., M. Laamari, S. Ouamen and E.C. Bernard**, 2013. Thysanoptera survey on *Vicia faba* (broad bean) in the arid Biskra region of Algeria. *Agriculture and Biology Journal of North America*, 4(3), 268-274.
49. **Reitz, S.R. and J. Funderburk**, 2015. Management strategies for western flower thrips and the role of insecticides. In: F. Perveen ed., *Insecticides – Pest Engineering*, 355-384.
50. **Ripa, R., J. Funderburk, F. Rodriguez, F. Espinoza and L. Mound**, 2009. Population abundance of *Frankliniella occidentalis* (Thysanoptera: Thripidae) and natural enemies on plant hosts in Central Chile. *Environmental Entomology*, 38 (2), 333-344.
51. **Sabelis, M.W. and P.C.J. van Rijn**, 1997. Predation by insects and mites. In: Lewis T (ed) *Thrips as crop pests*. CAB Int, *Wallingford*, 259-354.
52. **Summers, C.G., L.D. Godfrey, M. Rethwisch, D.R. Haviland, P.B. Goodell and R.F. Long**, 2006. Alfalfa thrips. UC IPM Pest Management Guidelines: Alfalfa. UC IPM Online Available at: <http://www.ipm.ucdavis.edu/PMG/r1302011.html> [Accessed 21 March 2012].
53. **Tao, G.H., Z. JunRui and S. Meng**, 2011. Effects of temperature on the survival and fecundity of *Frankliniella occidentalis* and *Frankliniella intonsa* (Thysanoptera: Thripidae). *Acta Phytophylacica Sinica*, 38 (6), 521-526.
54. **ThripsWiki**, 2015. ThripsWiki-providing information on the World's thrips. [cited 7 Aug. 2015.] Available from: <http://thrips.info/wiki>
55. **Tomov, R., K. Trencheva and G. Trenchev**, 2010. Foreign insects as a

problem for the sustainable development of Bulgaria. *Governance and Sustainable Development*, 1/2010 (25), 382-389 (Bg).

56. **Trdan, S.**, 2003. The occurrence of thrips species from the Terebrantia suborder on cultivated plants in Slovenia. Res. Rep., Biotech. Fac., Univ. Ljublj., *Agriculture*, 1, 81, 57-64.

57. **Trdan, S., L. Andjus, E. Raspudi and M. Ka** , 2005. Distribution of *Aeolothrips intermedius* Bagnall (Thysanoptera: Aeolothripidae) and its potential prey Thysanoptera species on different cultivated host plants. *Journal of Pest Science*, 78 (4), 217-226.

58. **Trenchev, G.**, 1991. A New Dangerous insect pest, *Earth*, p. 115 (Bg).

59. **Tyagi, K., V. Kumar and L.A. Mound**, 2008. Sexual dimorphism among Thysanoptera Terebrantia, with a new species from Malaysia and remarkable species from India in Aeolothripidae and Thripidae. *Insect Systematics and Evolution*, 39 (2), 155-170.

60. **von Zegula, T., C. Sengonca and P. Blaeser**, 2003. Entwicklung, Reproduktion und Prädatationsleistung von zwei Raubthrips-Arten *Aeolothrips intermedius* Bagnall und *Franklinothrips vespiformis* Crawford (Aeolothripidae) mit Ernährung zweier natürlicher Beutearten. *Gesunde Pflanz*, 55, 169-174.

61. **Yongfu, W.**, 1991. A study on the alfalfa thrips. *Acta Agrestia Sinica*, 01.



## Hemiptera, Cicadomorpha

## Fulgoromorpha

5800 , . , 89,

### Order Hemiptera, Suborder Fulgoromorpha and Cicadomorpha in Alfalfa Agrocenosis

Ivelina Nikolova

*Institute of Forage Crops, General Vladimir Vazov 89, 5800 Pleven, Bulgaria*

*E-mail: imnikolova@abv.bg*

*Review paper*

Cicadomorpha Fulgoromorpha  
Hemiptera

42 000

#### SUMMARY

Suborders Fulgoromorpha and Cicadomorpha in Hemiptera order are a highly diverse group of insects containing about 42,000 species described from around the world. They are spread everywhere, and all cicada species eat and damage plants.

The literature review covered the composition of species of cicada insects in alfalfa agrocenosis. They are diverse and numerous group that causes direct and indirect plant damage. Main and economically important cicadas are presented in different regions of the world and in Bulgaria, which as a result of their nutritional activities lead to a decrease in productivity and its quality.

It is described the damage mechanism to the dominant species and the injury occurring as a response in the plant organism. Cicada ability to transfer phytopathogenic organisms (viruses, mycoplasma, spiroplasma, bacteria) that

Cicadomorpha, : Fulgoromorpha,

cause various plant diseases is underlined.

**Key words:** Fulgoromorpha, Cicadomorpha, cicadas, damage, alfalfa agroecosis

Cicadomorpha Fulgoromorpha Hemiptera -  
42 000 (Mifsud et al., 2010; Cryan and Urban, 2011).

**STUDIES ABROAD**

Fulgoromorpha and Cicadomorpha suborder (order Hemiptera) are extremely diverse group of insects, containing about 42,000 described species worldwide (Mifsud et al., 2010; Cryan and Urban, 2011). They are everywhere spread and eat and damage plants.

(Hall, 2009).

Their common feature is the send out of clear and strong signals or specific vibration as a form of communication and reproductive behavior (Hall, 2009). Species of the two subdivisions have different degrees of specialization. Some of them are very broad polyphagous, others are oligophagous and third - monophagous, specialized on only one host species.

and Grimaldi, 2004). (2004) Ritzmann

These insects have a high reproductive ability, pronounced sexual dimorphism, ability to fly, great species diversity and distribution, small size (ngel and Grimaldi, 2004). According to Ritzmann et al. (2004) their last feature allowed settling in almost all ecosystems.

Fulgoromorpha -  
10 000 20 (Holzinger et al., 2003).

Fulgoromorpha suborder covers a large group of phytophagous insects that are distributed worldwide. It includes about 10, 000 described species, divided into 20 families (Holzinger et al., 2003).

This suborder is represented by some of the most harmful cicadas in major crops in the world. The relationship of this group to host plants is particularly important, because they use the plant organism not only as a source of food but also as a suitable site for fecundation and egg production, and as a means of communication (Holzinger et al., 2003).

(Holzinger et al., 2003).  
Fulgoromorpha

(Kastal'eva et al., 2016).

Cixiidae

Delphacidae

(Nickel, 2003).

),

(

),

Fulgoromorpha species damage plants by egg-laying and passing eggs into plant tissues, when fed with phloem cell juice, as well as in the transmission of various plant pathogens (Kastal'eva et al., 2016).

According to the authors, Cixiidae species were carriers of phytoplasma infections, while Delphacidae species are carriers of viruses primarily. These insect pests cause serious damage to crops and infectious diseases.

Cicadas are insect pests that can cause serious damage in the cultivation of crops, including alfalfa.

However, in most of Western and Central Europe, the damage to crops is limited, and the cicadas role in the functioning of the ecosystem is direct and indirect because they feed on the plant juice from xylem and phloem and are the transmission of pathogenic microorganisms (Nickel, 2003).

However, their importance and presence should not be underestimated, as insects have to consume large amounts of vegetable juice to receive the necessary nutrients.

Direct damage associated with feeding, whereby cicadas suck tissue juices (which slows down the growth and development of the plant organism) as well as associated with egg-laying, which causes mechanical damage.

Common symptoms of damage often resemble the aging process, and they appear after eating, regardless of species identification. These symptoms include wilting in young plants, leaf chlorosis (yellowing or browning of the leaves), followed by premature leaf fall, and inhibiting plant growth caused by the reduction of stem elongation (Backus et

<p>(Backus et al., 2005).</p> <p>-</p> <p>,</p>	<p>al., 2005). Although the leaf chlorosis is the more visible and quickly establishing feature, the most important one for reduced yield is suppressed vegetative and reproductive growth and development.</p>
<p>(2005)</p> <p>. Backus</p> <p>,</p> <p>,</p>	<p>Backus et al. (2005) found that alfalfa development during the growing season was of particular importance in the formation of forage production such as hindering or stopping the growth directly led to a proportional decrease in yield.</p>
<p>.</p> <p>,</p> <p>,</p>	<p>- The damage to the cicada feeding is related not only to a decrease in the forage production but also to a change in the forage quality.</p>
<p>,</p> <p><i>Empoasca</i></p> <p>,</p> <p>(Rethwisch, 2000).</p>	<p>- Recent studies involving the influence of <i>Empoasca</i> cicadas on forage quality showed a decrease in digestible protein concentration with increasing cicada density (Rethwisch, 2000).</p>
<p>Cicadellidae (Weintraub and Beanland, 2006).</p> <p>,</p>	<p>- One of the main vectors for spreading the Stolbur phytoplasma disease are the Cicadellidae species (Weintraub and Beanland, 2006).</p>
<p>(Alhudaib et al., 2007).</p> <p>,</p>	<p>- They can cause substantial plant damage and productivity losses in a wide variety of crops, including alfalfa (Alhudaib et al., 2007). The most common characteristic symptom of this disease is reducing leaf mass and reproductive organs.</p>
<p>50%,</p> <p>(Girsova et al., 2017).</p> <p>Mazzoni (2005)</p>	<p>- The vegetative organs and seeds weight decreases by more than 50%, and depending on the damage degree, flower buds and seeds may not form (Girsova et al., 2017).</p>
<p>,</p> <p>,</p> <p>,</p>	<p>- Mazzoni (2005) examined the species diversity of cicadas in the Tuscany region, Italy, depending on plant hosts and observed the high prevalence of monophagous and polyphagous species. The author found that in alfalfa mainly dominated by 9 species, in</p>

9  
*Empoasca decipiens* Paoli,  
*Euscelis lineolata*, *Psammotettix alienus*  
 Dahlbom, *Aphrodes makarovi* Zachvatkin,  
*Austroagallia sinuata* Mulsant & Rey,  
*Empoasca alsiosa* Ribaut, *Reptalus*  
*quinguecostatus* Dufour

*Macrosteles laevis*, *Psammotettix*  
*alienus*, *Empoasca pteridis*, *Javesella*  
*pellucida* Fabricius *Laodelphax striatellus*  
 Fallen

(Malenovský and Auterer, 2002).  
 (Nickel, 2003; Nickel and  
 Hildebrandt, 2003),

*Empoasca fabae*

(Sulc and Lamp, 2007).

220 26 (62%)  
 Fabaceae (Lamp  
 et al., 1994).

*Empoasca*

(2008)  
*Empoasca decipiens* Paoli

*M. sativa*.

(2005) 4  
 (*Cercopis sanguinolenta* Scopoli,

particular *Empoasca decipiens* Paoli,  
*Euscelis lineolata*, *Psammotettix alienus*  
 Dahlbom, *Aphrodes makarovi* Zachvatkin,  
*Austroagallia sinuata* Mulsant & Rey,  
*Empoasca alsiosa* Ribaut, *Reptalus*  
*quinguecostatus* Dufour and others. Also,  
 he concluded that the ecology of the  
 species depending on various factors  
 such as the stage of development,  
 climate, season, and the host plant  
 presence.

*Macrosteles laevis*, *Psammotettix*  
*alienus*, *Empoasca pteridis*, *Javesella*  
*pellucida* Fabricius and *Laodelphax*  
*striatellus* Fallen were dominant cicada  
 species in arable fields, and mainly alfalfa,  
 in the Czech Republic (Malenovský and  
 Auterer, 2002). According to some  
 authors (Nickel, 2003; Nickel and  
 Hildebrandt, 2003), those species easily  
 migrated to agroecosystems with disturbed  
 habitats in the early successional stages.

*Empoasca fabae* is one of the most  
 serious economic pests affecting forage  
 production of alfalfa in the Midwest and  
 the United States and the species  
 reached the economic threshold of harm  
 every year (in summer) (Sulc and Lamp,  
 2007). Cicada has a rich and diverse  
 range of hosts, including 220 species of  
 plants from 26 families, and the majority  
 of plants (62%) belong to the Fabaceae  
 family (Lamp et al., 1994).

*Empoasca* genus is the most  
 common and reported alfalfa pest  
 worldwide. In a study conducted in  
 different regions of Egypt, Shebl et al.  
 (2008) found that *Empoasca decipiens*  
 Paoli was the only major species with  
 economic importance among cicadas in  
*M. sativa*.

In Albania, Kullaj et al (2005) studied the  
 alfalfa entomofauna and reported 4 cicada  
 species (*Cercopis sanguinolenta* Scopoli,  
*Ceresa bubalus* Fabricius, *Cicadella*

*Ceresa bubalus* Fabricius, *Cicadella viridis* Linnaeus, *Empoasca* spp., *Philaenus spumarius* Linnaeus),

-  
-  
*Empoasca*.  
*E. pteridis*

,  
*Lepyronia coleoptrata*  
Linné *Euscelis plebeia* Fallén,  
- *Sonronius binotata*  
Sahlberg *Philaenus spumarius* (Girsova et al., 2015).

, *E. pteridis*

(Bogoutdinov et al., 2008; Bogoutdinov, 2012; Bakunov and Dmitrieva, 2015; Br án, 2012).

*Cicadella viridis* (Cicadellidae)

-  
-  
( Duduk et al., 2008; Ando et al., 2010; Janse and Obradovic, 2010; Trivellone et al., 2012).

. *viridis*,

*Macrosteles laevis* Ribaut,  
Cicadellidae,

(Remane, 2005).

*Macrosteles*,

(Nickel, 2003).

,  
3  
(Tóthová et al., 2004, Olivier et al., 2009, Girsova et al., 2015).

*Euscelis plebejus* Fallén

*viridis* Linnaeus, *Empoasca* spp., *Philaenus spumarius* Linnaeus). They found that the most numerous were *Empoasca* genus. In the Moscow region, *E. pteridis* predominated in the second half of the alfalfa growing season, together with *Lepyronia coleoptrata* Linné and *Euscelis plebeia* Fallén, and in the first half of the season- *Sonronius binotata* Sahlberg and *Philaenus spumarius* (Girsova et al., 2015).

A number of authors reported that *E. pteridis* is a carrier of Stolbur phytoplasma in alfalfa (Bogutdinov et al., 2008; Bogutdinov, 2012; Bakunov and Dmitrieva, 2015; Br án, 2012).

*Cicadella viridis* (Cicadellidae) is one of important insect pests and carrier of pathogens in economically important crops (corn, vineyards, carrots, some fruit, alfalfa) (Duduk et al., 2008; Ando et al., 2010; Janse and Obradovic, 2010; Trivellone et al., 2012).

The species is polyvoltine whose development goes through five larval stages. There is a growing interest in the study of the microbial community associated with *C. viridis* as with other insect vectors to identify micro-organisms that can be used as alternative control strategies.

*Macrosteles laevis* Ribaut, a representative of the Cicadellidae, occurred in high numbers in modern agroecosystems in Europe and was one of the main species in the new pastures (Remane, 2005). The cicada, like other *Macrosteles* species, preferred and inhabited disturbed habitats as well as fertilized pastures (Nickel, 2003). *Macrosteles laevis* is a polyphagous and carrier of stolbur and develops three generations per year (Tóthová et al., 2004, Olivier et al., 2009, Girsova et al., 2015).

*Euscelis plebejus* Fallén is another common species in alfalfa, and the

(Girsova et al., 2015).  
*Psammotettix striatus* Linnaeus  
 (Kastalyeva  
 et al., 2016).  
 22  
 10  
*Medicago sativa*  
*P. striatus*,  
*Paraphlepsius apertinus* Osborn &  
 Lathrop (Nielson, 1968) *Paraphlepsius*  
*(Sabix) irroratus* Say (Chiykowski, 1985),  
*Austroagallia sinuata* Mulsant &  
 Rey  
 (Nickel,  
 2003; Mazzoni, 2005).  
 Aphrophoridae,  
*Philaenus spumarius* Linnaeus  
 -  
 (Yurtsever,  
 2000).  
 (

biology of the species is well known (Girsova et al., 2015).

*Psammotettix striatus* Linnaeus is a major pest of wheat and other cereal crops such as barley, oats, and alfalfa at the same time (Castaleva et al., 2016). As a result of a nine-year study, the authors reported that 22 plant species from 10 families were hosts of the species among them *Medicago sativa* is one of the main hosts. *Psammotettix striatus* is a carrier of stolbur on the plants too.

*Paraphlepsius apertinus* Osborn & Lathrop (Nielson, 1968) and *Paraphlepsius (Sabix) irroratus* Say (Chiykowski, 1985) also were reported as important harmful species among alfalfa cicadas in studies of the last century (Chiykowski, 1985). The latter cicada was a common species in clover and various grass and weed species.

*Austroagallia sinuata* Mulsant & Rey is a species found in various hosts, including alfalfa in Germany, Italy, Turkey (Nickel, 2003; Mazzoni, 2005).

*Philaenus spumarius* Linnaeus (Aphrophoridae) is one of the most frequently occurring species in legumes. As a result of interesting aspects of its biological development, for decades the species had received much attention from biologists.

The species was one of the most widely studied species in the ecology and genetics area (Yurtsever, 2000). Cicada had a cosmopolitan distribution, a wide variety of habitats and strong sexual dimorphism (depending on different geographical areas). *Philaenus spumarius* sucked the plant juice from the xylem, and the nymphs, through salivary separation, formed a foamy mixture on the leaves. The foamy mixture surrounded their delicate and soft body and provided some protection from drying and predator attack (Ossiannilsson, 1978). The species was characterized by hundreds of hosts

<p>(Yurtsever, 1999).</p> <p>(<i>Medicago sativa</i>, <i>Trifolium</i> spp., <i>Vicia</i> spp., <i>Xanthium strumarium</i>) (Byers et al., 2001).</p> <p><i>Philaenus spumarius</i></p> <p>(Yurtsever, 1999).</p> <p><i>Hyalesthes obsoletus</i> Signoret (Cixiidae)</p> <p>(Milanesi et al., 2005; Riolo et al., 2012).</p> <p><i>Hyalesthes obsoletus</i></p> <p>(B ezíková and Linhartová, 2007; Weintraub, 2010).</p> <p><i>Reptalus panzeri</i> Low (Cixiidae)</p> <p><i>H. obsoletus</i>,</p> <p>(Weintraub, 2010).</p> <p>(2009)</p> <p>Membracidae Rafinesque (Hemiptera: Cicadomorpha) 3500 (Dietrich, 2008; Deitz et al., 2011).</p>	<p><i>P. spumarius</i></p> <p>- ranging from grasses to tree species in Europe, including meadow plants, herbs, and shrubs due to its great flexibility and adaptability (Yurtsever, 1999).</p> <p>- The nitrogen-fixing herbaceous legumes and some others that contain high concentrations of amino acids were the most preferred by cicadas (<i>Medicago sativa</i>, <i>Trifolium</i> spp., <i>Vicia</i> spp., <i>Xanthium strumarium</i>) (Byers et al., 2001).</p> <p>- <i>Philaenus spumarius</i> was a multivoltine species, and in some parts of Greece, the British Isles and some areas in Turkey, the species exhibited as a bivoltine (Yurtsever, 1999).</p> <p>- <i>Hyalesthes obsoletus</i> Signoret (Cixiidae) is a widespread polyphagous that prefer herbaceous hosts, including alfalfa (Milanesi et al., 2005; Riolo et al., 2012). The species wintered in a larval stage and only perennial plant species served as hosts although adult individuals can be found on a wide range of additional crops as food sources.</p> <p>- The species was a carrier of stolbur and once infected, remained a carrier of phytoplasma infections for the rest of its life (B ezíková and Linhartová, 2007; Weintraub, 2010).</p> <p>- <i>Reptalus panzeri</i> Low (Cixiidae) was also polyphagous, which appeared at the same time with <i>H. obsoletus</i> but was not as widespread (Weintraub, 2010). In Serbia, Jovi et al. (2009) found that <i>R. panzeri</i> may be a vector of phytoplasma infections causing stolbur.</p> <p>- Membracidae Rafinesque (Hemiptera: Cicadomorpha) is a large family and includes some 3500 known species in the world (Dietrich, 2008; Deitz et al., 2011). The species from that family are interesting not only for their strange shapes but also for their unusual behavior. Many of them form large and</p>
--	---



al., 2011).

Meneguzzi (2009) reported that *Ceresa nigripectus* Remes Lenicov (Membracidae)

*Ceresa bubalus* Fabricius

1-3

(wierczewski and Stroi ski, 2011).

Fulgoromorpha Cicadomorpha

(Holzinger et al., 2003; Nickel, 2003).

(Mazzoni, 2005).

Yosifov (1962)

20 : *Empoasca pteridis*, *Laodelphax striatellus* (*Psammotettix striatus*), *Macrosteles laevis* Ribaut, *Aphrodes costatus* Panzer, *Lepironia coleoptrat* Linnaeus, *Limotettix* (*Scleroracus*) *corniculus* Marshall, *Hardya*

often noticeable clusters of larvae or adults. In addition, some species exhibit mutualism together with social ants, which associations may be beneficial to plants (Deitz et al., 2011). In general, the species are not considered to be serious agricultural or forestry insect pests, although some of them mechanically damage the plant stems during the oviposition.

Meneguzzi (2009) reported that *Ceresa nigripectus* Remes Lenicov (Membracidae) was one of the most common alfalfa cicadas in Argentina and the species is also a carrier of phytoplasma infections.

*Ceresa bubalus* Fabricius was another common member of that family, preferring to lay their eggs on 1-3-year-old branches of deciduous trees and shrubs. Crops such as alfalfa and red clover served as the best source of food for the larvae (wierczewski and Stroi ski, 2011).

The awareness for species from the Fulgoromorpha and Cicadomorpha suborder in alfalfa through a faunal ecological study in Central Europe has been considerably improved in recent decades thanks to advances in knowledge of both their taxonomy and biology (Holzinger et al., 2003; Nickel, 2003). The knowledge provided additional information about the relationship between species and their typical habitat (Mazzoni, 2005).

### STUDIES IN BULGARIA

In Bulgaria, the first information on the species composition of cicadas in alfalfa was reported by Yosifov (1962). He found 20 species: *Empoasca pteridis*, *Laodelphax striatellus* (*Psammotettix striatus*), *Macrosteles laevis* Ribaut, *Aphrodes costatus* Panzer, *Lepironia coleoptrat* Linnaeus, *Limotettix* (*Scleroracus*) *corniculus* Marshall, *Hardya*

(Hardya) *tenuis* Germar, *Caligypona minuscula* Horvath, *Selenocephalus obsoletus* Germar, *Hysteropterum grylloides* F., *Psammotettix confinis* Dahlb., *Tettigometra reticulates* Pnz., *Euscelis plebeia* Fallén, *siraca clavicornis* Fabricius, *Dryodukgades reticulates* Sign, *Balclutha punctata* Fabricius, *Balclutha saltuella* Kirschbaum, *Hyalesthes obsoletus* Signoret, *Cicadella viridis* Linnaeus, *Dikraneura mollicula* Boh.

*Empoasca pteridis* *Psammotettix striatus*.

- , Bajryamova (1976, 1982)

*Empoasca pteridis*, *Macrosteles laevis*, *Euscelis plebeia* Fallén, *Aphrodes bicinctus* Schrank, *Cicadella viridis* Linnaeus, *Philaenus spumarius* Linnaeus, *Psammotettix alienus* Dahlbom, *P. provincialis* Ribaut. Popova (1968)

18

: *Cicadella viridis* Linnaeus, *Lepironia coleoptrat* Linnaeus, *Aphrodes bicinctus* Schrank, *Reptalus quinguecostatus* Dufour, *Ceresa bubalus* Fabricius, *Asiraca flavicornis* F., *Philaenus spumarius* Linnaeus, *Cixium desertorum* Fibr., *Macrosteles laevis* Ribaut, *Scleroracrus decumanus* Kontk., *Empoasca pteridis*, *Deltocephalus* sp., *Anacerataquallia* sp., *Artianus* sp., *Philaenus spumarius* Linnaeus, *Cercopis sanguinolenta* Scopoli, *Macrosteles* sp., *Macrosteles quadripunctata* Kbn., *Aphrodes fuscofascitus* Gn., *Selenocephalus obsoletus* Germar.

E.

*pteridis*

Atanasova (2011),

*Cicadella viridis*, *Empoasca pteridis*, *Philaenus spumarius* *Cercopis vulnerata* Rossi (Cercopidae).

*M. sativa*

-

(Hardya) *tenuis* Germar, *Caligypona minuscula* Horvath, *Selenocephalus obsoletus* Germar, *Hysteropterum grylloides* F., *Psammotettix confinis* Dahlb., *Tettigometra reticulates* Pnz., *Euscelis plebeia* Fallén, *siraca clavicornis* Fabricius, *Dryodukgades reticulates* Sign, *Balclutha punctata* Fabricius, *Balclutha saltuella* Kirschbaum, *Hyalesthes obsoletus* Signoret, *Cicadella viridis* Linnaeus, *Dikraneura mollicula* Boh.

Among them, *Empoasca pteridis* and *Psammotettix striatus* were in the highest numbers.

Later, Bayryamova (1976, 1982) reported as characteristic cicada pests in alfalfa *Empoasca pteridis*, *Macrosteles laevis*, *Euscelis plebeia* Fallén, *Aphrodes bicinctus* Schrank, *Cicadella viridis* Linnaeus, *Philaenus spumarius* Linnaeus, *Psammotettix alienus* Dahlbom, *P. provincialis* Ribaut. Popova (1968) identified 18 species of cicadas in alfalfa: *Cicadella viridis* Linnaeus, *Lepironia coleoptrat* Linnaeus, *Aphrodes bicinctus* Schrank, *Reptalus quinguecostatus* Dufour, *Ceresa bubalus* Fabricius, *Asiraca flavicornis* F., *Philaenus spumarius* Linnaeus, *Cixium desertorum* Fibr., *Macrosteles laevis* Ribaut, *Scleroracrus decumanus* Kontk., *Empoasca pteridis*, *Deltocephalus* sp., *Anacerataquallia* sp., *Artianus* sp., *Philaenus spumarius* Linnaeus, *Cercopis sanguinolenta* Scopoli, *Macrosteles* sp., *Macrosteles quadripunctata* Kbn., *Aphrodes fuscofascitus* Gn., *Selenocephalus obsoletus* Germar.

The author reported *E. pteridis* as dominant species and presented the population dynamics of cicadas in alfalfa. Atanasova (2011), studying the harmful entomofauna of Multifoliate and trifoliate alfalfa varieties, reported following cicadas: *Cicadella viridis*, *Empoasca pteridis*, *Philaenus spumarius* and *Cercopis vulnerata* Rossi (Cercopidae)

In Bulgaria, information on that group of cicadas in *M. sativa* is scarce. Most cicadas studies focused on cereals

(Karadjova and Krusteva, 2017; Karadjova and Krusteva, 2016).	-	(Karadjova and Krusteva, 2016; Karadjova and Krusteva, 2017). Attention to the problem of cicadas was also given to some other crops, such as vegetable and annual legumes (Bogatsevskaja et al., 2008; Zhekova, 2012).
(Bogatsevskaja et al., 2008; Zhekova, 2012).	-	The need for a more in-depth study of alfalfa cicadas is the result of insufficient information on these species.
Fulgoromorpha (Hemiptera)	Cicadomorpha	Until now, studies have been conducted on the Fulgoromorpha and Cicadomorpha (Hemiptera) suborder species in different geographical regions of the country.
	-	Determination of the species composition and population density of alfalfa cicadas will help to find rational solutions for controlling their numbers and setting the term for plant protection.

## / REFERENCES

1. **Alhudaib, K., Y. Arocha, M. Wilson and P. Jones**, 2007. First Report of A 16Srl, Candidatus Phytoplasma Asteris Group Phytoplasma Associated with a Date Palm Disease In Saudi Arabia. *Plant Pathology* NDR 15, Feb. 2007.
2. **Ando, Y., S. Utsumi and T. Ohgushi**, 2010. Community Structure of Insect Herbivores on Introduced and Native Solidago Plants in Japan. *Entomologia Experimentalis et Applicata*, 136, 174-183.
3. **Atanasova, D.**, 2011. A Study of the Insect Pests and Their Entomophages on Multifoliate Alfalfa (Medicago Sativa L.). PhD thesis, Agrarian University, Plovdiv, pp.1-35 (Bg).
4. **Backus, E.A., M.S. Serrano and C.M. Ranger**, 2005. Mechanisms of Hopperburn: An Overview of Insect Taxonomy, Behavior, and Physiology. *Annual Review of Entomology*, 50, 125-51.
5. **Bajryamova, V.**, 1976. Cicadas (Homoptera. Auchenorrhyncha) on Cultivated Plants in Sofia Field. Terrestrial fauna of Bulgaria. Materials, Sofia, 121-141 (Bg).
6. **Bajryamova, V.**, 1982. Study of the Number Dynamics of the Cicadas Entomocenosis in Sofia Field Crops. *Ecology*, 10, 13-21 (Bg).
7. **Bakunov, A.L. and N.N. Dmitrieva**, 2015. Stolbur - A Harmful Disease of Potatoes. *Bulletin of the Samara Scientific Center of the Russian Academy of Sciences*, 17, 4 (3), 570-572 (Ru).
8. **Bogatsevskaja, N., D. Hristova, S. Simova, E. Staneva, R. Nakov, Ts. Dimitrova, I. Kiryakov and P. Grigorova**, 2008. A Guide to Integrated Pest Management in Cereals. Sofia (Bg).
9. **Bogoutdinov, D.Z.**, 2012. The Role of Cicadas in Agrocenoses of Solanaceae Crops. *Brief communications*, UDC 632.753.1, 45-46.
10. **Bogoutdinov, D.Z., D. Valyunas, M. Navalinskens, M. Samuytene**, 2008. On

The Species Identification of Pathogens of Phytoplasmas of Solanaceae Crops. *Agricultural Biology*, 1, 77-80 (Ru).

11. **Brán, J.**, 2012. Leafhopper And Planthopper Vectors of Plant Disease Agents in Central and Southern Europe. In: Karl Maramorosch and H. Karris. Leafhopper Vectors and Plant Disease Agents. Elsevier, at: [https://books.google.bg/books?id=ko0UM0-nz4C&dq=Empoasca+pteridis&hl=bg&source=gbs\\_navlinks\\_s](https://books.google.bg/books?id=ko0UM0-nz4C&dq=Empoasca+pteridis&hl=bg&source=gbs_navlinks_s)
12. **Bezíková, M. and S. Linhartová**, 2007. First Report of Potato Stolbur Phytoplasma in hemipterans in Southern Moravia. *Plant Protection Science*, 43, 73-76.
13. **Byers, R.A., C.C. Bahler, W.L. Stout, K.T. Leath and L.D. Hoffman**, 2001. The Establishment of Alfalfa into Different Maize Residues by Conservation-Tillage and Its Effect on Insect Infestation. *Grass and Forage Science*, 54(1), 77-86.
14. **Cryan, J.R. and J.M. Urban**, 2012. Higher-Level Phylogeny of the Insect Order Hemiptera: Is Auchenorrhyncha Really Paraphyletic? *Systematic Entomology*, 37 (1), 7-21.
15. **Deitz, L., M. Wallace, C. Dietrich, S. Mckamey and M. Rothschild**, 2011. Treehoppers. (<http://treehoppers.insectmuseum.org/public/site/treehoppers/home>),
16. **Dietrich, C.**, 2008. Treehoppers. [http:// www.inhs.uiuc.edu/~dietrich/treehome.html](http://www.inhs.uiuc.edu/~dietrich/treehome.html).
17. **Duduk, B., P. Peri , D. Mar i , T. Drobnjakovi , L. Picciau, A. Alma and A. Bertaccini**, 2008. Phytoplasmas in Carrots: Disease and Potential Vectors in Serbia. *Bulletin of Insectology*, 61 (2), 327-331.
18. **ngel, M.S. and D.A. Grimaldi**, 2004. New Light Shed on the Oldest Insect. *Nature*, 427, 627-630.
19. **Girsova, H.B., T.B. Kastalyeva, U.I. Sacks, K.A. Mozhaev and D.Z. Bogoutdino**, 2015. Legumes Plants Phytoplasmas. Proceedings of the TSHA, 2, 58-73 (Ru).
20. **Girsova, V., D.K. Bottner-Parker, D.Z. Bogoutdinov, T.B. Kastalyeva, Y.I. Meshkov, K.A. Mozhaeva and I-M. Lee**, 2017. Diverse Phytoplasmas Associated With Leguminous Crops In Russia. *European Journal of Plant Pathology*, DOI 10.1007/s10658-017-1209-6.
21. **Hall, M.L.**, 2009. A Review of Vocal Duetting in Birds. *Advances in the Study of Behavior*, 40, 67-121.
22. **Holzinger, W.E., I. Kammerlander and H. Nickel**, 2003. The Auchenorrhyncha of Central Europe: Die Zikaden Mitteleuropas. Brill, Leiden – Boston, 673 pp.
23. **Holzinger, W.E., I. Kammerlander and H. Nickel**, 2003. The Auchenorrhyncha of Central Europe: Die Zikaden Mitteleuropas. Brill, Leiden – Boston, 673 pp.
24. **Janse, J.D. and A. Obradovic**, 2010. Xylella fastidiosa: Its biology, Diagnosis, Control and risks. *Journal of Plant Pathology*, 92 (S1), 35-48.
25. **Jovi , J., T. Cvrkovi , M. Mitrovi , S. Krnjaji , A. Petrovi , M.G. Redinbaugh, C. Pratt, S.A. Hogenhout and I. Toševski**, 2009. Stolbur Phytoplasma Transmission to Maize by Reptalus Panzeri and the Disease Cycle of Maize Redness in Serbia. *Phytopathology*, 99, 1053-1061.
26. **Karadjova, O. and H. Krusteva**, 2016. Species Composition and Population Dynamics of the Harmful Insect Fauna (Hemiptera: Cicadomorpha, Fulgoromorpha and Sternorrhyncha) of Winter Triticale. *Bulgarian Journal of Aricultural Science*, 22, 619-626.
27. **Karadjova, O. and H. Krusteva**, 2017. Scientific Opinion on the Spread of Cicadas (Hemiptera: Cocadomorpha And Fulgoromorpha) in Cereals and Sunflower Crops in Bulgaria and the Potential Vectors Species of Xylella Fastidiosa, A Regulated Pathogen in The EU (Bg).
28. **Kastal'eva, T.B., D.Z. Bogoutdinov, K.D. Botiner-Parker, N.V. Girsova and IM.**

- Lee, 2016. Diverse Phytoplasmas Associated with Diseases in Various Crops in Russia – Pathogens and Vectors. *Agricultural Biology*, 51(3), 367-375.
29. **Kastalyeva, T.B., D.Z. Bogoutdinov, C.D. Bottnerparker, H.B. Girsova and I.M.O. Lee**, 2016. Diversity of Phytoplasmas in Agricultural Crops in Russia: Pathogens and Their Carriers. *Agricultural Biology*, 3, 367-375 (Ru).
30. **Kullaj, E., A. Çakalli, S. Shahini and S. Varaku**, 2005. Entomofaunistic Study on Alfalfa, A Precondition to The Biological Control of Its Pests. Conference: 1st Congress of Plant Protection “Environmental Concern and Food Safety” and XXXth Meeting for Plant Protection in the Republic of Macedonia.
31. **Lamp, W., G. Nielson, and S. Danielson**, 1994. Patterns among Host Plants of Potato Leafhopper, *Empoasca fabae* (Homoptera: Cicadellidae). *Journal of the Kansas Entomological Society*, 67(4), 354-368.
32. **Malenovský, I. and P. Auterer**, 2002. Species Assemblages of Auchenorrhyncha and Psylloidea (Hemiptera) in red-clover fields in the Brno environs. Pp. 69-70. In: B RYJA J. & Z UKAL J.: Zoologické dny Brno 2002. Abstrakta referátů z konference, 14.-15. února 2002. Ústav biologie obratlovců AVÈR, Brno, 191 pp. (CZ).
33. **Mazzoni, V.**, 2005. Contribution to the Knowledge of the Auchenorrhyncha (Hemiptera Fulgoromorpha and Cicadomorpha) of Tuscany (Italy). *REDIA*, LXXXVIII, 85-102.
34. **Meneguzzi, N.**, 2009. Caracterización Molecular, Taxonomía Y Diagnóstico De Fitoplasmas Del Grupo Ash Yellows (VII). Tesis de Doctorado en Ciencias Biológicas. Fac. de Cs. Exactas, Físicas y Naturales. Univ. Nac. de Córdoba. 146 pp.
35. **Mifsud, D., C. Cocquempot, R. Mühlethaler, MR. Wilson and J.-C. Streito**, 2010. Other Hemiptera Sternorrhyncha (Aleyrodidae, Phylloxeroidea, and Psylloidea) and Hemiptera Auchenorrhyncha. Chapter 9.4, BioRisk – Biodiversity and Ecosystem Risk Assessment – Biodiversity and Ecosystem Risk Assessment, 4 (1), 511–552.
36. **Milanesi, L., R. Bondavalli N. Mori, D. Dradi, I. Menozzi and A. Bertaccini**, 2005. Remarks On the phytoplasma vector of black wood of the vine, *Hyalesthes obsoletus*, in Emilia Romagna. *Petria*, 15, 59-61 (I).
37. **Nickel, H. and J. Hildebrandt**, 2003. Auchenorrhyncha Communities as Indicators Of Disturbance In Grasslands (Insecta, Hemiptera) - A Case Study From The Elbe Flood Plains (Northern Germany). *Agriculture, Ecosystems and Environment*, 98, 183-199.
38. **Nickel, H.**, 2003. The Leafhoppers and Planthoppers of Germany (Hemiptera, Uchenorrhyncha): Patterns and Strategies in a Highly Diverse Group of Phytophagous Insects. A. Pensoft publishers, 460 pp.
39. **Nielson, M.**, 1968. The Leafhopper Vectors Of Phytopathogenic Viruses (Homoptera: Cicadellidae), Taxonomy, Biology, and Virus Transmission. *Technical Bulletin 1382*, Washington, D. C., 233.
40. **Olivier, C.Y., D.T. Lowery and LW. Stobbs**, 2009. Phytoplasma Diseases and Their Relationships with Insect and Plant Hosts in Canadian Horticultural and Field Crops. *Canadian Entomologist*, 14, 425-462.
41. **Ossiannilsson, F.**, 1978. Fauna Entomologica Scandinavica: The Auchenorrhyncha (Homoptera) of Fennoscandia and Denmark. *Scandinavian Science Press, Klampenborg*, 7, 1-222.
42. **Popova, V.**, 1968. Entomofauna on Alfalfa. Academy of Agricultural Sciences, IPP, Kostinbrod, Publishing House of the Bulgarian Academy of Sciences, Sofia, 5-151 (Bg).
43. **Remane, R.**, 2005. The Cicada Fauna of Iceland, a Geologically "Old" Volcanic Island: Analysis of a 2001 by Dr. med. M. v. Tschirnhaus Collected Material (Insecta Rhyncota Auchenorrhyncha). *Marburger entomologische Publikationen*, 3(3), 111-123 (D).

44. **Rethwisch, M.D.**, 2000. Comparisons of Differing Rates of Baythroid® 2 And WarriorT® Insecticides For Insect Control In Fall Alfalfa. Pp. 21-30. In University of Arizona College of Agriculture 2000 Forage and Grain Report, Series P-124. M. Ottman, ed. 152 pp.
45. **Riolo, P., R.L. Minuz, G. Anfora, MV. Rossi Stacconi, S. Carlin, N. Isidoro and R. Romani**, 2012. Perception of Host Plant Volatiles in *Hyalesthes Obsoletus*: Behavior, Morphology, and Electrophysiology. *Journal of Chemical Ecology*, DOI 10.1007/s10886-012-0154-2.
46. **Ritzmann, R.E., R.D. Quinn and M.S. Fischer**, 2004. Convergent Evolution and Locomotion through Complex Terrain By Insects, Vertebrates and Robots. *Arthropod Structure & Development*, 33, 361-379.
47. **Shebl, M.A., S.M. Kamel, T.A. Abu Hashesh and M.A. Osman**, 2008. The Most Common Insect Species in Alfalfa Field in Egypt. *Academic Journal of Entomology*, 1(2), 27-31.
48. **Sulc, M. and W.O. Lamp**, 2007. Insect Pest Management. In Forages: The Science of Grassland Agriculture, R.F. Barnes, D.A. Miller, and C.J. Nelson (eds.), 6th ed., Vol. II, pp. 411-424. Iowa State Univ. Press.
49. **wierczewski, D. and A. Stroi ski**, 2011. The First Records of the Nearctic Treehopper Stictoccephala Bisionia in Poland (Cicadomorpha: Membracidae) with Some Comments on This Potential Pest. *Polish Journal of Entomology*, 80, 13-22.
50. **Tóthová, M., P. Tóth and . Cagá** , 2004. Leafhoppers, Planthoppers, Froghoppers and Cixiids (Auchenorrhyncha) on Pigweeds As Vectors of Plant Diseases. *Acta fytotechnica et zootechnica*, 7, Special Number, Proceedings of the XVI. Slovak and Czech. Plant Protection Conference organised at Slovak Agricultural University in Nitra, Slovakia, pp. 322-326.
51. **Trivellone, V., L.P. Paltrinieri, M. Jermini and M. Moretti**, 2012. Management Pressure Drives Leafhopper Communities in Vineyards In Southern Switzerland. *Insect Conservation and Diversity*, 5, 75-85 (D).
52. **Weintraub, P. and L. Beanland**, 2006. Insect Vectors of Phytoplasmas. *Annual Review of Entomology*, 51, 91-111.
53. **Weintraub, P.G.**, 2010. Auchenorrhyncha Phytoplasma Vectors. In: M.Maixner, P. Weintraub B. Jarausch. COST Action FA0807 - Working group 2 Grapevine Yellows Vector Sampling and Monitoring Training School, pp. 7-17.
54. **Yosifov, M.**, 1962. Quantitative and Qualitative Studies of the Entomofauna of Alfalfa Fields in Sofia with a View to Insects of the Heteroptera Order. *Notifications of the Zoological Institute and Museum, BAS, Sofia*, XI, 111-140 (Bg).
55. **Yurtsever, S.**, 1999. Inheritance of Three Dorsal Colour/Pattern Morphs in Some Turkish *Philaenus Spumarius* (Homoptera: Cercopidae) Populations. *Israel Journal of Zoology*, 45, 361-369.
56. **Yurtsever, S.**, 2000. On the Polymorphic Meadow Spittlebug, *Philaenus Spumarius* (L.) (Homoptera: Cercopidae). *Turkish Journal of Zoology*, 24, 447-459.
57. **Zhekova, E.D.**, 2012. Monitoring of the Entomofauna in Ecological Crop Rotation. *Banat's Journal of Biotechnology*, III (2), 104-110.