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Study on grain yield in spring vetch variety "Tempo" depending on the technology of cultivation

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SUMMARY

The aim of the experiment is to determine grain yield in spring vetch variety "Tempo" depending on the technology of cultivation. The study was conducted in the period 2011-2013, under field conditions, on soil subtype slightly-leached chernozem. Variants of cultivation: 1. At conventional technology – control; 2. Without use of preparations of inorganic origin; 3. Treatment with bio insecticides "Ecofil P". It was found that the best biometric and structural parameters of the extraction, as well as higher yields of grain (221,00 kg/da biological and 165,50 kg/da economic) distinguishes the crop cultivated in the conventional technology. The lowest yields of grain (biological 122,49 kg/da and economic 98,90 kg/da) were harvested from the crop grown without the use of preparations of inorganic origin. Yields derived from the crop treated with bio preparation "Ecofil P" occupy an intermediate position.

Keywords: spring vetch, structural elements, bio preparation, technology, grain yield

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|-----------------|------|--------------|---|
| “ ” | - | 2011-2013 .. | |
| : | 1. | | |
| - | ; 2. | | |
| | | ; 3. | |
| ” | ” | , | - |
| (221,00 kg/da) | | 165,50 kg/da | |
| | | - | |
| | | (| |
| 122,49 kg/da | | 98,90 kg/da) | |
| | | ” | |
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INTRODUCTION

(*Vicia sativa* L.)

2010). (Kertikov,

et al., 1997; Jimenez, 1999).

Kertikov, 2003),
(Kertikov, 2000),

al., 1994).

, (Katayama,
1991; Kertikov and Vasileva 2000; Vasileva
and Kertikov, 2007).

— (Kertikov, 2005; Kertikova et
al., 2012).

2011-2013

Common vetch (*Vicia sativa* L.) is well known in our country forage legumes. Although less disseminated by the other legumes, it is important to solve protein problem in the country (Kertikov, 2010). In recent years, researches in this culture become particularly topical.

Basically they are agro-technical orientation. It is the influence of certain biotic and abiotic factors on seed yield, yields of grain and feed at separate and mixed crops (Mousa et al., 1997; Jimenez, 1999). An important share occupy studies determining the quantitative and qualitative parameters of yield (Orak, 2000; Kertikov, 2003), seed and fertilizer rates (Kertikov, 2000), the nitrogen-fixing ability of liquid chromatography have its effect on the soil nutritive regime (Ynkov et al., 1994). Thorough evaluation of the use of biologically active substances to control growth, development and increased yield of spring vetch was conducted (Katayama, 1991; Kertikov and Vasileva, 2000; Vasileva and Kertikov, 2007). As a result of comprehensive studies on the cultivation and use of new varieties and intensive breeding-improvement work created a new variety of spring vetch – Tempo (Kertikov, 2005; Kertikova et al., 2012).

The aim of the experiment was to determine effect of different technologies for cultivation of spring vetch cv. Tempo on the level of lodging, yield, structural elements in yield and grain losses.

MATERIAL AND METHODS

The experiment was conducted during the period 2011-2013 in the second experimental field in Institute of Forage Crops with spring vetch cv. Tempo.

The variety is a good productivity,

(Kertikova et al., 2012).

10 m².
 1 (Kostov and Pavlov, 1999),
 2 – ; 3 –
 („ „) „
 3,5 l/da.

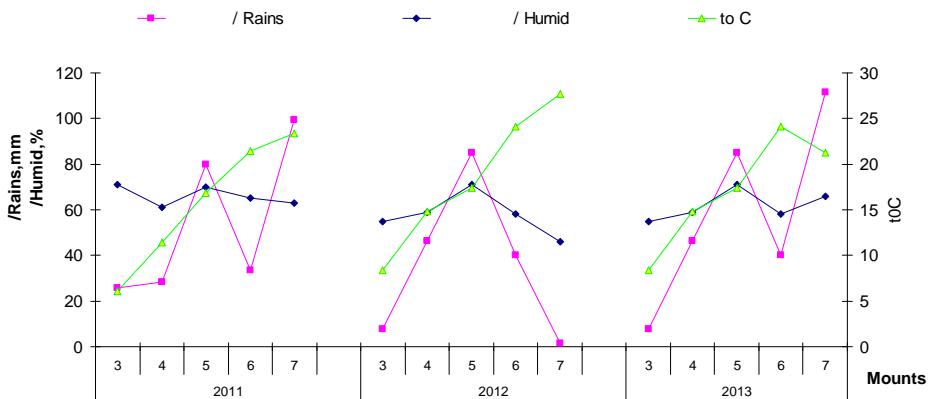
:
 (cm)
 () ;
 (cm) (L);
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 (kg/da);
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 (g). STATGRAPHICS plus for
 Windows Version 2.1.

earliness, resistant to lodging and good adaptability. It is suitable in the direction of grain and green mass (Kertikova et al., 2012). The study was conducted on the soil subtype slightly leached chernozem, without irrigation. It used the split plot method with four repetitions of the variants and a size of 10 m² of harvest plot. Variants of the field experience: Variant 1 control – a conventional technology (Kostov and Pavlov, 1999), including fertilization and treatment with herbicides and insecticides; Variant 2 - without the use of preparations of inorganic origin; Variant 3 - treatment only with bio insecticide ("Ecofil P") of organic origin. Treatment with bio preparation "Ecofil P" is performed in phenophase full flowering at a dose of 3,5 l/da.

Agro-meteorological and phenological observations and readings were carried. The following indicators were recorded: stand height (cm) in its natural condition before harvesting (H); stem length (cm) upright (L); lodging rate in % (C); economic and biological grain yield (kg/da); grain losses (kg/da). Grain harvest is done with miniature plot harvester. Structural analysis of grain yield include: stem length (cm); height of the first pod (cm); number of pods per plant; number of seeds per pod, seeds weight per plant (g); weight of 1000 seeds (g). The data were processed with the software STATGRAPHICS plus for Windows Version 2.1.

RESULTS AND DISCUSSION

Agro-meteorological conditions for the period March-July (Figure 1) show that the real opportunity for sowing of spring vetch and the three experimental years there in the third ten days of March – 21.03 - 28.03 (Table 1), i.e. as soon as possible to work under field conditions at the existing temperature conditions and good humid soil.



. 1.
Fig. 1. Klimatogram for the vegetation period of spring vetch cv. Tempo

- In March precipitation are relatively sufficient as a whole, but are not evenly distributed. Given the rising average daily soil and air temperature in April, complete germination in plants and the three options was registered at the beginning of the second decade of the month. Period phenophase germination to 3-4 leaf and the beginning of flowering is characterized by frequent rainfall, good humid and optimum temperature for development of the crop. In phenophase 50% flowering and liquid chromatography have three variants explored are no reported differences in their development. It appears in full flowering phases – beginning of pod formation – the end of May and beginning of June. The crop from the first variant (conventional technology) passes said two phases in a shorter period of time. Relative to the crop from a second variant (biological) this period is 3-4 days, and from 1-3 days compared to the crop of the third variant. This trend continued until the harvesting of spring vetch for grain production. In general weather conditions are characterized as favourable, with normal temperatures and rainfall during the growing of the spring vetch. The

| | | | | | | |
|---|----|-----|---|-----|---|---|
| (| 1) | 120 | (| 2). | - | measured vegetation period is in the range of 111 days (variant 1) to 120 days (variant 2). |
|---|----|-----|---|-----|---|---|

1.

, 2011-2013 .

Table 1. Number of plants, periods for the sowing and the harvesting and vegetation period in the spring vetch cv. Tempo, 2011-2013

| Treatment | m ² | Sowing | Harvesting | () |
|---------------------|-------------------------------------|-----------------|-----------------|--------------------------|
| | Number of plants per m ² | | | Vegetation period (days) |
| 1 – | | | | |
| Variant 1 – control | 446,20 | 21.03. - 28.03. | 10.07. - 16.07. | 111 - 117 |
| 2 / Variant 2 | 382,35 | 21.03. - 28.03. | 15.07. - 19.07. | 115 - 120 |
| 3 / Variant 3 | 384,12 | 21.03. - 28.03. | 12.07. - 18.07. | 112 - 119 |

| | | | |
|---------------------------|---|----|---|
| , | - | - | The data on the reported number of plants per unit of area in the storage of seed grain indicates that the use of standard technology to create more favourable cultivation conditions for the emergence and development of a large number of plants. |
| 446,20 ./m ² , | , | - | In this variant the number reaches 446,20 number/m ² , while the remaining two, the numbers of plants significantly reduced, respectively 382,35 number/m ² in biological and 384,12 number/m ² using "Ecofil P". |
| 382,35 ./m ² | , | - | In the results of the structural analysis of the yield (Table 2) on spring vetch, according to the appended growing technologies show significant differences. They are very good and good mathematical significant differences in individual indicators, compared to results of plants in the control variant. |
| 384,12 ./m ² | , | - | The length of the plants in the upright position is the greatest in the variant cultivated without the use of preparations of inorganic origin – 72,14 cm, followed by the length in the plants treated with "Ecofil P" – 69,00 cm. |
| " . | (| 2) | |
| , | , | , | |
| - 72,14 cm, | , | , | |
| " – 69,00 cm. | , | , | |

Table 2. Structural analysis of spring vetch cv. Tempo, 2011-2013

| Treatment | / Indicators | | |
|---------------------|---------------------------|------------------------------|------------------------------|
| | Length of the stems, (cm) | Height of the first pod (cm) | Number of pods per plant |
| 1 – | | | |
| Variant 1 – control | 67,10 ^c | 40,89 ^b | 10,58 ^a |
| 2 / Variant 2 | 72,14 ^a | 46,34 ^a | 8,17 ^b |
| 3 / Variant 3 | 69,00 ^b | 41,07 ^b | 8,75 ^b |
| | | | 1000 |
| Treatment | Number of seeds per pod | Seed weight per plant (g) | Weight of the 1000 seeds (g) |
| 1 – | | | |
| Variant 1 – control | 5,0 ^a | 13,09 ^a | 69,13 ^a |
| 2 / Variant 2 | 5,0 ^a | 12,77 ^a | 58,00 ^c |
| 3 / Variant 3 | 5,0 ^a | 13,10 ^a | 62,83 ^b |

LSD 99.5% –

LSD 99.5% – means of the same column followed by the same letter was not significant different

| | | |
|---------|---|---|
| - | - | Lowest are the plants of the variant with applied conventional technology – 67,10 cm. Most likely this is due to the higher level of weeding in the first and second variants and stronger downloading plants. |
| - | - | A similar trend is observed in the height of the first pod. The difference between height of the first pod plants at second and third variants is minimal and is not significant. The plants of the first variant form a relatively lower first pod – 40,89 cm. |
| - | - | The different heights of the first pod, largely determines the possibility of different pod formation and thus obtain different grain yield. The results show that when use the same variety, depending on the applied technology of cultivation, the height of the first pod, the number of reported pods varies between variants. |
| - | - | The largest number of pods per plant (10,58) formed plants of the first variant, while the second and third variants, the number of pods reduced to 8,75 to 8,17 units. |
| (10,58) | , | , |

| | | | |
|------|------|---|--|
| 8,75 | 8,17 | | Average for the period of the study the reported seeds per pod at the various options are equal in numbers. In the case here stronger factor is characteristic of biological factor compared to technology of growing. |
| , | , | 3 | The data listed in Table 3 show that the technology of cultivation of spring vetch grain production has a significant impact on the rate lodging at the time of harvesting the crop. |
| , | , | - | The obtained results are very good significant differences. The stand grown on conventional technology was greatly lodging. |

3.

, 2011-2013 .

Table 3. Degree of lodging depending on the technology of cultivation of cv. Tempo, 2011-2013

| Treatment | / Indicators | | |
|---------------------|--------------------|--------------------|--------------------|
| | H cm | L cm | (%) Lodging (%) |
| 1 – | | | |
| Variant 1 – control | 26,00 ^b | 83,67 ^b | 61,32 ^a |
| 2 / Variant 2 | 35,67 ^a | 86,33 ^a | 53,90 ^b |
| 3 / Variant 3 | 33,00 ^a | 83,00 ^b | 55,84 ^b |

LSD 99.5% –

LSD 99.5% – means of the same column followed by the same letter was not significant different

Note: H -
the natural state; L -
erect stems in the stand

| | | |
|---------------------------------|---------------------|--|
| 26,0 cm, 61,32%, 83,67 cm | | The reported natural height was 26,0 cm, and the degree of lodging is equal to 61,32% when measured length of 83,67 cm of upright stems. When the crops of the other two variants, thanks to a strong weeding, registered levels of lodging are lower, the differences have no mathematical significant. |
| 53,90% 55,84% | " " | In plants grown without use of preparations of inorganic origin lodging crop was 53,90% and 55,84% in plants treated with the "Ecofil P". |
| (| 4) | Reported results average for the period on grain yield of spring vetch cv. Tempo (Table 4) show that the |

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221,00 kg/da

165,50 kg/da.

differences between the three variants are significant. Biological and economic grain yield is highest at the crop grown in the conventional technology, respectively 221 kg/da and 165,50 kg/da.

4. (kg/da)

, 2011-2013

Table 4. Grain yield (kg/da) depending on the technology of cultivation of cv. Tempo, 2011-2013

| Treatment | Biological yield | Economic yield | Grain losses |
|-------------------------|---------------------|---------------------|--------------------|
| 1 – Variant 1 – control | 221,00 ^a | 165,50 ^a | 55,50 ^a |
| 2 / Variant 2 | 122,49 ^c | 98,90 ^c | 23,77 ^b |
| 3 / Variant 3 | 140,03 ^b | 115,40 ^b | 24,63 ^b |

LSD 99.5% –

LSD 99.5% – means of the same column followed by the same letter was not significant different

55,50 kg/da. -

,

(– 122,49 kg/da

98,90 kg/da) -

,

” ”, 140,03 kg/da

115,40 kg/da ,

23,77 kg/da.

In this stand however grain losses are high and reach 55,50 kg/da. The lowest values of the losses of the seed in a crop grown without the use of preparations of inorganic origin, but also under it, and yields (biological – 122,49 kg/da and economic – 98,90 kg/da) are the lowest.

Grain yields, as well as the losses in the treated stand with bio preparation "Ecofil P", occupy an intermediate position, respectively 140,03 kg/da biological and 115,40 kg/da economic yield in losses of 23,77 kg/da.

CONCLUSIONS

The crop grown in conventional technology is with two to six days shorter growing period of development with respect to the crops grown without the use of preparations of inorganic origin and those treated with bio insecticides "Ecofil P".

The values for the structural analysis of grain yield are higher in plants grown in conventional technology compared to those grown by treatment with bio insecticides "Ecofil P" and without the use of preparations of inorganic origin.

| | | | |
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| | | | At the time of harvesting of the spring vetch grain, the crop cultivated in the conventional technology is with the highest degree of lodging – 61,32%. |
| 61,32%. | - | (165,50 kg/da) | With the highest grain yield (165,50 kg/da) is distinguished the crop grown in the conventional technology, while the lowest (98,90 kg/da) crop grown without the use of preparations of inorganic origin. |
| | - | (98,90 kg/da) | |

/ REFERENCES

1. **Jimenez, M.**, 1999. Herbage and seed yield of common vetch (*V. sativa*), In: Proceedings of the XVI International Grassland Congress, Nice, France, pp. 571-572.
2. **Katayama, M.**, 1991. Studies of chlorinated indol auxins and antiauxin. *Chemical Regulation of Plants*, 26(1), 11-24.
3. **Kertikov, ..**, 2000. Establishing optimal seed and fertilizer norms of spring vetch for grain production. *Rastenievadni nauki*, 37(8), 625-628 (Bg).
4. **Kertikov, ..**, 2003. Quantitative and qualitative parameters of the yield of spring vetch (*Vicia sativa L.*) depending on harvesting phenophase. *Rastenievadni nauki*, 40(6), 525-531 (Bg).
5. **Kertikov, T.**, 2005. Comparative characterization of economic and qualitative Indices two varieties of spring vetch (*Vicia sativa L.*). *Bulgarian Journal of Agricultural Science*, 11(4), 475-481.
6. **Kertikov, T.**, 2010. Study on agronomic aspects of forage pea and vetch. Habilitation work, Pleven, Bulgaria.
7. **Kertikov, T. and V. Vasileva**, 2000. Effect of some biologically active substances on grain yield and its chemical composition in spring vetch. *Journal of Mountain Agriculture on the Balkans*, 3(2), 190-198.
8. **Kertikova, D., T. Kertikov, B. Ynkov, M. Pehlivanov and I. Dimitrov**, 2012. Tempo – a new variety of spring vetch. *Agrarni Nauki*, 11, 119-124 (Bg).
9. **Kostov, .., and D. Pavlov**, 1999. Forage production, Akademitchno izdatelstvo na VSI Plovdiv (Bg).
10. **Mousa, M., M. Ahmed and A. Mohamed**, 1997. Effect of phosphorus fertilizer and seeding rates of forage yield of *V. sativa*. *Egyptian Journal of Agricultural Research*, 70, 861-872.
11. **Orak, A.**, 2000. An investigation on yield and yield components of some common vetch (*V. sativa*) genotypes. *Acta Agronomica Hungarica*, 48, 295-299.
12. **Vasileva, V. and T. Kertikov**, 2007. Influence of the treatment with Humustim on sowing qualities and grain yield in spring vetch. *Field Crops Studies*, 4(2), 311-316 (Bg).
13. **Ynkov, B., I. Dimitrov and T. Kertikov**, 1995. Productivity of some vetch species and forms and their effect on the soil nutritive regime. *Rastenievadni nauki*, 32(6), 123-125 (Bg).

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Entomofauna of heteroptera in alfalfa agrocenoses

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SUMMARY

The report is a brief overview of entomofauna of heteroptera suborder in alfalfa agrocenoses – one of the richest and most numerous groups among insects. Presented are essential and economically important herbivorous bugs in various regions of the world, as a result of their feeding reduced productivity, seed quality and length of life of feed plants.

Described the mechanism of injury of the dominant species and damages that occur as a response in the plant organism. Discussed the composition, role and importance of heteroptera entomophaga for regulating the number of pests in alfalfa agrocenoses. Indicated some directions for future research.

: Heteroptera,

Key words: heteroptera, phytophaga, entomophaga, alfalfa agrocenoses

(*Medicago sativa* L.),

Alfalfa (*Medicago sativa* L.) due to its floristic and faunal status possesses a special microclimate suitable for the habitat of numerous insects and arthropods (Mirab-balou et al., 2007).

(Mirab-balou et al., 2007).

(Hemiptera: Heteroptera)

(Konjević and Kereši, 2014).

Among them herbivorous bugs (Hemiptera: Heteroptera) are basic and important pests of alfalfa and many other crops and their nutritional activity causes reduction of productivity, seed quality and duration of life of forage plants (Konjević and Kerešić, 2014).

Usually these pests are polyphagous and feed on wild plants, which are hosts and play an important role in raising the population levels of different types of bugs and are important food resources for the development of nymphs and adults reproduction.

World fauna of heteroptera order is divided into approximately 75 families, including the number of the most popular and well known terrestrial species ranging from 2,000 to 5,000 (Chaplin, 2009). According to Gerling et al (2001), one of the most abundant group of predator insects in alfalfa fields is precisely suborder Heteroptera.

In his collection of insects and mites from alfalfa fields, Harper (1988) found that 41 out of totally 437 species, from order Heteroptera, belong to the family Miridae, which is the highest number compared to the other families. Genus *Lygus* (Miridae) is represented by the most numerous species.

Fauna survey of representatives of the order Heteroptera in alfalfa crops in Serbia identify 26 species belonging to nine families. It is dominated by the family Pentatomidae, comprising more than 68% of the total number of bugs, followed by families Miridae (13%), Nabidae (6%), Lygaeidae (5%) and Anthocoridae (3%) (Konjević and Kerešić, 2014). Within the dominant family Pentatomidae, *Dolycoris baccarum* (Linnaeus, 1758) occupies more than 28% of all captured bugs, followed by *Eurydema ornatum* (Linnaeus, 1758) with 15% participation and *Piezodorus lituratus* (Fabricius, 1794) - with 7%. According to the authors this is the only kind of dominant family

| | | | |
|--|---|---|--|
| | Miridae | | |
| Lygus pratensis | (Linnaeus, 1758) (3%), <i>Chlamydatus pulicarius</i> (Fallen, 1807) (2%), <i>Stenodema calcarat</i> (Fallen, 1807) (1.5%), <i>Polymerus vulneratus</i> (Panzer, 1806) (1.5%) 1% | | |
| 14 | , | | |
| 10 | 6 | | |
| , | | | |
| Stenodema | <i>Lygus, Adelphocoris,</i> <i>Nezara</i> (Mirab-balou et al., 2007). | | |
| <i>L. pratensis</i> | <i>Lygus rugulipennis,</i> <i>A. lineolatus</i> (Mirab-balou et al., 2008). | | |
| | <i>Lygus,</i> | | |
| | | | |
| | (Mirab-balou et al., 2007). | | |
| , | | | |
| | <i>Deraeocoris</i> | | |
| Nabis, | , | | |
| | | | |
| | | | |
| 34 | , | | |
| | 24 | 8 | |
| : | Alydidae (<i>Camptopus</i>), | | |
| Anthocoridae (<i>Anthocoris Orius</i>), | | | |
| Berytidae (<i>Metacanthus</i>), Lygaeidae (<i>Geocoris, Nysius Leptodemus</i>), | | | |
| Nabidae (<i>Nabis</i>), Miridae (<i>Adelphocoris, Lygus, Polymerus, Stenodema, Campylomma Deraeocoris</i>), | | | |
| Pentatomidae (<i>Eysarcoris, Aelia, Eurydema, Dolycoris, Holcostethus Carpocoris</i>) Rhopalidae (<i>Brachycareus, Liorhyssus Rhopalus</i>). | | | |
| Adelphocoris lineolatus Goeze, 1778 | <i>Orius niger Wolff, 1804</i> | | |
| | | | |
| | (Havaskary et al., 2010). | | |
| Gharaat | (2009) | - | |
| , | | | |
| | | | |
| 73 | , | - | |
| 18 | . | | |

associated with alfalfa and other legumes. The second-largest family Miridae is represented by *Lygus pratensis* (Linnaeus, 1758) (3%), *Chlamydatus pulicarius* (Fallen, 1807) (2%), *Stenodema calcarata* (Fallen, 1807) (1.5%), *Polymerus vulneratus* (Panzer, 1806) (1.5%) and other species with less than 1% involved.

In western Iran reported a total of 14 types of bugs belonging to 10 genera and 6 families. From the collected species, herbivorous bugs belong to the genus *Lygus, Adelphocoris, Stenodema* and *Nezara* (Mirab-balou et al., 2007). With the highest numbers and economic importance are *L. rugulipennis, L. pratensis* and *A. lineolatus* (Mirab-balou et al., 2008). Among these major pests on seeds are species of *Lygus*, because of their feeding in phenophase blooms cause the most damaging and reducing yields in the second and third cut in alfalfa grown for seed (Mirab-balou et al., 2007). The authors report that among predatory bugs, species of the genus *Deraeocoris* and *Nabis*, which feed on aphids and nymphs of herbivorous bugs, are able to reduce populations of these pests to some extent.

In Northeast Iran established a total of 34 types of bugs belonging to 24 genera and 8 families: Alydidae (genus *Camptopus*), Anthocoridae (genera *Anthocoris* and *Orius*), Berytidae (genus *Metacanthus*), Lygaeidae (genera *Geocoris, Nysius* and *Leptodemus*), Nabidae (genus *Nabis*), Miridae (genera *Adelphocoris, Lygus, Polymerus, Stenodema, Campylomma* and *Deraeocoris*), Pentatomidae (genera *Eysarcoris, Aelia, Eurydema, Dolycoris, Holcostethus* and *Carpocoris*), Rhopalidae (genera *Brachycareus, Liorhyssus* and *Rhopalus*). Among them phytophaga Adelphocoris lineolatus Goeze, 1778 and predatory species *Orius niger Wolff, 1804* are the dominant species in fields of study area (Havaskary et al., 2010).

Gharaat et al. (2009) reported that entomofauna of heteroptera order in eastern and western Azerbaijan is represented by 73 species belonging to

| | |
|---|-------------------------|
| | : |
| Anthocoridae - 8, Berytidae - 1, Coreidae - 4, Cydnidae - 2, Miridae - 20, Nabidae - 2, Reduviidae - 3, Tingidae - 1, Corixidae - 3, Notonectidae - 1, Gerridae - 2, Hydrometridae - 1, Alydidae - 1, Lygaeidae - 7, Pentatomidae - 12, Pyrrhocoridae - 2, Scutelleridae - 2 Stenocephalidae - 1. | |
| <i>Mozena lunata</i> (Burmeister, 1835 .) | |
| (Coreidae) | |
| | - |
| | - |
| Anthocoridae (41.5%), Miridae (29.3%) (7.4%). | Pentatomidae |
| | Khaghaninia |
| | (2013) |
| | , 650 |
| | , 29 |
| 12 | Anthocoridae, |
| Miridae, Tingidae, Nabidae, Lygaeidae, | |
| Pyrrhocoridae, Coreidae, Stenocephalidae, | |
| Rhopalidae, Cydnidae, Scutelleridae | |
| Pentatomidae. | |
| | - |
| | - |
| 8 | , |
| 6 | . |
| | - |
| | <i>Deracoris</i> sp. |
| <i>Campylomma diversicornis</i> Reuter, 1878, | |
| <i>Lygaeus pandurus</i> Scop. | |
| <i>Pyrrhocorius apterus</i> (Linnaeus 1758) | |
| | - |
| (Augul et al., 2012). | |
| <i>Orius albidipennis</i> <i>Deracoris</i> sp. | |
| | - |
| | Mirab-balou |
| (2007), | . |
| <i>Orius</i> | . |
| | , |
| | , |
| (Augul et al., 2012). | |
| | Hemiptera |
| | , |
| | , |
| | <i>L. lineolaris</i> A. |
| <i>lineolatus</i> (Jensen et al., 1991). | |
| | , |
| | , |
| | , |
| | , |
| | Mueller |
| (2005) | , |

18 families. The identified families and the number of species are as follows: Anthocoridae - 8, Berytidae - 1, Coreidae - 4, Cydnidae - 2, Miridae - 20, Nabidae - 2, Reduviidae - 3, Tingidae - 1, Corixidae - 3, Notonectidae - 1, Gerridae - 2, Hydrometridae - 1, Alydidae - 1, Lygaeidae - 7, Pentatomidae - 12, Pyrrhocoridae - 2, Scutelleridae - 2 and Stenocephalidae - 1. *Mozena lunata* (Burmeister, 1835) (Coreidae) is reported as a new species of Palearctic ecozone. The most diverse and the highest numbers are the families Anthocoridae (41.5% of total), Miridae (29.3%) and Pentatomidae (7.4%). In a later study Khaghaninia et al (2013) reported about 650 collected species, 29 of them belong to 12 families, including Anthocoridae, Miridae, Tingidae, Nabidae, Lygaeidae, Pyrrhocoridae, Coreidae, Stenocephalidae, Rhopalidae, Cydnidae, Scutelleridae, Pentatomidae.

Species of Heteroptera in alfalfa in Abu Ghraib, Baghdad represented by 8 species belong to 8 genera and 6 families. The most common species are *Deracoris* sp. and *Campylomma diversicornis* Reuter, 1878, while *Lygaeus pandurus* Scop. and *Pyrrhocorius apterus* (Linnaeus 1758) are the lowest numbers in the study period (Augul et al., 2012). In addition, the species *Orius albidipennis* and *Deracoris* sp. are also registered in high numbers and authors confirm the findings in an earlier study of Mirab-Balou et al. (2007) that the predatory bugs of the genus *Orius* have high density, and feed on herbivorous mites, insect eggs, aphids, thrips and small caterpillars. (Augul et al., 2012).

Major pests of Hemiptera in alfalfa fields, used for forage in Washington, are *L. lineolaris* and *A. lineolatus* (Jensen et al., 1991). The authors found that the damage caused by nymphs and adults, is associated with a reduction of dry weight and length of the stems, especially in soon cut crops and increases the crude protein content. In a later study, Mueller et al. (2005) reported that the species composition of the genus *Lygus* in alfalfa

| | | | |
|---------------------|--|------------------|--|
| | <i>Lygus</i> | , | |
| (Knight) | <i>Lygus hesperus</i> | | |
| | <i>L. elisus</i> (van Duzee) | , | |
| | | , | |
| | | 1: 1. | |
| | , | , | |
| | , | , | |
| Miridae, | <i>M. sativa</i> | | |
| | : | | |
| | <i>A. lineolatus</i> (Goeze), <i>A. rapidus</i> (Say), <i>Lopidea marginalis</i> (Reuter), <i>Lygus lineolaris</i> (Palisot de Beauvois), <i>Plagiognathus chrysanthemi</i> (Wolff) <i>P. politus</i> Uhler (Wheeler, 2012). | | |
| | , | , | |
| Miridae | | , | |
| | | , | |
| Miridae | Miridae | | |
| | 200 | | |
| | (Lu and | | |
| Wu, 2011). | , | , | |
| , | , | , | |
| Heteroptera, | | , | |
| | , | , | |
| | , | , | |
| | (Wheeler, 2001). | | |
| | Miridae | (<i>Lygus</i> , | |
| <i>Adelphocoris</i> | .), | , | |
| | , | , | |
| | , | , | |
| | (Lu and Wu, | | |
| 2011). | | | |
| | | , | |
| | (Khanjani, 2005). | | |
| | , | , | |
| | , | , | |
| | , | , | |

grown for seed and forage is represented only by *Lygus hesperus* (Knight) and *L. elisus* (van Duzee) in the San Joaquin Valley California as sex ratio male to female is 1:1.

In Ithaca, New York are studied feeding habits, seasonal dynamics, population density and biology of species of Miridae, associated with *M. sativa* and established the following phytophaga: *A. lineolatus* (Goeze), *A. rapidus* (Say), *Lopidea marginalis* (Reuter), *Lygus lineolaris* (Palisot de Beauvois), *Plagiognathus chrysanthemi* (Wolff) and *P. politus* Uhler (Wheeler, 2012). In conclusion, it reported that predatory insects Miridae are frequently found in lucerne crops.

Family Miridae

Bugs of the family Miridae are polyphagous insects with more than 200 species of host plants (Lu and Wu, 2011). According to the authors the recent appearance of these bugs and damages caused by them, will progressively increase. This is the largest family in the suborder Heteroptera, where there is a huge morphological diversity and trophic plasticity, and consists of number of key phytophaga and predators (Wheeler, 2001).

The species of Miridae (*Lygus*, *Adelphocoris* etc.), with their oral apparatus for thrusting and sucking, are preferentially fed with plant tissues with a high content of nutrients such as apical meristems and primordial cells forming the leaves, or the formation of the reproductive tissues (Lu and Wu, 2011). The representatives of this family are key pests during flowering (Khanjani, 2005). The most common symptoms as a result of the inflicted damage, are forming of an excessive number of branches, small, deformed leaves and necrosis at the border of the leaves, death of buttons abortion of flowers, weak flowering and formation of shriveled seeds and seeds with reduced mass of 1000 seeds

(Blackmer et al., 2004).

(Leal, 2013).
Linnaviuori (2007)

204 , Miridae

-
L. rugulipennis,

(Khanjani, 2005).
Adelphocoris

fasciaticollis Reuter
2015). (Zhang et al.,

Miridae - A.
lineolatus Lygus (-
Exolygus), -

L. rugulipennis, *L. pratensis* *L. gemellatus*.

25-50%, (Sekuli et
al., 2005).

() -
Lygus lineolaris, *L. elisus*
L. borealis, *L. lineolaris* -

Adelphocoris *A. lineolatus* -
lineolaris , L.

(Nagalingam and
Holliday, 2010).
Lygus spp. (Hemiptera: Miridae)

L. lineolaris, *L. elisus* *L. hesperus* L.

plants do not contain (Blackmer et al., 2004). The attraction is due to the key proteins in the olfactory receptor neurons through which the olfactory system of bugs cause dynamic process of perception of odors and their differentiation (Leal, 2013).

Linnaviuori (2007) presents a list of 204 species belonging to Miridae in exploration in Gilan and neighboring provinces of Iran as one of the most important pest is *L. rugulipennis*, occupying a key role in the flowering phase of the seed production (Khanjani, 2005). In northern China *Adelphocoris fasciaticollis* Reuter became a key pest on alfalfa (Zhang et al., 2015).

Most common species of family Miridae in Serbia are *A. lineolatus* and species of the genus *Lygus* (synonymous *Exolygus*), which are particularly harmful in seed productive alfalfa crops. Among *Lygus*, *L. rugulipennis*, *L. pratensis* and *L. gemellatus* are dominating. They are present throughout the all growing period and are especially numerous in July and August. By sucking plant sap, they injure youngest plant parts, stop the growth and cause deformation of the seeds. The losses in the yield of seeds often reach 25-50% and sometimes occurs a complete loss of seed yield (Sekuli et al., 2005).

In southern Manitoba (Canada) the major pests are *L. lineolaris*, *L. elisus* and *L. borealis*, amongst which *L. lineolaris* is the most numerous and widespread. Several species of the genus *Adelphocoris* are important, but *A. lineolatus* is the dominant. It is assumed that the species, along with *L. lineolaris*, have similar effects on alfalfa and the method of calculating the economic threshold of harm refers to both species (Nagalingam and Holliday, 2010).

Lygus spp. (Hemiptera: Miridae) originates from North America and the most common species in alfalfa are *L.*

lineolaris, *L. elisus* and *L. hesperus* as *L. lineolaris*, *L. hesperus*, *A. lineolatus* are the most economically important plant bugs in North America (Blackmer et al., 2010; Henry and Wheeler, 2014). Several species of *Lygus* have two or more generations in the southern latitudes of the prairies. Within a species, the number of generations may vary directly with the accumulation of temperatures above 10 °C (Champlain and Butler, 1976). Some species have a wide range of hosts such as *L. lineolaris* feeds on over 300 plant species from different families, and *Lygus rugulipennis* – more than 400 species of plants (Young, 1986; Capinera, 2001; Lefort et al., 2014). These species are very mobile and are usually found in high numbers. Information on the preference of important insect pests such as bugs from the genus *Lygus*, may be useful in predicting their occurrence and future movement between crops and wild host plants. It is estimated that between five studied hosts: cotton, alfalfa (*Medicago sativa* L.), wild sunflower (*Helianthus annuus* L.), Russian thistle (*Salsola iberica* L.) and amaranth (*Amaranthus palmeri* L.), alfalfa and thistle are the two most preferred hosts with a high degree of damage (Barman et al., 2010).

Species of the genus *Lygus* spp. may not have a significant impact on the physiological condition of the plant, but economic importance tackle the onset of the reproductive phase of development by acquiring the status of the most important pests of alfalfa grown for seed, as feed on the reproductive parts of the plant and reduced yield seeds (Day et. al., 2003; Jorgensen, 2005). In alfalfa grown for forage, bugs do not usually affect productivity, but crops can serve as a reservoir from which they migrate after cutting (Day et. Al., 2003).

Lygus bugs are the most important pests in seed production of alfalfa in Idaho, USA and Vashington especially when exceed the economic threshold

eastern half of the continent (Burange et al., 2012). In France and Spain, *Lygus rugulipennis* and *L. pratensis* are major pests of the genus *Lygus* and a successful biological control upon their number carry out pupal parasitoids *Peristenus digoneutis* and *P. stygicus* (Coutinot et al., 2005). To similar conclusions in Poland reaches Bilewicz-Pawinska (2006). According to her the most common parasitoids in *L. rugulipennis*, *Stenodema virens* (L.) and *A. lineolatus* are species of the genus *Peristenus* spp. (Hymenoptera: Braconidae), which play an important role in reducing the populations of the three harmful species. It was found that parasitism reached the highest level when the nymphs of plant bugs are in the highest quantity. In California by deliberately introducing parasitoids *P. digoneutis* and *P. relictus* (Ruthe) monitor successfully reduction in number of *Lygus* spp. (Pickett et al., 2009) as the peak of parasitism coincide with the peak activity of *Lygus* bugs. The natural presence of *Peristenus* spp. in seed production crops is associated with low levels of parasitism, reaching only to 8.9%, so do not expect a significant impact on bioagents on reducing population density of species of the genus *Lygus* and damages caused by them (Schreiber, 2008).

Adelphocoris lineolatus (Goeze, 1778) (Miridae) is a species from Old World widespread in Western Europe, North Africa, the Middle East and much of Asia (Wheeler, 2001), but was imported to North America (Scudder and Foottit, 2006). This pest is characterized by a wide range of host and feeds mainly on plant petals and flower, causing premature petal fall. The species is one of the most damaging and widespread pests in Ukraine, which represents 56% of the total number of insect pests of alfalfa seed (Grikun al., 2008).

Alfalfa bug, because of their feeding, can reduce seed yield of crops by 50% or more (Wheeler, 2001).

| | |
|--|-----------------------|
| (Sweet, 2000). | |
| et al., 2012). | <i>Lygaeus</i> (Augul |
| (Lygaeidae) | <i>Geocoris</i> |
| , | , |
| , | , |
| , | <i>Geocoris</i> spp. |
| , | , |
| (Hagler and Cohen, 1994). | , |
| <i>Geocoris pallidipennis</i> | , |
| | <i>A.lineolatus</i> (|
| 24 |) |
| | 37.9 |
| | 1- |
| 15 °C 35 °C, 1.78 | , |
| 35 °C, 15 °C (Tong et al., | , |
| 2011). , <i>G. pallens</i> Stål G. | , |
| <i>bullatus</i> Say | , |
| 16 : <i>Cymus glandicolor</i> H, <i>Cymus</i> <i>melanocephalus</i> Fieb., <i>Geocoris arenarius</i> (Jak.), <i>Geocoris ater</i> (F.), <i>Picocoris</i> <i>erythrocephalus</i> (P.S.), <i>Lygaeus saxatilis</i> (Scop.), <i>Melanocoryphus tristrami</i> (D.Sc.), <i>Nysius cymoides</i> Spin, <i>Nysius ericae</i> (Schl.), <i>Nysius graminicola</i> (Klt.), <i>Oxycarenus longiceps</i> Wgn., <i>Oxycarenus</i> <i>pallens</i> (H.-S.), <i>Emblethis griseus</i> W., <i>Lamprodema maurum</i> F., <i>Lethaeus</i> <i>cibratissimus</i> (Stal), <i>Lethaeus picipes</i> (H.-S.). (Önder et al., 1999). | |

stress and strong drought, migrate from wild plant species - hosts to alfalfa and cause considerable damages (Sweet, 2000). Into phytophaga, damaging lucerne seed also relates species of the genus *Lygaeus* (Augul et al., 2012).

Species of the genus *Geocoris* (Lygaeidae) are among the most numerous and important predators of insects in many agro-ecosystems that feed on small insects, butterflies eggs, nymphs of herbivorous bugs, aphids and mites. The potential of *Geocoris* spp. as bioagents is justified, because adults and immature individuals can consume dozens of specimens per day (Hagler and Cohen, 1994). For example, *Geocoris pallidipennis* as a potential biological agent for *A. lineolatus* (in China) has a high consuming power - for 24 hours adult females feed on 37.9 nymphs of phytophaga bug in the first stage of its development. In addition, predation rate gradually increases with increase of temperature from 15 °C to 35 °C and it is 1.78 times higher at 35 °C, than at 15 °C (Tong et al., 2011). In Montana, *G. pallens* Stål and *G. bullatus* Say are reported as one of the main predators that feed on eggs and nymphs of pest *Lygus* genus in alfalfa (Blodgett, 2009).

As a result of 8-year fauna study of the family Lygaeidae in the Western Black Sea, Central Anatolia and the Mediterranean region of Turkey, which are characterized by different climates and habitats in alfalfa, have been identified 16 species. They are *Cymus glandicolor* H., *Cymus melanocephalus* Fieb., *Geocoris arenarius* Jak., *Geocoris ater* F., *Piocoris erythrocephalus* PS., *Lygaeus saxatilis* Scop., *Melanocoryphus tristrami* D.-Sc., *Nysius cymoides* Spin., *Nysius ericae* Schl., *Nysius graminicola* Klt., *Oxycarenus longiceps* Wgn., *Oxycarenus pallens* H.-S., *Emblethis griseus* W., *Lamprodema maurum* F., *Lethaeus cibratissimus* Stal, *Lethaeus picipes* H.-S. (Önder et al., 1999).

Heteroptera is the richest and most numerous suborder, presented by predatory insects in alfalfa, the most common species belong to families Nabidae, Anthocoridae and Miridae (Pons et al., 2005). Bugs predators, including *Nabis* spp., comprise from 40 to 89% of the total number of predatory insects in Argentina (Cornelis et al., 2012). Similar results reported Razmjoo (2012) as the dominant species of the genus *Deracoris*, *Nabis*, *Orius* and *Geocoris* constitute 60% of the total number of individuals collected in the Central part of Iran.

Family Nabidae

Nabidae family consists of 31 genera and about 386 species distributed in all biogeographical regions of the world (Henry, 2009). Genus *Nabis* belongs to the subfamily Nabinae and is represented by 15 species (Volpi and Coscarón, 2010). Bugs of this genus are polyphaga predators, feeding on a wide variety of small arthropods such as aphids, *Lygus* bugs and others, and many of them have a high numerical value in crops such as alfalfa, soybean, cotton and others. (Mirabbalou, et al., 2007). In the absence of enough food *Nabis* species can express cannibalistic aptitude to each other (Wheeler, 2001). Jorgensen (2005) found that when choosing between species of *Lygus* and *Acyrthosiphon pisum*, predatory bugs of the genus *Nabis* spp. prefer to consume nymphs of *Lygus* bugs. In support of this assertion, amprag (2007) reported that *Nabis* species feed mainly with representatives of the family Miridae, and especially with *Lygus* genus, which explains the high number of predators in lucerne fields where *Lygus* dominate.

Nabis ferus L. has a key role in the critical reduction in the number of nymphs of herbivorous bugs and aphids (Mirabbalou et al., 2007) and it is the dominant species in different geographical areas. As common predators, feeding on herbivores bugs and other pests present

Nabis provencalis Remane, *Nabis capsiformis* (Germar, 1837) . (Pons et al., 2005; Cornelis et al., 2012)

Anthocoridae

Anthocoridae

(Bokina, 2008). Anthocoridae
(Jorgensen, 2005; Blodgett, 2009).
Orius niger Wolff., *O. minutus* L., *O. majusculus* *Anthocoris confosus* Reut. *O. minutus* *O. niger*

(Pons et al., 2005; Mirab-balou, et al., 2007; Konjevi and Kereši, 2014).
Orius

Orius,

(Bahsi and Tunç, 2008; Bosco and Tavella, 2013).

Orius,

(Bosco and Tavella, 2008; Bosco et al., 2008).

Miridae,
Deraeocoris pallens Rt., *D. ruber* (L.), *D. serenus* Dgl. Sc., *Campylomma verbasci* (Meyer-Duer, 1843),

Lygus
(Lodos et al., 2003;
Augul et al., 2012).

Pyrrhocoridae,

in alfalfa, depending on geographical areas, *Nabis provencalis* Remane, *Nabis capsiformis* (Germar, 1837) and others are reported (Pons et al., 2005; Cornelis et al., 2012).

Family Anthocoridae

The species of family Anthocoridae feed on small insects, mites and eggs. They kill their prey, through pre-piercing it with stiletto, injecting salivary secretion and suck body fluids. It defines them as useful biological agents that control the numbers of insect pests (Bokina, 2008). Anthocoridae are common predators in seed production areas with alfalfa (Jorgensen, 2005; Blodgett, 2009). From this family, *Orius niger* Wolff., *O. minutus* L., *O. majusculus* and *Anthocoris confosus* Reut. are prevalent. *Orius minutus* and *O. niger* are essential for a number of herbivorous mites, eggs of insects, aphids, thrips and small caterpillars, due to the rapid development of all their stages, which identifies them as very effective predators (Pons et al., 2005; Mirab-balou, et al., 2007; Konjevi and Kereši, 2014). In addition, the species of the genus *Orius* have a key role in controlling of the density of phytophaga thrips worldwide. Prevailing on alfalfa *Orius* are known for important bioagents against harmful thrips in Europe and the Middle East (Bahsi and Tunç, 2008; Bosco and Tavella, 2013). In fact, the most important role in the biological control of thrips have species of the genus *Orius*, which have proved good adaptability to the weather conditions (Bosco and Tavella, 2008; Bosco et al., 2008).

The useful species of Miridae, *Deraeocoris pallens* Rt., *D. ruber* L., *D. serenus* Dgl. Sc., *Campylomma verbasci* Meyer-Duer, found in high density and have an important role in reducing the number of aphids and nymphs of the genus *Lygus* in alfalfa fields (Lodos et al., 2003; Augul et al., 2012).

From the family Pyrrhocoridae,

Pyrrhocoris apterus L. is a common species in alfalfa that feeds with various insects, as well as the seeds, plants on the surface and others, but does not constitute a threat to alfalfa (Mirabbalou, et al., 2007 ; Augul et al., 2012).

In Bulgaria more thorough studies on quantitative and qualitative composition of the suborder Heteroptera in agrobiocenoses of alfalfa were made in the 60 years in Sofia district and Plovdiv district (Iosifov 1962; Popova, 1966). They are related to biology, as well as seasonal and quantitative dynamics of the main heteroptera entomofauna, which has great practical importance for a successful and targeted control on harmful bugs. Popova (1966) sets 17 species phytophaga, of which a major dominant species is *Lygus rugulipennis* (42.5%) (Miridae). Significantly, lower part of the same family has *L. pratensis* (8.2%) and *Adelphocoris lineolatus* (2.4%). From Pentatomidae, only *Piezodorus lituratus* is more closely related to alfalfa, but it is present insignificantly in crops. In a comparative analysis between the different categories (harmful, useful and indifferent), the author found that the harmful bugs predominate significantly over the others, and among predators the dominant species is *Nabis pseudoferus* (29.6%). Similar results for major harmful heteroptera species in alfalfa reported later Pirgozlieva and Dochkova (1996).

In a recent study, Atanasova (2006), for the region of Plovdiv, established 13 species of harmful bugs, the predominant species is *Lygus rugulipennis* (44.7%), followed by *Adelphocoris lineolatus* (37.1%) and *L. pratensis* (13.3%) (Miridae). The family Pentatomidae is represented mainly by *Nezara viridula* (1.8%) and *Piezodorus lituratus* (1.2%). The species of the genus *Nabis*, *Orius*, *Deraeocoris* and *Geocoris* are reported to be predators.

Regardless of the earlier conducted studies in Bulgaria, there is a definite necessity for relevant and profound research on species composition.

population dynamics of the most common bugs, determination of objective laws regarding the formation of phytoagroecosystem under different climatic conditions and seasons of the year as well as biocenological dependencies between alfalfa and insects. The ultimate aim is controlling the population of phytophaga insects, preservation of the diversity and abundance of natural enemies that govern the population dynamics of the pest, and where reasonable to apply the most effective environmental protection measures for the control of economically important pests.

/ REFERENCES

1. **Atanasova, D.**, 2006. Species composition and population density dynamics of economically important pests in multifoliate alfalfa, grown for hay, in the region of Plovdiv. Agricultural University - Plovdiv, *Scientific Works*, vol. LI, 17-24 (Bg).
2. **Augul, RSH., NN. Mzhr and M. S. Abul-Rassoul**, 2012. Occurrence of hemipteran species on alfalfa plant in Baghdad. *Bull. Iraq nat. Hist. Mus*, 12 (2), 7-13.
3. **Bah i, .Ü. and I. Tunç**, 2008. Development, survival and reproduction of *Orius niger* (Hemiptera: Anthocoridae) under different photoperiod and temperature regimes. *Biocontrol Science and Technology*, 18 (8), 767-778.
4. **Bancroft, J.S.**, 2005. Dispersal and abundance of *Lygus hesperus* in field crops. *Environmental entomology*, 34 (6), 1517-1523.
5. **Barman, A.K., M.N., Parajulee and S.C. Carroll**, 2010. Relative preference of *Lygus hesperus* (Hemiptera: Miridae) to selected host plants in the field. *Insect Science*, 17: 542-548. doi:10.1111/j.1744-7917.2010.01334.x
6. **Bilewicz-Pawinska, T**, 2006. Natural enemies of some plant bugs (Miridae) in Poland. *Roczniki Nauk Rolniczych*, 6(1), 125-135.
7. **Blackmer, J.L., C. Rodriguez-Saona J.A. Byers, K.L. Shopeand and J.P. Smith**, 2004. Behavioral response of *Lygus hesperus* to conspecifics and headspace volatiles of alfalfa in a Y-tube olfactometer. *J. Chem. Ecol.*, 30: 1547-1564.
8. **Blackmer, LJ., AJ. Byers and C. Rodriguez-Saona**, 2010. Evaluation of color traps for monitoring *Lygus* spp.: design, placement, height, time of day, and non-target effects. Second International Lygus Symposium Asilomar Conference Grounds, Pacific Grove, California, April 15-19 2007. *Journal of Insect Science*, 8 (49); 4.
9. **Blodgett, S**, 2009. Lygus and Alfalfa Plant Bugs. Alfalfa. High Plains Integrated Pest Management. Montana State University, 2009, Bozeman, Montana. http://wiki.bugwood.org/HPIPM:Crops/Alfalfa_Seed/Lygus_and_Alfalfa_Plant_Bugs [accessed 15 March 2011].
10. **Bochkareva, Z.A. and S.M. Vdovichenko**, 2004. The protection of lucerne seed crops. *Zashchita Rastenii*, 7, pp. 19.
11. **Bokina, I. G**, 2008. On the biology of anthocorid bugs (Heteroptera, Anthocoridae), predators of grass aphids in the forest-steppe zone of West Siberia. *Entomological Review*, 88 (8), 1060-1063.
12. **Borbour, J**, 2007. Insecticide resistance and efficacy of novaluron and flonicamid for control of Lygus bugs in alfalfa seed. Second International Lygus

Symposium Asilomar Conference Grounds, Pacific Grove, California, April 15-19 2007.
Journal of Insect Science, 8 (49), 3

13. **Bosco, L. and L. Tavella**, 2008. Collection of Orius species in horticultural areas of northwestern Italy. *Bulletin of Insectology*, 61: 209-210.
14. **Bosco, L. and L. Tavella**, 2013. Distribution and abundance of species of the genus Orius in horticultural ecosystems of northwestern Italy. *Bulletin of Insectology*, 66 (2), 297-307. ISSN 1721-8861.
15. **Bosco, L., E. Giacometto and L. Tavella**, 2008. Colonization and predation of thrips (Thysanoptera: Thripidae) by Orius spp. (Heteroptera: Anthocoridae) in sweet pepper greenhouses in Northwest Italy. *Biological Control*, 44: 341-340.
16. **Burange, P.S., R.L. Roehrdanz and M.A. Boetel**, 2012. Geographically based diversity in mitochondrial DNA of North American Lygus lineolaris (Hemiptera: Miridae). *Ann. Entomol. Soc. Am.*, 105(6), 917-929.
17. **Burdfield-Steel, E.R. and D.M. Shuker**, 2014. The evolutionary ecology of the Lygaeidae. *Ecol Evol*, 4(11), 2278-2301.
18. **amprag, D**, 2007. Razmnožavanje šteto ina ratarskih kultura u Srbiji i susednim zemljama tokom 20. veka. SANU - Ogranak u Novom Sadu, Novi Sad, 2007, pp. 348 (Sr).
19. **Capinera, JL**, 2001. Handbook of Vegetable Pests. Academic Press, San Diego, pp. 729.
20. **Champlain R. A. and G.D.Jr. Butler**, 1976. Temperature effects on development of the egg and nymphal stages of Lygus hesperus (Hemiptera: Miridae). *Ann. Entomol. Soc. Am.*, 60: 519-521.
21. **Chaplin, N**, 2009. The world Mirids. last update 22 December 2009. <http://www.faunaeur.org/>
22. **Conti, E. and F. Bin**, 2006. Native Lygus spp. (Heteroptera: Miridae) damaging introduced Hibiscus cannabinus in Italy. *J. Econ. Ento.*, 94: 648-657.
23. **Cornelis, M., E.M. Quirán and M.C. Coscarón**, 2012. Description of some immature stages of Nabis (Tropiconabis) capsiformis (Hemiptera: Nabidae). *Revista Mexicana de Biodiversidad*, 83: 1009-1012.
24. **Coutinot, D., K.A. Hoelmer and C.H. Pickett**, 2005. Exploration in Europe, exportation and establishment of parasitoids for biological control of Lygus hesperus (Hem. : Miridae) in California. 8pp. 7th International Conference on Pests in Agriculture, Montpellier, France. 26-27 Octobre 2005.
25. **Day, W.H., A.T. Eaton, R.F. Romig, K.J. Tilmon, M. Mayer and T. Dorsey**, 2003. Peristenus digoneutis (Hymenoptera: Braconidae), a parasite of Lygus lineolaris (Hemiptera: Miridae) in northeastern United States alfalfa, and the need for research on other crops. *Entomological News*, 114: 105-111.
26. **Després, L., JP. David and C. Gallet**, 2007. The evolutionary ecology of insect resistance to plant chemicals. *Trends Ecology and Evolution*, 22: 298-307.
27. **Domek, J.M. and D.R. Scott**, 1985. Species of the genus Lygus Hahn and their host plants in the Lewiston-Moscow area of Idaho. *Entomography*, 3: 75-105.
28. **Ebrahimi, A., R. Hosseini and R.V. Shoushtari**, 2012. A faunal study of plant bugs (Hemiptera: Miridae) in Ghorveh and its counties (Kurdistan province, Iran). *Ansfelden*, 2: 25-38.
29. **Erdelyi, C. and P. Benedek**, 1974. Effect of climate on the density and distribution of some Mirid pests of lucerne (Heteroptera: Miridae). *Acta Phytopathologica Academiae Scientiarum Hungaricae*, 9(1-2), 167-176.
30. Fauna Europaea version 2.5. <http://www.faunaeur.org>
31. **Gerling, D., O. Almar and J. Amo**, 2001. Biological control of Bemesia tabaci

predators and parasitoids. *Crop Protection*, 20: 779-799.

32. **Gharaat, M.A., M. Hassanzadeh, M.H. Safaralizadeh and M. Fallahzadeh**, 2009. Notes on the true bug (Heteroptera) fauna of Azerbaijan province. *Iran. Turk J Zool*, 33: 421-431. doi:10.3906/zoo-0804-3
33. **Hagler, J.R. and A.C. Cohen**, 1994. Prey selection by in vitro- and field-reared *Geocoris punctipes*. *Entomol. Exp. Appl.*, 59: 201-205.
34. **Havaskary, M., F.Hossein Pour and M. Modares Awal**, 2010. Cimicomorpha and Pentatomomorpha (Heteroptera) of alfalfa from Mashhad and vicinity, NE Iran. *Munis Entomology and Zoology*, 5 (1), 253-261.
35. **Henry, T.J.**, 2009. Biodiversity of Heteroptera. In *Insect Biodiversity: science and society*, R. Foottit and P. Adler (eds.). Wiley-Blackwell, pp. 223-263.
36. **Henry, T.J. and Jr.A.G. Wheeler**, 2014. Plant bugs on alfalfa. In: Samac D., L. Rhodes and W. Lamp, editors. *Compendium on Alfalfa Diseases*. St. Paul, MN: APS Press., pp. 93-95.
37. **Hrykun, OA., LC. Antipova and BC. Kryvoguz**, 2008. Insect complex in crops of alfalfa. *Scientific Production Journal*, April, 142 (4), 16-19 (Ukr).
38. **Jensen, B.M., J.L. Wedberg and D.B. Hogg**, 1991. Assessment of damage caused by tarnished plant bug and alfalfa plant bug (Hemiptera: Miridae) on alfalfa grown for forage in Wisconsin. *Journal of economic entomology*, 84 (3).
39. **Jorgensen, A. E.**, 2005. Conserving natural enemies control *Ligus* in Washington state alfalfa fields. PhD theses Washington state University, Department of Entomology, pp. 1-55.
40. **Jorgensen, A.E.**, 2005. Conserving Natural Enemies to Control *Lygus* in Washington State Alfalfa Fields. M.Sc. thesis, Washington State University, Pullman.
41. **Khaghaninia, S., R. Farshbaf Pour, O. Abad Askari and M. Havaskary**, 2013. A faunistic study of true bugs (Heteroptera) from Horand grasslands, NW Iran. *Munis Entomology & Zoology*, 8 (1), 468-474.
42. **Khanjani, M.**, 2005. Field Crop Pests in Iran (3rd edition). Bu-Ali Sina University Publication, Iran, pp. 732.
43. **Khanjani, M.**, 2007. Vegetables Pests in Iran (3rd edition). Bu-Ali Sina University Publication, Iran, pp. 467.
44. **Konjevi , A. and T. Kereši**, 2014. Fauna of Heteroptera in alfalfa fields in the region of Ba ka (Northwest Serbia): past and present situation. *Research Journal of Agricultural Science*, 46 (2), 115-124.
45. **Leal, W.S**, 2013. Odorant reception in insects: roles of receptors, binding proteins, and degrading enzymes. *Annual Review of Entomology*, 58: 373-391.
46. **Lefort, F., D. Fleury, I. Fleury, C. Coutant, S. Kuske, P. Kehrli and P. Mainget**, 2014. Pathogenicity of Entomopathogenic Fungi to the Green Peach Aphid *Myzus persicae* Sulzer (Aphididae) and the European Tarnished Bug *Lygus rugulipennis* Poppius (Miridae). *Egyptian Journal of Biological Pest Control*, 24(2), 379-386.
47. **Linnavuori, R.E.**, 2007. Studies on the Miridae (Heteroptera) of Gilan and the adjacent provinces in northern Iran. II. List of species. *Acta Entomologica Musei Nationalis Pragae*, 47: 17-56.
48. **Lodos, N., F. Önder, W. Pehlivan, R. Atalay, E. Erkin, Y. Karsavuran, S. Tezcan and S. Aksoy**, 2003. Faunistic studies on Miridae (Heteroptera) of Western Black Sea, Central Anatolia and Mediterranean regions of Turkey. *The Scientific and Technical Research Council of Turkey*, Project Numbers: TOAG/336 & 502, 2003, Izmir.
49. **Lu, Y. and K. Wu**, 2011. Mirid Bugs in China: Pest Status and Management Strategies. *Outlooks on Pest Management*, 22 (6), 248-252.

50. **McPherson, JE. and RM. McPherson**, 2000. Stink bugs of economic importance in America North of Mexico. CRC Press, Florida, USA, 2000, pp. 253.
51. **Mirab-balou, M., M. Khanjani and M. Zolfaghari**, 2007. The preliminary study of true bugs (Hemiptera: Heteroptera) fauna in the alfalfa field of Medan province (Western Iran). *Pak. Entomol.*, 29(1).
52. **Mirab-balou, M. and R. Radjabi**, 2013. Lygus rugulipennis Poppius (Hemiptera: Miridae): A key pest on alfalfa (*Medicago sativa L.*) in west of Iran and checklist of the insect pests. *Persian Gulf Crop Protect.*, 2: 57-66.
53. **Mirab-balou, M., Gh.R. Rasoulian, M. Khanjani and Q. Sabahi**, 2008. The preliminary study of true bugs (Hemiptera: Heteroptera) fauna in the alfalfa field of Hamedan province (Western Iran). *Pak. Entomol.*, 30(1), 55-60.
54. **Mirab-balou, M. and C. Xue-xin**, 2009. Lygus rugulipennis Poppius (Hemiptera: Heteroptera: Miridae): a key pest on alfalfa (*Medicago sativa L.*) in Western Iran. 5th European Hemiptera Congress, 31 August–4 September 2009, Velence, Hungary – Abstracts, pp. 32.
55. **May, W. E., J. J. Soroka, H. A. Loeppky and D. C. Murrell**, 2003. The effects of trichlorfon and deltamethrin on alfalfa plant bug and lygus bug (Heteroptera: Miridae) populations in alfalfa grown in Canada. *Crop Prot.*, 22: 883–889.
56. **Mueller, S.C., C.G. Summers and B. Peter**, 2005. Composition of Lygus species found in selected agronomic crops and weeds in the San Joaquin valley, California. *Southwestern Entomologist*, 30(2), 121-127.
57. **Nagalingam, T. and N.J. Holliday**, 2010. Plant bugs (Miridae) on edible beans cumulative report of research activities and results 2008 and 2009 to the Manitoba pulse growers association Ins. 1-16. <http://www.manitobapulse.ca/wp-content/uploads/Edible-bean Holliday lygus-bugs year-1-and-year-2-report.pdf>
58. **Önder, F., E. Pehlivan, R. Atalay, E. Erkin, Y. Karsavuran, S. Tezcan and E.S. Akso**, 1999. Faunistic studies on Lygaeidae (Heteroptera) of Western Black Sea, Central Anatolia and Mediterranean region of Turkey. Project Numbers: TOAG/336 & 502 (The Scientific and Technical Research Council of Turkey), zmir, pp. 58.
59. **Pan, H, B. Liu, Y. Lu and N. Desneux**, 2014. Life Table Parameters of Three Mirid Bug (*Adelphocoris*) Species (Hemiptera: Miridae) under Contrasted Relative Humidity Regimes. *PLoS ONE* 9(12): e115878, 2014.
doi:10.1371/journal.pone.0115878
60. **Panizzi, A. R.**, 1997. Wild hosts of Pentatomids. Ecological Significance and Role in Their Pest Status on Crops. *Annual Review of Entomology*, vol. 42, 99-122.
61. **Panizzi, AR, JE. McPherson, DG. James, M. Javahery and RM. McPherson**, 2000. Economic importance of stink bugs (Pentatomidae), pp.421–474 In: Schaefer CW, Panizzi AR [eds.], *Heteroptera of Economic Importance*, CRC Press, Boca Raton, FL., USA, 2000, pp. 828.
62. **Panizzi, AR**, 2002. Stink bugs on soybean in Northeastern Brazil and new record on the southern green stink bug, *Nezara viridula* (L.) (Heteroptera: Pentatomidae). *Neotropical Entomology*, 31: 331-332.
63. **Panizzi, AR. and JRP. Parra**, 2012. Insect bioecology and nutrition for integrated pest management (IPM), pp. 687-704. In A. R. Panizzi and J. R. P. Parra. *Insect Bioecology and Nutrition for integrated Pest Management*. CRC Press, New York, USA, 2012, pp. 732.
64. **Pansa, M.G., L. Guidone and L. Tavella**, 2012. Distribution and abundance of nymphal parasitoids of *Lygus rugulipennis* and *Adelphocoris lineolatus* in northwestern Italy. *Bulletin of Insectology*, 65 (1), 81-87. ISSN 1721-8861

65. **Pickett, C.H., S.L. Swezey, D.J. Nieto, J.A. Bryer, M. Erlandson, H. Goulet and M.D. Schwartz**, 2009. Colonization and Establishment of *Peristenus. relictus* (Hymenoptera: Braconidae) for Control of *Lygus* spp. (Heteroptera: Miridae) in Strawberries on the California Central Coast. *Biocontrol Control*, 49: 27-37.
66. **Pirgozlieva, E. and B. Dochkova**, 1996. Plant bugs - important pests in alfalfa. *Agrokompas*, 5: 24 (Bg).
67. **Pons, X., E. Nú ez, B. Lumbierres and R. Albajes**, 2005. Epigaeal aphidophagous predators and the role of alfalfa as a reservoir of aphid predators for arable crops. *Eur. J. Entomol.*, 102: 519-525.
68. **Popova, V.**, 1966. Studies on the Heteroptera in the alfalfa biocoenoses in Plovdiv region. *Plant Science*, III (7), 49-57 (Bg).
69. **Proti , L**, 2011. Heteroptera. Prirodna ki muzej u Beogradu. *Posebna izdanja*, knjiga 43, Beograd. ISBN 978-86-82145-36-3, pp. 259 (Sr).
70. **Rämert, B., S. Hellqvist and M.K. Petersen**, 2005. A survey of *Lygus* parasitoids in Sweden. *Biocontrol Science And Technology*, 15(4), 411-426.
71. **Razmjoo, M**, 2012. A Faunistic survey of Hemiptera-Heteroptera found in Isfahan hay alfalfa. *Journal of Research in Agricultural Science*, 8(1), 89-92.
72. **Rosenheim, J.A., R.E. Gomez and E.F. Thatcher**, 2004. Omnivore or herbivore? Field observations of foraging *Lygus hesperus* (Hemiptera: Miridae). *Environmental Entomology*, 33(5), 1362-1370.
73. **Schreiber, A.S**, 2009. Alfalfa and Alfalfa Seed Production. In The Washington Ag. Project. Retrieved from World Wide Web on January 12, 2009.
74. **Schuh, R. T. and J. A. Slater**, 1995. True bugs of the world (Hemiptera: Heteroptera) classification and natural history. Cornell Unive. Press, Ithaca, NY and London, pp.336.
75. **Scudder, GG. and RG. Foottit**, 2006. Alien true bugs (Hemiptera: Heteroptera) in Canada: Composition and adaptation. *Can. Entomol.*, 138: 24-51.
76. **Sekuli , R., S. Kati , . Karagi and T. Kereši**, 2005. Biljne stenice šteto ine lucerke i deteline. Biljni lekar, Poljoprivredni fakultet, Departman za zaštitu bilja i životne sredine „Dr Pavle Vukasovi ”, Novi Sad, 33(5), 517-526.
77. **Shackel, K.A., M. de la P. Celorio-Mancera, H. Ahmadi, L.C. Greve, L.R. Teuber, E.A. Backus and J.M. Labavitch**, 2005. Micro-injection of *Lygus* salivary gland proteins to simulate feeding damage in alfalfa and cotton flowers. *Archives of insect biochemistry and physiology*, 58(2), 69-83.
78. **Smaniotto, L. F. and A.R. Panizzi**, 2015. Interactions of selected species of stink bugs (Hemiptera: Pentatomidae) from Leguminous crops with plants in the Neotropics. *Florida Entomologist*, 8(1), 7-17.
79. **Soroka, J. and J. Otani**, 2011. Arthropods of Legume Forage Crops.In Arthropods of Canadian Grasslands (Volume 2): Inhabitants of a Changing Landscape. Edited by K. D. Floate. *Biological Survey of Canada*, pp. 239-264. © 2011 Biological Survey of Canada. ISBN 978-0-9689321-5-5doi:10.3752/9780968932155.ch10
80. **Sweet, M.H.**, 2000. Seed and Chinch Bugs Lygaeoidea. In: Schaefer C. W., A. R. Panizzi. 2000. Heteroptera of Economic Importance. CRC Press, pp. 856.
81. **Swezey, S.L., D.J. Nieto and J.A. Bryer**, 2007. Control of western tarnished plant bug *Lygus hesperus* Knight (Hemiptera: Miridae) in California organic strawberries using alfalfa trap crops and tractor-mounted vacuums center for agroecology and sustainable food systems. *Environ. Entomol.*, 36(6), 1457-1465.
82. **Tong, Y-J., Y-H. Lu and K-M. Wu**, 2011. Predation of *Geocoris pallidipennis* on *Adelphocoris lineolatus*. *Chinese Journal of Applied Entomology*, 1: 1245-1251.

83. **Volpi, L.N. and M. C. Coscarón**, 2011. Catalog of Nabidae (Hemiptera: Heteroptera) for the Neotropical Region. *Zootaxa*, 2513: 50-68.
84. **Wheeler, A.G.**, 2001. Biology of the Plant Bugs (Hemiptera: Miridae): Pests, Predators, Opportunists. Cornell University Press, Ithaca, New York.
85. **Wheeler, A.G.**, 2012. Studies on the arthropod fauna of alfalfa: VI. Plant bugs (Miridae). *The Canadian Entomologist*, 106 (12), 1267-1275.
86. **Williams, L., J.L. Blackmer, C. Rodriguez-Saona and S. Zhu**, 2010. Plant volatiles influence electrophysiological and behavioral responses of *Lygus hesperus*. *J Chem Ecol*, 36: 467-478.
87. **Wu, U.Y., K.W. Kongming and Y. Guo**, 2009. Comparative study of temperature-dependent life histories of three economically important *Adelphocoris* spp. *Physiological Entomology*, 34: 318-324. doi:10.1111/j.1365-3032.2009.00692.x
88. **Yosifov, M.**, 1962. Quantitative and qualitative studies of entomofauna of lucerne fields with regard to insects of the order Heteroptera, *Sitona* spp. Proceedings of the Zoological Institute and Museum, vol. XI (Bg).
89. **Young, OP**, 1986. Host plants of the tarnished plant bug, *Lygus lineolaris* (Heteroptera: Miridae). *Annals of the Entomological Society of America*, 79: 747-762.
90. **Zhang, T., X. Mei, L. Zhang, K. Wu and J. Ning**, 2015. Identification of female sex pheromone of a plant bug, *Adelphocoris fasciaticollis* Reuter (Hemiptera: Miridae). *J. Appl. Entomol.*, 139: 87-93.
91. **Zink, A.G. and J.A. Rosenheim**, 2004. State-dependent sampling bias in insects: implications for monitoring western tarnished plant bugs. *Entomol. Exp. Appl.*, 113: 117-123.

Sorghum sudanense Piper Stapf Sorgum bicolor L.

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Response of *Sorghum sudanense* Piper Stapf and *Sorgum bicolor* L. after treatment with plant growth regulators with retardant activity

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SUMMARY

| | | | | |
|-----------------------|---------------------|-------------------|---|--|
| (| | | - | In order to establish the biological influence of the plant growth regulators (with retardant activity) CCC 750 SL (750 g/l chlormequat chloride) and Toprex 375 SC (250 g/l difenoconazole + 125 g/l paclobutrazole) in the experimental field of the Institute of Forage Crops, Pleven during the period 2014-2016 was conducted field experiences with <i>Sorghum sudanense</i> Piper. Stapf. and <i>Sorghum bicolor</i> L. |
| 125 g/l | |) | - | |
| 375 | (250 g/l |) | + | |
| 60,120 | 180 ml/da | 750 | - | |
| 25, 50 | 75 ml/da, | 375 | - | |
| | BBCH-13-15 | | - | |
| | (1) | | - | |
| | <i>S. sudanense</i> | <i>S. bicolor</i> | | |
| bicolor. | | | | |
| 750 | | 60 | | |
| ml/da | | (=0.05) | | |
| | | | | |
| <i>S. sudanense</i> , | | <i>S. bicolor</i> | | |
| | | . | | |
| 120 | 180 ml/da | | - | |
| | | | - | |
| <i>S. sudanense</i> | <i>S. bicolor</i> , | | | |

were statistically significant at the highest applied dose ($P=0.05$).

CCC 750 SL did not have a depressant effect on the germination and initial development of *S. sudanense* and *S. bicolor* with excluding for the highest dose – GI ranging from 87.7 to 123.0. Treatment with the Toprex 375 SC had a strong inhibitory effect on the initial development of test plants – GI ranges from 20.0 to 79.7 at all doses.

Key words: *Sorghum sudanense* Piper. Stapf., *Sorghum bicolor* L. selectivity, plant growth regulators

INTRODUCTION

In recent years, research has focused on physiological effects and economic effects following the application of plant growth regulators (with retardant activity) to a number of cereals as a means of increasing the efficiency of seed production (Tams et al., 2004; Eduardo, 2006; Acreche, 2011; Berry and Spink, 2012; Diallo et al., 2015).

Studies have been carried out to determine optimal concentrations for the application of plant growth regulators (with retardant activity) to a number of crops depending on the agro-meteorological conditions (Vardhini and Rao, 2003, Görtz et al., 2012).

Sporadic and contradictory are reports of the use of plant growth regulators (with retardant activity) in sudangrass and sorghum. Singh and Sunder (2012), Pirasteh-Anosheh et al. (2014) find that most plant growth regulators reduce the negative effects of osmotic stress on plant growth. In studies of the GIII et al. (1976), Pando and Srivastava (1985; 1987) found that the use of chlormequat chloride in *Sorghum* species increased the yield and quality of the seeds.

Limited studies in Bulgaria are an argument to establish on the physiological and to assess the economic effect of the

MATERIAL AND METHODS

The studies were conducted during the period 2014-2016 in the experimental field of the Institute for Forage Crops, Pleven on leached black ground under non-irrigated conditions.

The field trial were conducted perpendicularly methods in three replicates with a test area of 5 m².

The following factors are observed:

The following factors are used: Factor A – Species of genus *Sorghum*: a_1 – *Sorghum sudanense* (Piper) Stapf. (Mutant form – M 200/256) and a_2 – *Sorghum bicolor* (L.) Moench (Line 16113); Factor B – plants growth regulators (with retardant activity): b_1 – CCC 750 SL (750 g/l chlormequat chloride) and b_2 – Toprex 375 SC (250 g/l difenoconazole + 125 g/l paclobutrazole). Application: c_1 – control treatment; c_2 – 50%; c_3 – 100% and c_4 – 150% of the registered dose of the plant growth regulators by the manufacturer (Table 1).

1

Table 1. Experimental treatment and characteristics of test plant growth regulators (with retardant activity)

| Species | Genotypes | Active ingredients | Commercial product | / Dose, ml/dia* | | |
|---|-----------------------------------|---|--------------------|-----------------|------|------|
| | | | | 50% | 100% | 150% |
| <i>Sorghum sudanense</i> Piper. Stapf. | 200/256 Mutant form 200/256 | 750 g/l | 750 | 60 | 120 | 180 |
| | | 750 g/l chlormequat chloride | CCC 750 SL | | | |
| | | 250 g/l + 125 g/l | 375 | 25 | 50 | 75 |
| | | 250 g/l difenoconazole + 125 g/l paclobutrazole | Toprex 375 SC | | | |
| <i>Sorghum bicolor</i> L. | 16113 Line 16113 | 750 g/l | 750 | 60 | 120 | 180 |
| | | 750 g/l chlormequat chloride | CCC 750 SL | | | |
| | | 250 g/l + 125 g/l | 375 | 25 | 50 | 75 |
| | | 250 g/l difenoconazole + 125 g/l paclobutrazole | Toprex 375 SC | | | |

*

* Percentage of the registered dose of the commercial product from the manufacturer.

(GI)
Gariglio et al. (2002):

The plant development index (GI) was determined according to the method of Gariglio et al. (2002):

$$GI = \left[\left(\frac{G}{G_0} \right) \cdot \left(\frac{L}{L_0} \right) \right] \cdot 100 \quad (1)$$

G – G_0 –
(%); L – (cm)

; L_0 – , 100%.

$$Y = \arcsin \sqrt{\left(x_{\%} / 100 \right)} \quad (\text{Anant, 1996}).$$

STATGRAPHICS Plus
Windows Ver. 2.1 STATISTICA Ver. 10

where G and G_0 – germinated seeds in each treatment and the control, respectively, %; L – average length (cm) of seedlings in treatment transformed into percentage as against the control treatment; L_0 – average length (cm) of the seedlings in the control treatment taken as 100%.

The percentage of germinated seeds in each treatment was calculated previously transformed $Y = \arcsin \sqrt{\left(x_{\%} / 100 \right)}$ (Anant, 1996). The mathematical-statistical processing of the experimental data was performed with the software product STATGRAPHICS Plus for Windows Ver. 2.1 and STATISTICA Ver. 10.

RESULTS AND DISCUSSION

- Estimating the complex effect of some major meteorological factors, rainfall amount and average 24-hour air temperatures, the studied years can be divided conventionally into two groups: 2014 and 2015 – conditions for the growth and development of the genus *Sorghum* at aridity (I_{ar-DM}) of the growing season (III-IX): 29.6 and 25.41, respectively and 2016 – unfavorable at aridity of 22.3 (Table 2).

(2).

2016<2015<2014,
2014<2015<2016

2014 .)
-1.3 +3.4 °

The total rainfall amount during the period of study can be ranged in the following ascending order: 2016<2015<2014 and in inverse relation 2014<2015<2016 with regard to average 24-hour air temperatures. The agro-meteorological conditions during the years of study (except for 2014) showed temperature deviations from -1.3 to +3.4 ° and stronger variability in the rainfall amount from 12.7 to 314.8%, as

12.7 314.8% - | compared to those for the many-year period (Table 2)
(2).

2.

(2014-2016 .)

Table 2. Meteorological indicators during the years and period average (2014-2016)

| Period of study | / Vegetation period | | | | | | | III – IX, $^{\circ}\text{C}$ Average for III – IX, $^{\circ}\text{C}$ | |
|--|---|-------|-------|-------|-------|------|-------|---|--|
| | a The average monthly temperature of the air, $^{\circ}\text{C}$ | | | | | | | | |
| | III | IV | V | VI | VII | VIII | IX | | |
| 2014 | 9.7 | 14.9 | 16.7 | 20.6 | 23.1 | 23.7 | 17.9 | 18.1 | |
| $\text{Deviation } ^{\circ}$ | 3.3 | 2.9 | -1.0 | -0.6 | -0.3 | 0.8 | -0.4 | 0.0 | |
| 2015 | 6.8 | 12.2 | 18.8 | 20.7 | 25.5 | 24.4 | 20 | 18.3 | |
| $\text{Deviation } ^{\circ}$ | 0.4 | 0.2 | 1.1 | -0.5 | 2.1 | 1.5 | 1.7 | 0.2 | |
| 2016 | 8.5 | 15.4 | 16.4 | 23 | 24.6 | 23.5 | 19.4 | 18.7 | |
| $\text{Deviation } ^{\circ}$ | 2.1 | 3.4 | -1.3 | 1.8 | 1.2 | 0.6 | 1.1 | 0.6 | |
| 50 . (1964 – 2013) Average 50 years (1964–2013) | 6.4 | 12.0 | 17.7 | 21.2 | 23.4 | 22.9 | 18.3 | 18.1 | |
| Period of study | / Vegetation period | | | | | | | III - IX, mm Amount for III- IX, mm | |
| | , mm | | | | | | | | |
| | Monthly rainfall, mm | | | | | | | | |
| 2014 | 39.7 | 32.3 | 83 | 54.3 | 71.8 | 23.9 | 142.6 | 447.6 | |
| e, % / Deviation, % | 111.5 | 66.3 | 132.0 | 85.2 | 116.7 | 52.5 | 314.8 | 123.2 | |
| 2015 | 76.9 | 43.6 | 30.6 | 95.9 | 21.5 | 29.9 | 130.3 | 428.7 | |
| e, % / Deviation, % | 216.0 | 89.5 | 48.6 | 150.5 | 35.0 | 65.7 | 287.6 | 118.0 | |
| 2016 | 68.4 | 72.5 | 77.2 | 46.1 | 7.8 | 31.2 | 61.8 | 365.0 | |
| e, % / Deviation, % | 192.1 | 148.9 | 122.7 | 72.4 | 12.7 | 68.6 | 136.4 | 100.5 | |
| 50 . (1964 – 2013) Average 50 years (1964–2013) | 35.6 | 48.7 | 62.9 | 63.7 | 61.5 | 45.5 | 45.3 | 363.2 | |
| Period of study | De Martonne | | | | | | | III-IX Average for III – IX | |
| | De Martonne aridity index, $I_{\text{a}}-\text{DM}$ | | | | | | | | |
| | III | IV | V | VI | VII | VIII | IX | | |
| 2014 | 46.8 | 15.6 | 37.3 | 21.3 | 26 | 8.5 | 61.3 | 29.6 | |
| 2015 | 48.9 | 23.6 | 12.8 | 37.5 | 7.3 | 10.4 | 52.1 | 25.4 | |
| 2016 | 49.9 | 34.3 | 35.1 | 16.8 | 2.7 | 11.2 | 25.2 | 22.3 | |
| 50 . (1964 – 2013) Average 50 years (1964–2013) | 26 | 26.6 | 27.3 | 24.5 | 22.1 | 16.6 | 19.2 | 22.7 | |

) 750 - | Plant growth regulators (with retardant activity) CCC 750 SL and Toprex 375 SC applied to *S. sudanense* and *S. bicolor* in growth stage BBCH-13-15 did not have a phytotoxicity effect (score 1) for the study period of the test species (Table 3).
375 S. *bicolor* (1)
sudanense BBCH-13-15 (3).

)

Table 3. Phytotoxicity and crop vigor in *Sorghum* species depending on the type and dose of applied plant growth regulators (with retardant activity) averaged for the period

| Treatments | /Dose, ml/da | 0 DBA | 7 DAA | 14 DAA | 20 DAA | 30 DAA | 45 DAA |
|---------------------------|-----------------|---------------------------------------|----------|--------|--------|--------|--------|
| | | <i>Sorghum sudanense</i> Piper Stapf. | | | | | |
| | | / Phytotoxicity | | | | | |
| /Control | | 1 | 1 | 1 | 1 | 1 | 1 |
| 750 CCC 750 SL | 60 | 1 | 1 | 1 | 1 | 1 | 1 |
| | 120 | 1 | 1 | 1 | 1 | 1 | 1 |
| | 180 | 1 | 1 | 1 | 1 | 1 | 1 |
| /Control | | 1 | 1 | 1 | 1 | 1 | 1 |
| 375 Toprex 375 SC | 25 | 1 | 1 | 1 | 1 | 1 | 1 |
| | 50 | 1 | 1 | 1 | 1 | 1 | 1 |
| | 75 | 1 | 1 | 1 | 1 | 1 | 1 |
| / Crop vigor | | | | | | | |
| /Control | | 100 | 100 | 100 | 100 | 100 | 100 |
| 750 CCC 750 SL | 60 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 120 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 180 | 100 | 100 | 100 | 100 | 100 | 100 |
| /Control | | 100 | 100 | 100 | 100 | 100 | 100 |
| 375 Toprex 375 SC | 25 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 50 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 75 | 100 | 100 | 100 | 100 | 100 | 100 |
| <i>Sorghum bicolor</i> L. | | | | | | | |
| / Phytotoxicity | | | | | | | |
| /Control | | 1 | 1 | 1 | 1 | 1 | 1 |
| 750 CCC 750 SL | 60 | 1 | 1 | 1 | 1 | 1 | 1 |
| | 120 | 1 | 1 | 1 | 1 | 1 | 1 |
| | 180 | 1 | 1 | 1 | 1 | 1 | 1 |
| /Control | | 1 | 1 | 1 | 1 | 1 | 1 |
| 375 Toprex 375 SC | 25 | 1 | 1 | 1 | 1 | 1 | 1 |
| | 50 | 1 | 1 | 1 | 1 | 1 | 1 |
| | 75 | 1 | 1 | 1 | 1 | 1 | 1 |
| / Crop vigor | | | | | | | |
| /Control | | 100 | 100 | 100 | 100 | 100 | 100 |
| 750 CCC 750 SL | 60 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 120 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 180 | 100 | 100 | 100 | 100 | 100 | 100 |
| /Control | | 100 | 100 | 100 | 100 | 100 | 100 |
| 375 Toprex 375 SC | 25 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 50 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 75 | 100 | 100 | 100 | 100 | 100 | 100 |

: WRS (European Weed Research), 1 - ; DBA - ; DAA - ; 100 - ; 0 - ;

Legend: EWRS (European Weed Research), Ball 1 - no damage, and at score 9 - culture is completely destroyed; DBA - day before treatment; DAA - days after application; crop vigor 100 - no damage, and 0 - the culture is completely destroyed

| | | | |
|------------------|--------|-------|---|
| <i>sudanense</i> | (—) | S. | Plant height and degree of lodging (score 1) in <i>S. sudanense</i> after treatment with CCC 750 CL and Toprex 375 SC varied in a narrow range (from 200.4 to 246.7 cm) and did not depend on the factors studied – the type of plant growth regulator and the dose of application, the differences were statistically not significant (Table 4). |
| 750 | 375 | | |
| (cm) | (200.4 | 246.7 | |
| — | , | - | |
| (—) | 4). | | |

4. *Sorghum sudanense Piper Stapf.*

Table 4. Structural elements of productivity in *Sorghum sudanense Piper Stapf.* depending on the type of plant growth regulators (with retardant activity) and the applied doses

| Years | Treatments | / Dose, ml/da | / Indicators | | | | | | |
|-------------------|-------------------|---------------|----------------|--------|--------|--------|----------|---------|--|
| | | | M ^l | SH | LP | WT | WS | NSP | |
| 2014 | /Control | | 1 | 235.1a | 32.4bc | 20.4b | 16.2ab | 887.1ab | |
| | 750 CCC 750 SL | 60 | 1 | 240.7a | 30.1c | 24.5b | 20.9b | 1185.0c | |
| | | 120 | 1 | 228.6a | 30.1b | 17.4a | 15.2a | 994.5bc | |
| | | 180 | 1 | 200.4a | 29.5a | 16.6a | 11.5a | 749.2a | |
| | /Control | | 1 | 235.1a | 32.4b | 20.4b | 16.2b | 887.1b | |
| | 375 Toprex 375 SC | 25 | 1 | 246.7a | 30.2b | 22.0b | 16.4b | 995.5b | |
| | | 50 | 1 | 231.5a | 24.9a | 13.2a | 9.5a | 631.1a | |
| | | 75 | 1 | 222.1a | 25.9a | 12.7a | 9.7a | 630.9a | |
| | /Control | | 1 | 195.8a | 32.4a | 21.8b | 17.3b | 819.6a | |
| | 750 CCC 750 SL | 60 | 1 | 201.4a | 33.1a | 27.0c | 23.3c | 1120.3b | |
| 2015 | | 120 | 1 | 189.3a | 31.2a | 18.1ab | 15.9b | 704.3ab | |
| | | 180 | 1 | 272.9a | 30.9a | 17.4a | 12.1a | 756.5ab | |
| /Control | | 1 | 197.6a | 32.4ab | 20.4b | 16.4ab | 766.9a | | |
| 375 Toprex 375 SC | 25 | 1 | 207.4a | 35.6ab | 26.0c | 19.4ab | 1011.7b | | |
| | 50 | 1 | 192.2a | 30.3a | 16.8ab | 12.1b | 624.53ab | | |
| | 75 | 1 | 182.8a | 31.5a | 15.4a | 11.8a | 461.1a | | |
| /Control | | 1 | 215.5a | 32.4a | 21.1b | 16.8b | 853.4ab | | |
| 750 CCC 750 SL | 60 | 1 | 221.1a | 31.6a | 25.8c | 22.1c | 1152.7b | | |
| | 120 | 1 | 209.0a | 30.7a | 17.8a | 15.6b | 849.4a | | |
| | 180 | 1 | 236.6a | 30.2a | 17.0a | 11.8a | 752.9a | | |
| /Control | | 1 | 216.3a | 32.4a | 20.4bc | 16.3a | 827.0bc | | |
| 375 Toprex 375 SC | 25 | 1 | 227.0a | 32.9a | 24.0c | 17.9a | 1003.6c | | |
| | 50 | 1 | 211.9a | 27.6a | 15.0ab | 10.8b | 627.8ab | | |
| | 75 | 1 | 202.4a | 28.7a | 14.1a | 10.8b | 546.0a | | |

: M^l – (1-9); Vera et al. (2012); SH – , cm; WT – , g; WS – , g; NSP – , g;

Legend: M^l – lodging plants; SH - height of plants, cm; LP - length of panicle, cm; WT - total weight of panicle, g; WS - seed weight, g; NSP - number of seeds from one panicle, num.

S. bicolor

The influence of the plant growth regulators tested on *S. bicolor* on the height of the plants is not one sided.

750 375
 (5).

Growth dynamics of plants treated with CCC 750 CL and Toprex 375 SC depend on dose applied and degree of lodging did not depend on the studied factors (Table 5).

5. *Sorghum bicolor L.*
 ()

Table 5. Structural elements of productivity in *Sorghum bicolor L.* depending on the type of plant growth regulators (with retardant activity) and the applied doses

| Years | Treatments | /Dose, ml/da | / Indicators | | | | |
|------------------------------|------------------|-----------------|----------------|--------|--------|-------|-------|
| | | | M ^l | SH | LP | WT | WS |
| 2014 | /Control | | 1 | 78.0c | 17.6b | 9.9b | 7.8b |
| | CCC 750 SL | 60 | 1 | 78.1c | 17.6b | 6.1a | 4.7a |
| | | 120 | 1 | 73.2b | 14.9a | 6.2a | 4.9a |
| | | 180 | 1 | 69.0a | 14.9a | 6.2a | 4.9a |
| | /Control | | 1 | 78.0b | 17.6c | 9.9a | 7.8a |
| | Toprex 375 SC | 375 | 25 | 85.0c | 15.3ab | 12.7b | 10.7b |
| | | 50 | 1 | 74.2b | 16.0b | 9.8a | 8.6ab |
| | | 75 | 1 | 77.4ab | 14.5a | 9.3a | 7.7a |
| 2016 | /Control | | 1 | 67.9b | 16.4b | 12.4b | 7.7b |
| | CCC 750 SL | 750 | 1 | 67.7b | 16.8b | 6.2a | 4.6a |
| | | 120 | 1 | 63.4a | 14.9a | 6.2a | 4.8a |
| | | 180 | 1 | 59.5a | 14.6a | 6.1a | 4.9a |
| | /Control | | 1 | 66.3a | 17.6b | 11.1a | 8.7a |
| | Toprex 375 SC | 375 | 25 | 70.1b | 15.5ab | 13.6a | 11.5a |
| | | 50 | 1 | 65.0a | 16.1ab | 9.3a | 8.1a |
| | | 75 | 1 | 63.0a | 14.7a | 10.1a | 8.1a |
| Average for the period | /Control | | 1 | 73.0a | 17.0b | 11.2b | 7.8c |
| | CCC 750 SL | 750 | 1 | 72.9a | 17.2b | 6.2a | 4.6b |
| | | 120 | 1 | 68.3a | 14.9a | 6.2a | 4.9b |
| | | 180 | 1 | 64.3a | 14.7a | 6.1a | 4.9a |
| | /Control | | 1 | 72.2a | 17.6d | 10.5a | 8.3a |
| | Toprex 375 SC | 375 | 25 | 77.6a | 15.4b | 13.2b | 11.1b |
| | | 50 | 1 | 69.6a | 16.1c | 9.5a | 8.4a |
| | | 75 | 1 | 70.2a | 14.6a | 9.7a | 7.9a |

: M_l – lodging plants; SH - height of plants, cm; LP - length of panicle, cm; WT - total weight of panicle, g; WS - seed weight, g; NSP - number of seeds from one panicle, num.

With the increasing dose was found to disproportionately reduce the height of the plants, the differences were

| | | | |
|--------------------------------------|-------------------|-----------|---|
| | | (=0.05), | statistically significant against the control variant ($P = 0.05$), but only at the higher doses. An exception to the described dependence was found at the applied lowest dose, where Toprex 375 SC had differences were statistically not significant on the studied indicator. |
| 375 | | | The results obtained with respect to the length of the broom are similar. Increasing the dose of the plant applied growth regulator (with retardant activity) disproportionately reduces the length of panicle, the differences being statistically proven only at the applied highest doses. |
|) | | , | |
| a 4 5 | 750 | | |
| 375 | | S. | On Table 4 5 was present data on the Influence of CCC 750 SL and Toprex 375 SC on some structural elements of the yield of <i>S. sudanense</i> and <i>S. bicolor</i> . |
| <i>sudanense</i> <i>S. bicolor</i> . | | | The structural elements of the yield show that the total weight of the panicle and the weight of the seeds of one panicle g are strongly influenced by the dose of the applied plant growth regulator (with retardant activity) and are not dependent of the type of plant growth regulator. |
| , | | | |
| , g | | (| |
|) | | | |
| (| | | |
|) | | | |
| 0.7 45.5%, | | | With increasing dose of growth regulators (with retardant activity), the studied parameters statistically proven decrease from 0.7 to 45.5%, but only at higher doses. More significant differences are found in the number of seeds of one panicle. |
| ml/da) | 750 | (60 | |
| | (=0.05) | | The lowest applied dose (60 ml/da) of CC 750 SL statistically significant ($P = 0.05$) increased the number of seeds in one panicle in <i>S. sudanense</i> , while <i>S. bicolor</i> produced a depressant effect. |
| <i>sudanense</i> , | <i>S. bicolor</i> | S. | With increasing doses of 120 and 180 ml/da there was a decrease in the number of seeds in one panicle In both species – <i>S. sudanense</i> and <i>S. bicolor</i> , he differences were statistically significant at the highest applied dose ($P = 0.05$) (Tables 4 and 5). |
| 120 180 ml/da | | | |
| <i>sudanense</i> <i>S. bicolor</i> , | | – S. | |
| | | (=0.05) | |
| (4 5). | | | |

Stapf. *Sorghum bicolor* L.

Table 6. Influence of plant growth regulators (with retardant activity) on germination and initial development of *Sorghum sudanense* Piper Stapf. and *Sorghum bicolor* L. depending on the applied doses under laboratory conditions

| Genus | Treatments | /Dose ml/da | / Indicators | | | GI |
|---------------------|----------------------|----------------|------------------|----------------------|----------------------------------|-------|
| | | | Germination % | Length of germ cm | Fresh biomass per seedling, g | |
| <i>S. sudanense</i> | /Control | | 45.0b | 1.0a | 0.04a | 100.0 |
| | 750 CCC 750 SL | 60 | 42.1ab | 1.3a | 0.05a | 123.0 |
| | | 120 | 46.4b | 0.9a | 0.04a | 87.7 |
| | | 180 | 30.0a | 0.8a | 0.05a | 61.5 |
| | /Control | | 45.0c | 1.0a | 0.04b | 100.0 |
| | 375 Toprex 375 SC | 25 | 30.0bc | 0.8a | 0.02a | 59.1 |
| | | 50 | 18.4ab | 0.6a | 0.02a | 38.6 |
| | | 75 | 9.1a | 0.5a | 0.01a | 20.0 |
| <i>S. bicolor</i> | /Control | | 80.9b | 1.0b | 0.05b | 100.0 |
| | 750 CCC 750 SL | 60 | 80.9b | 0.9b | 0.04b | 94.9 |
| | | 120 | 65.3b | 0.6ab | 0.04b | 54.9 |
| | | 180 | 33.2a | 0.3a | 0.02a | 18.5 |
| | /Control | | 80.9a | 1.2b | 0.05b | 100.0 |
| | 375 Toprex 375 SC | 25 | 71.6a | 1.1ab | 0.05b | 79.7 |
| | | 50 | 80.9a | 0.6a | 0.02a | 47.1 |
| | | 75 | 67.2a | 1.0ab | 0.05b | 69.6 |

CONCLUSIONS

| | | | |
|-----------------------|---------------------|---|--|
| P | (| - | |
|) | 750 | | |
| 60,120 180 ml/da | 375 | | |
| 25, 50 75 ml/da, | | | |
| BBCH-13-15 | | | |
| (1) | - | | |
| <i>S. sudanense</i> | <i>S. bicolor</i> | | |
| 750 | 60 | | |
| ml/da | (=0.05) | | |
| <i>S. sudanense</i> , | <i>S. bicolor</i> | | |
| | . | | |
| 120 180 ml/da | | | |
| - <i>S. sudanense</i> | <i>S. bicolor</i> , | - | |
| - | | - | |
| (=0.05). | 750 | - | |
| | | - | |

Plant growth regulators (with retardant activity) CCC 750 SL at doses 60, 120 and 180 ml/da and Toprex 375 SC at doses – 25, 50 and 75 ml/da applied at the growth stage BBCH-13-15 did not have phytotoxic effect (score 1) of the tested plant species *S. sudanense* and *S. bicolor*.

CC 750 SL applied in dose 60 ml/da statistically significant ($P=0.05$) increased the number of seeds in one panicle in *S. sudanense*, while *S. bicolor* produced a depressant effect. With increasing doses of 120 and 180 ml/da there was a decrease in the number of seeds in one panicle In both species – *S. sudanense* and *S. bicolor*, the differences were statistically significant at the highest applied dose ($P=0.05$).

CCC 750 SL did not have a depressant effect on the germination

| <i>S. bicolor</i> | <i>S. sudanense</i> | | |
|-------------------|-------------------------|---|---|
| | - | | and initial development of <i>S. sudanense</i> and <i>S. bicolor</i> with excluding for the highest dose – GI ranging from 87.7 to 123.0. Treatment with the Toprex 375 SC had a strong inhibitory effect on the initial development of test plants – GI ranges from 20.0 to 79.7 at all doses. |
| 87.7 | - GI 123.0, 375 , | - | |
| | - GI 20.0 79.7 | - | |

/ REFERENCES

1. **Acreche, M. M. and G. A. Slafer**, 2011. Lodging yield penalties as affected by breeding in Mediterranean wheats. *Field Crops Research*, 122, 40-48. <http://dx.doi.org/10.1016/j.fcr.2011.02.004>
2. **Anant, K.**, 1996. Book reviews: Hinkelmann, K and O. Kempthorne, 1994. Design and analysis of experiments. Vol. I: Introduction to experimental design. New York: John Wiley and Sons, Inc. pp. 495.
3. **Berry, P. M. and J. Spink**, 2012. Predicting yield losses caused by lodging in wheat. *Field Crops Research*, 137, 19-26. <http://dx.doi.org/10.1016/j.fcr.2012.07.019>
4. **Diallo, L., Q. Cao, F. Yang, Z. Yang, J. Cui, L. Gang, T. I. M. Dafaalla, M. Diarso and W. Ahmad**, 2015. Seed priming effects of yuhuangjin on spring maize. *The Journal of Animal & Plant Sciences*, 25(3), 747-754.
5. **Eduardo, C.**, 2006. Effect of induced lodging on grain yield and quality of brewing barley. *Crop Breeding and Applied Biotechnology*, 6, 215-221.
6. **Gariglio, N. F., M. Buyatti, R. Pillati, R. D. Gonzales and M. Acosta**, 2002. Use a germination bioassay to test compost maturity of willow (*Salix sp.*) sawdust. *New Zealand Journal of Crop of Horticultural Science*, 30, 135-139.
7. **Gill, A. S., N. D. Mannikar and C. T. Adhichandani**, 1976. Effect of cycocel on growth and seed yield of sorghum bieour. *Indion I. Agron.*, 21(4), 486-487.
8. **Görtz, A., E. C.Oerke, T. Puhl, and U. Steiner**, 2012. Effect of environmental conditions on plant growth regulator activity of fungicidal seed treatments of barley. *Journal of Applied Botany and Food Quality*, 82(1), 60-68.
9. **Hess, M., G.Barralis, H. Bleiholder, L. Buhr, T. Eggers, H. Hack and R. Stauss**, 1997. Use of the extended BBCH scale-general for the descriptions of the growth stages of mono; and dicotyledonous weed species. *Weed Research*, 37(6), 433-441.
10. **Pando, S. B. and G. C. Srivastava**, 1985. Physiological studies on seed set in sunflower IIL Significance of dwarfening the plant size Using growth regulator. *Indian Journal of Plant Physiology*, 28(1), 72-80.
11. **Pando, S. B. and G. C. Srivastava**, 1987. Influence of cycocel on seed yield and oil content in seed of sunflower (*Helianthus annuus L.*). *Indian Journal of Plant Physiology*, 30(3), 305-307.
12. **Pirasteh-Anosheh , H., Y. Emam and Muhammad Ashraf**, 2014. Impact of cycocel on seed germination and growth in some commercial crops under osmotic stress conditions. *Archives of Agronomy and Soil Science*, 60, 91277-1289.
13. **Rajala, A., and P. Peltonen-Sainio**, 2001. Plant Growth Regulator Effects on Spring Cereal Root and Shoot Growth. *Agronomy Journal*, 93, 936-943. doi:10.2134/agronj2001.934936x
14. **Singh, R. and S. Sunder**, 2012. Foot rot and bakanae of rice: an overview. *Rev. Plant Pathol*, 5, 565-604.

15. **Stall, W., S. Locascio, R. Hochmuth.** 1998. Premergence and postmergence weed control in snap beans. *Proc. Fla. State Hort. Soc.*, 102, 329-332.
16. **Tams, A. R., Sacha J. Mooney and P. M. Berry,** 2004. The Effect of Lodging in Cereals on Morphological Properties of the Root-Soil Complex. 3rd Australian New Zealand Soils Conference, 5-9 December 2004, University of Sydney, Australia. Published on CD ROM. www.region.org.au/au/asssi/
17. **Vera, C. L., S. D. Duguid, S. L. Fox, K. Y. Rashid, J. C. P. Dribbenki and F. R. Clarke,** 2012. Short Communication: Comparative effect of lodging on seed yield of flax and wheat. *Canadian Journal of Plant Science*, 92(1), 39-43.
10.4141/cjps2011-031
18. **Vardhini, V. B. and S. R. Rao,** 2003. Amelioration of osmotic stress by brassinosteroids on seed germination and seedling growth of three varieties of sorghum. *Plant Growth Regulation*, 41(1), 25-31.

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500

Sorghum

1*, 1,
2

1 , 5800
2 , 5200 ,

*E-mail: plserafimov@abv.bg

Influence of herbicides Wing-P, Stomp Aqua and Gardoprim Plus Gold 500 SC on seed germination and initial development of the species of the genus *Sorghum*

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SUMMARY

During the period 2016-2017, at the Institute of Forage Crops - Pleven was studied sensitivity of the ten genotypes of the genus *Sorghum* to herbicides Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin), Stomp Aqua (455 g/l pendimethalin) and Gardoprim Plus Gold 500 SC (312.5 g/l S-metolaclor + 187.5 g/l terbutilazin) under laboratory conditions.

Under the test conditions herbicides and application doses have a strong negative effect on seed germination and growth of seedlings in species of the genus *Sorghum*.

It was found that Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin) and Gardoprim Plus Gold

Sorghum.

- , - (212.5 g/l
250 g/l)
500 (312.5)

| | | | |
|---------------------|---|------|---|
| g/l S-) 10.5 | + 187.5 g/l 100%) | (IR) | 500 SC (312.5 g/l S-metolaclor + 187.5 g/l terbutilazin) have an inhibitory effect (IR from 10.5 to 100%) on seed germination for all tested species of the genus <i>Sorghum</i> . Stomp Aqua (455 g/l pendimethalin) at all applied concentrations had no statistically significant ($P = 0.05$) inhibition effect on germination of genotypes Kazitachi, M300 / 43, GL15A, Mi16N and G16. |
| Mi16N G16. | Kazitachi, M300/43, GL15A, (= 0.05) | - | The application in different concentrations of Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin), Stomp Aqua (455 g/l pendimethalin) and Gardoprime Plus Gold 500 SC (312.5 g/l S-metolaclor + 187.5 g/l terbutilazin) suppress the growth of seedlings (IR from 50.9 to 100%) in all tested species of the genus <i>Sorghum</i> (<i>Sorghum sudanense</i> (Piper.) Stapf.; <i>Sorghum vulgare</i> var. <i>technicum</i> [Körn.]), and the differences were statistically significant ($P = 0.05$) compared with control treatments. |
| 50.9 100%) | Sorghum (<i>Sorghum sudanense</i> (Piper.) Stapf.; <i>Sorghum vulgare</i> var. <i>technicum</i> [Körn.]), (= 0.05) | - | Key words: Sorghum, seeds, herbicides, germination, selectivity |
| | : Sorghum, , | , | |
| | Sorghum, (-) (Martin et al., 1982; Dimitrova and Tsukov, 1996; Moyer et al., 2003; Dimitrova, 2005). | - | INTRODUCTION Efficacious destruction of weed species is an essential element of agrotechnics in species of the genus <i>Sorghum</i> for weed control, because of their slow rate of early growth stage and initial development (germination - third leaves) (Martin et al., 1982; Dimitrova and Tsukov, 1996; Moyer et al., 2003; Dimitrova, 2005). |
| | (Dimitrova and Tsukov, 1996; Dimitrova, 2005; Hristova, 2005; 2006; 2007). | - | Weed infestation during this period reduces the productive potential and yield decreases (Dimitrova and Tsukov, 1996; Dimitrova, 2005; Hristova, 2005; 2006; 2007). |
| | (Tahir et al., | - | Application of a complex of agronomic techniques can be achieved relatively good results for restricting the degree of weeding to improve the quality of the resulting production (Tahir et al., 2005; Bibi et al., 2012; Kikindonov et al., |

INTRODUCTION

Efficacious destruction of weed species is an essential element of agrotechnics in species of the genus *Sorghum* for weed control, because of their slow rate of early growth stage and initial development (germination - third leaves) (Martin et al., 1982; Dimitrova and Tsukov, 1996; Moyer et al., 2003; Dimitrova, 2005).

Weed infestation during this period reduces the productive potential and yield decreases (Dimitrova and Tsukov, 1996; Dimitrova, 2005; Hristova, 2005; 2006; 2007).

Application of a complex of agronomic techniques can be achieved relatively good results for restricting the degree of weeding to improve the quality of the resulting production (Tahir et al., 2005; Bibi et al., 2012; Kikindonov et al.,

2005; Bibi et al., 2012; Kikindonov et al., 2013a, 2013b 2013c).

Sorghum

(Pannaccie, et al., 2010; Yu et al., 2015),

(Enchev and Georgieva-Andreeva, 2013; Marinov-Serafimov and Golubinova, 2015a; 2015b).

- (212.5 g/l
g/l),
500 (312.5 g/l S-
g/l)

- 250
(455 g/l
+ 187.5

Sorghum

(,
(,
(1).

2013a; 2013b and 2013c).

Tolerance to herbicides in species and varieties of the genus *Sorghum* was partially studied globally (Pannaccie et al., 2010; Yu et al., 2015) and in us such studies are extremely limited (Enchev and Georgieva-Andreeva, 2013; Marinov-Serafimov and Golubinova, 2015a; 2015b).

The objective of this study was to determine influence of the herbicides Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin), Stomp Aqua (455 g/l pendimethalin) and Gardoprim Plus Gold 500 SC (312.5 g/l S-metolachlor + 187.5 g/l terbutilazin) on seeds germination on species of the genus *Sorghum* and detection of genotypes with low sensitivity to these herbicides for inclusion as components in future breeding programs .

MATERIAL AND METHODS

The study was conducted during the period 2016-2017 in the laboratory of the Institute of Forage Crops in Pleven.

Collection and Preparation of Plant Material. Used seeds of genotypes of the genus *Sorghum* (varieties mutant forms and local varieties), created by different breeding methods (Table 1).

1.
Sorghum
Table 1. Studied species of the genus *Sorghum*

| | / Species | | Method of creation |
|-----|--|-------------------|--------------------|
| | / Scientific name | / Genotype | |
| 1. | <i>Sorghum sudanense</i> (Piper.) Stapf. | -1 / Endje-1 | / Variety |
| 2. | <i>Sorghum sudanense</i> (Piper.) Stapf. | / Kazitachi | / Variety |
| 3. | <i>Sorghum sudanense</i> (Piper.) Stapf. | 9 / Voronejkaya 9 | / Variety |
| 4. | <i>Sorghum sudanense</i> (Piper.) Stapf. | / Vercors | / Variety |
| 5. | <i>Sorghum sudanense</i> (Piper.) Stapf. | M300/43 | / Mutant form |
| 6. | <i>Sorghum sudanense</i> (Piper.) Stapf. | M200/86 | / Mutant form |
| 7. | <i>Sorghum vulgare</i> var. <i>technicum</i> [Körn.] | GL15A | / Local variety |
| 8. | <i>Sorghum vulgare</i> var. <i>technicum</i> [Körn.] | Mi16N | / Local variety |
| 9. | <i>Sorghum vulgare</i> var. <i>technicum</i> [Körn.] | S14 | / Local variety |
| 10. | <i>Sorghum vulgare</i> var. <i>technicum</i> [Körn.] | G16 | / Local variety |

Sorghum) 100 (90 mm
) 5 ml ,
 : -
Sorghum: 1 – *S. sudanense* – Endje-1;
 2 – *S. sudanense* – Kazitachi; 3 – *S. sudanense* – 9; 4 – *S. sudanense* – Vercors; 5 – *S. sudanense* – M300/43; 6 – *S. sudanense* – M200/86;
 7 – *S. vulgare* var. *technicum* – GL15A;
 8 – *S. vulgare* var. *technicum* – Mi16N;
 9 – *S. vulgare* var. *technicum* – S14
 10 – *S. vulgare* var. *technicum* – G16.
 B – : b₁ –
 - (212.5 g/l) 250
 g/l); b₂ – (455
 g/l) b₃ –
 500 (312.5 g/l S-
 + 187.5 g/l).
 :
 1 – 25%; 2 – 50% 3 – 100%
 (0.3, 0.5 1.0%
).
 :
 22 ± 2⁰
 : , %
 , mm
 (IR)
 (1) (Ahn
 and Chung, 2000).

Bioassay Techniques. In order to assess the effect of the tested herbicides on the seed germination and initial development of species of genus Sorghum, 100 seeds of each genotypes were placed in Petri dishes, 90 mm in diameter between filter paper, moistened with 5 ml of a solution prepared according to the study the factors B and C.

They were studied following factors: *Factor A* – species of the genus Sorghum: 1 – *S. sudanense* – Endje-1; 2 – *S. sudanense* – Kazitachi; 3 – *S. sudanense* – Voronejkaja 9; 4 – *S. sudanense* – Vercors; 5 – *S. sudanense* – M300/43; 6 – *S. sudanense* – M200/86; 7 – *S. vulgare* var. *technicum* – GL15A; 8 – *S. vulgare* var. *technicum* – Mi16N; 9 – *S. vulgare* var. *technicum* – S14 and 10 – *S. vulgare* var. *technicum* – G16.

Factor B – applied herbicides: b₁ – Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin); b₂ – Stomp Aqua (455 g/l pendimethalin) and b₃ – Gardoprim Plus Gold 500 SC (312.5 g/l S-metolaclor + 187.5 g/l terbutilazin)

Factor – application rates: 1 – 25%; 2 – 50% 3 – 100% of the dose determined by the manufacturer presented hereinafter in the text as equivalents of 0.3, 0.5 and 1.0% solution, respectively). Distilled water was used as a control. Each variant had eight replications. The samples were then placed in a thermostat operated device at a temperature of 22 ± 2⁰ for seven days.

The following characteristics were determined: Percentage of germinated seeds (%); seedling length, mm for all treatments of the study.

Inhibition rate (IR) was determined by the Equation (1) (Ahn and Chung, 2000).

$$IR = \left[\frac{C - T}{C} \right] \cdot 100 \quad (1)$$

C –

where C – studied parameters in each control treatments; – studied parameters in each treatment

(*TI*) e
Tahseen and Jagannath (2015) (2).

$$TI = \frac{LS_{TR}}{LS_{CT} \cdot 100} \quad (2)$$

LS_{TR} – , mm; LS_{CT} – , mm.
 \arcsin - where LS_{TR} – Longest of seedlings in each treatment, mm; LS_{CT} – Longest of seedlings in each control treatment, mm;
The percentage of seed germination was calculated after preliminary \arcsin - transformation following the formula

$$Y = \arcsin \sqrt{\left(\frac{X}{100} \right)}$$

Hinkelmann and Kempthorne (1994), LD₅₀, Hamilton et al. (1977).

Statgraphics Plus for Windows Ver. 2.1 Statistica Ver. 10.

forwarded by Hinkelmann and Kempthorne (1994), and to induce half-maximal inhibition of growth (LD₅₀) according to Hamilton et al. (1977). The collected data were analyzed using the software Statgraphics Plus for Windows Ver. 2.1 and Statistica Ver. 10.

(- , 500)
100% (IR 5,1)
Sorghum (2).

P=0.05.
Vercors GL15 25 50% (,
)
P = 0.05.

RESULTS AND DISCUSSION

The herbicides (Wing-P, Stomp Aqua and Gardoprim Plus Gold 500 SC) showed an inhibitory effect on the seed germination in all tested species of the genus *Sorghum* (Table 2).

With the increase concentration of the herbicides – Wing-P and Gardoprim Plus Gold 500 SC the decreased disproportionately germinated seed percentage in all treatments, as compared to the control treatment, the differences being statistically significantly smaller at P=0.05. An exception was found for genotypes Vercors and GL15A after treatment with the herbicide Wing-P at doses of 25 and 50% (depending on the dose specified by the manufacturer), where the differences are statistically no significant at P=0.05.

2.

*Sorghum***Table 2. Effect of tested herbicides on seed germination and seedling growth on the genotypes of the genus *Sorghum***

| Genotypes | / Concentration, % | | | | | | | | | | | |
|------------------------|--------------------|---------|--|---|-------|-------|--------|------------------|------------------------------|------|------------------|-------|
| | 0.0% | | | 0.3% | | | 0.5% | | | 1.0% | | |
| | GR% | GR% | IR | GR% | IR | GR% | IR | LS _{mm} | LS _{mm} | IR | LS _{mm} | IR |
| / Germination seeds, % | | | / Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin) | | | | | | / Length of the seedling, mm | | | |
| -P (212.5 g/l) | - | 250 g/l |) | Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin) | | | | | | | | |
| Endje-1 | 63.4c | 33.2b | 47.6 | 0.0a | 100 | 0.0a | 100.0a | 102.3c | 20.3b | 80.2 | 0.0a | 100.0 |
| Kazitachi | 71.6b | 39.2a | 45.2 | 26.6a | 62.9 | 26.6a | 62.9 | 115.6b | 30.6a | 73.5 | 22.3a | 80.7 |
| Voronejkaya 9 | 63.4c | 26.6b | 58.1 | 26.6b | 58.1 | 0.0a | 100.0 | 89.4c | 22.8b | 74.5 | 21.9b | 75.5 |
| Vercors | 71.6b | 63.4b | 11.4 | 56.8ab | 20.6 | 33.2a | 53.6 | 101.3b | 24.4a | 75.9 | 22.4a | 77.9 |
| M300/43 | 71.6c | 33.2b | 53.6 | 0.0a | 100 | 0.0a | 100.0 | 158.2b | 13.2a | 91.7 | 0.0a | 100.0 |
| M200/86 | 50.8b | 39.2b | 22.7 | 0.0a | 100 | 0.0a | 100.0 | 88.7b | 12.3a | 86.1 | 0.0a | 100.0 |
| GL15A | 63.4b | 45.0ab | 29.1 | 39.2ab | 38.2 | 18.4a | 70.9 | 109.1c | 30.1b | 72.4 | 12.0a | 89.0 |
| Mi16N | 90.0d | 56.8c | 36.9 | 26.6b | 70.5 | 0.0a | 100.0 | 102.3b | 15.4a | 84.9 | 6.0a | 94.1 |
| S14 | 71.6c | 39.2b | 45.2 | 0.0a | 100.0 | 0.0a | 100.0 | 67.8b | 13.6a | 79.9 | 0.0a | 100.0 |
| G16 | 71.6c | 33.2b | 53.6 | 0.0a | 100.0 | 0.0a | 100.0 | 140.6b | 15.4a | 89.0 | 0.0a | 100.0 |
| /Average | 68.9c | 40.9b | 40.3 | 17.6ab | 75.0 | 7.8a | 88.7 | 107.5b | 19.8a | 80.8 | 8.5a | 91.7 |
| (455 g/l) | | |) / Stomp Aqua (455 g/l pendimethalin) | | | | | | | | | |
| Endje-1 | 63.4b | 63.4b | 0.0 | 63.4b | 0.0 | 0.0a | 100.0 | 102.3d | 42.4c | 58.6 | 21.3b | 79.2 |
| Kazitachi | 71.6a | 63.a | 11.4 | 71.6a | 0.0 | 56.8a | 20.6 | 115.6c | 38.6b | 66.6 | 25.7ab | 77.8 |
| Voronejkaya 9 | 63.4b | 63.4b | 0.0 | 39.2ab | 38.2 | 26.6a | 58.1 | 89.4c | 43.4b | 51.5 | 23.6a | 73.6 |
| Vercors | 71.6c | 71.6c | 0.0 | 33.2b | 53.6 | 0.0a | 100.0 | 101.3c | 25.6b | 74.7 | 12.3ab | 87.9 |
| M300/43 | 71.6a | 63.4a | 11.4 | 56.8a | 20.6 | 50.8a | 29.1 | 158.2c | 33.1b | 79.1 | 21.1ab | 86.7 |
| M200/86 | 50.8b | 50.8b | 0.0 | 45.0b | 11.4 | 0.0a | 100.0 | 88.7b | 22.6a | 74.5 | 14.5a | 83.7 |
| GL15A | 63.4a | 58.7a | 7.5 | 58.7a | 7.5 | 39.2a | 38.2 | 109.1b | 22.5a | 79.4 | 20.3b | 81.4 |
| Mi16N | 90.0a | 90.0a | 0.0 | 90.0a | 0.0 | 71.6a | 20.5 | 102.3c | 49.3b | 51.8 | 32.1b | 68.6 |
| LV-S14 | 71.6b | 56.8ab | 20.6 | 45.0a | 37.1 | 33.2a | 53.6 | 67.8b | 33.3a | 50.9 | 20.2a | 70.2 |
| G16 | 71.6a | 71.6a | 0.0 | 56.8a | 20.6 | 50.8a | 29.1 | 140.6b | 26.1a | 81.4 | 22.3a | 84.1 |
| /Average | 68.9b | 65.3b | 5.1 | 56.0ab | 18.9 | 32.9a | 54.9 | 107.5c | 33.7b | 66.8 | 21.3ab | 79.3 |
| 500 (312.5 g/l S- | | | + 187.5 g/l)/ Gardoprime Plus Gold 500 SC (312.5 g/l S-metolaclor + 187.5 g/l terbutilazin) | | | | | | | | | |
| Endje-1 | 63.4b | 56.8b | 10.5 | 26.6a | 58.1 | 18.4a | 70.9 | 102.3c | 28.3b | 72.3 | 25.6ab | 75.0 |
| Kazitachi | 71.6c | 50.8bc | 29.1 | 45.0b | 37.1 | 0.0a | 100.0 | 115.6c | 25.6b | 77.9 | 18.4b | 84.1 |
| Voronejkaya 9 | 63.4b | 39.2ab | 38.2 | 33.2a | 47.6 | 26.6a | 58.1 | 89.4c | 25.3b | 71.7 | 15.7ab | 82.4 |
| Vercors | 71.6c | 33.2b | 53.6 | 26.6b | 62.9 | 0.0a | 100.0 | 101.3b | 15.7a | 84.5 | 6.3a | 93.8 |
| M300/43 | 71.6c | 50.8b | 29.1 | 0.0a | 100.0 | 0.0a | 100.0 | 158.2c | 18.6b | 88.2 | 0.0a | 100.0 |
| M200/86 | 50.8b | 39.2b | 22.7 | 0.0a | 100.0 | 0.0a | 100.0 | 88.7c | 15.6b | 82.4 | 0.0a | 100.0 |
| GL15A | 63.4c | 26.6b | 58.1 | 0.0a | 100.0 | 0.0a | 100.0 | 109.1b | 8.7a | 92.0 | 0.0a | 100.0 |
| Mi16N | 90.0b | 90.0b | 0.0 | 71.6b | 20.5 | 39.2a | 56.4 | 102.3c | 35.2b | 65.6 | 25.6ab | 75.0 |
| S14 | 71.6b | 56.8b | 20.6 | 71.6b | 0.0 | 0.0a | 100.0 | 67.8c | 31.6b | 53.4 | 25.1b | 63.0 |
| G16 | 71.6c | 50.8b | 29.1 | 0.0a | 100.0 | 0.0a | 100.0 | 140.6c | 16.7b | 88.1 | 0.0a | 100.0 |
| /Average | 68.9c | 49.4bc | 29.1 | 27.5ab | 62.6 | 8.4a | 88.5 | 107.5c | 22.1b | 77.6 | 11.7a | 87.3 |

: GR% -

, %; LS_{mm} -

; IR-

, %; a, b, c, d, e -

P=0.05;

Legend: GR% - Germination seeds, %; LS_{mm} – Length of the seedling, mm; IR- Inhibition rate,%; a, b, c, d, e statistically significant differences at P=0.05

Kazitachi, M300/43, GL15A,

Mi16N G16

(, 2).

Sorghum (2).

Practically, the seed germination of genotypes Kazitachi, M300/43, GL15A, Mi16N and G16 was not influenced after treatment with Stomp aqua at all applied concentrations according control treatment (Table 2).

Data from biometric measurements of increase in seedlings length allowed objective assessment of phytotoxic effects of the tested herbicides in the initial growth stage of development of species of the genus *Sorghum* (Table 2). All tested herbicides and applied concentrations, had statistically

(2).

(Tl) (1).
 - (Tl)
 27.7 49.1) -1,
 9, Mi16N S14

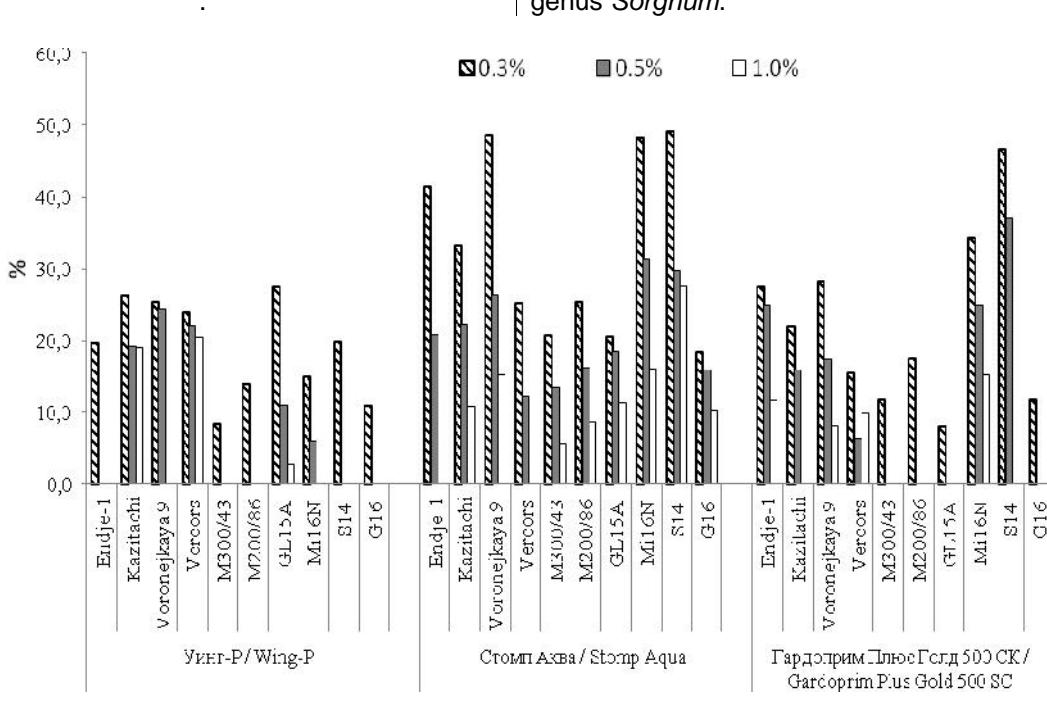


Fig. 1. Tolerance Index (T) of species of the genus *Sorghum* depending on herbicide applications.

Sorghum vulgare var.
technicum [Körn.] -
(T)

The genotypes belonging to the genus *Sorghum vulgare* var. *technicum* [Körn.] have a relatively higher tolerance (T_f) compared to the genotypes of the

Sorghum sudanense (Piper.) Stapf. - genus *Sorghum sudanense* (Piper.)
 (1 1). - Stapf. that can be used as components in development of varieties with improved tolerance to herbicides (Table 1 and Figure 1). The obtained results were analogous when determining LD₅₀ on seed germination of the tested genotypes of the genus *Sorghum* depending on the tested herbicide (Table 3).

LD₅₀ -

Sorghum (3).

3. (LD₅₀)

Sorghum

Table 3. Lethal concentration LD₅₀ of tested herbicides in species of the genus Sorghum

| Genotypes | / Herbicides | | | | | |
|---------------|------------------|----------------------|--------------------------|----------------------|--------------------------------|----------------------|
| | -P / Wing-P | | / Stomp Aqua | | 500 Gardoprim Plus Gold 500 SC | |
| | LD ₅₀ | Cl _{P=0.05} | LD ₅₀ | Cl _{P=0.05} | LD ₅₀ | Cl _{P=0.05} |
| Endje-1 | 103.30 | 90.7 ÷ 117.64 | 236.79 | 228.89 ÷ 244.96 | 195.43 | 161.86 ÷ 235.96 |
| Kazitachi | 118.92 | 85.78 ÷ 164.87 | | >79.2 | 216.34 | 183.95 ÷ 254.45 |
| Voronejkaya 9 | | >57.1 | 269.89 | 207.84 ÷ 350.45 | 237.84 | 130.70 ÷ 432.80 |
| Vercors | 366.80 | 305.60 ÷ 440.26 | 170.44 | 158.62 ÷ 183.14 | | >54.2 |
| M300/43 | | >54.2 | | >70.8 | 124.76 | 116.61 ÷ 133.48 |
| M200/86 | 129.63 | 122.76 ÷ 137.00 | 228.03 | 218.03 ÷ 238.49 | 127.92 | 121.48 ÷ 134.71 |
| LV-GL15A | 242.45 | 200.91 ÷ 292.62 | | >61.9 | | >57.1 |
| LV-Mi16N | 131.95 | 114.44 ÷ 152.14 | | >80.8 | 352.64 | 301.78 ÷ 412.06 |
| LV-S14 | 106.31 | 93.02 ÷ 121.49 | 294.31 (212.28 ÷ 408.04) | | 188.90 | 167.67 ÷ 212.82 |
| LV-G16 | | >54.4 | | >70.8 | 123.11 | 115.49 ÷ 131.24 |
| /Average | 121.64 | 104.14 ÷ 142.08 | 334.53 | 275.40 ÷ 406.55 | 158.57 | 136.60 ÷ 189.09 |

: LD₅₀- , mm; Cl -
 Legend: LD₅₀- lethal doses, mm; Cl- confidence interval

LD₅₀
Sorghum
 103.30 366.80 ml
 - , 195.43 352.64 ml
 500
 - 236.79
 334.53 ml

500
 LD₅₀

Sorghum,

250 g/l - (212.5 g/l)
 500 (312.5 g/l S-

The LD₅₀ values for tested species of the genus *Sorghum* varied from 103.30 to 366.80 ml for herbicide Wing-, from 195.43 to 352.64 ml for Gardoprim Plus Gold 500 SC and the relatively highest from 236.79 to 334.53 ml for Stomp Aqua could be conventionally grouped in the following ascending order: Stomp Aqua Gardoprim Plus Gold 500 SC Wing- .

Differences in the LD₅₀ values of herbicides can be explained by genetic differences between the tested genotypes of the genus *Sorghum*, as comparisons are made under the same conditions.

CONCLUSIONS

Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin) and Gardoprim Plus Gold 500 SC (312.5 g/l S-metolaclor

| | | | | | |
|--|--------------|------------|----------|-------|--|
| 187.5 g/l |) | (IR | 10.5 | 100%) | + 187.5 g/l terbutilazin) have an strong inhibitory effect (IR from 10.5 to 100%) on seed germination for all tested species of the genus <i>Sorghum</i> . Stomp Aqua (455 g/l pendimethalin) at all applied concentrations had no statistically significant ($P=0.05$) inhibitory effect on germination of genotypes Kazitachi, M300/43, GL15A, Mi16N and G16. |
| (455 g/l |) | | | | |
| | | | (=0.05) | | |
| M300/43, GL15A, Mi16N | G16. | | | | |
| 250 g/l | - (212.5 g/l |) | - | | The application in different concentrations of Wing-P (212.5 g/l dimethenamid-p and 250 g/l pendimethalin), Stomp Aqua (455 g/l pendimethalin) and Gardoprime Plus Gold 500 SC (312.5 g/l S-metolachlor + 187.5 g/l terbutilazin) suppress the growth of seedlings (IR from 50.9 to 100%) in all tested species of the genus <i>Sorghum</i> (<i>Sorghum sudanense</i> (Piper.) Stapf.; <i>Sorghum vulgare</i> var. <i>technicum</i> [Körn.]), and the differences were statistically significant ($P=0.05$) compared with control treatments. |
| (455 g/l |) | | | | |
| | 500 | (312.5 g/l | S- | | |
| + 187.5 g/l |) | | | | |
| 50.9 | 100%) | | (IR | | |
| | | | | | |
| Sorghum (<i>Sorghum sudanense</i> (Piper.) Stapf.; <i>Sorghum vulgare</i> var. <i>technicum</i> [Körn.]), | | | | | |
| | | (=0.05) | | | |
| Sorghum <i>vulgare</i> var. <i>technicum</i> | | | | | |
| [Körn.] | - | - | | | |
| | (Tl) | | | | |
| Sorghum <i>sudanense</i> (Piper.) | | | | | |
| Stapf. | | | | | |

/ REFERENCES

1. Ahn, J. K. and I. M. Chung, 2000. Allelopathic potential of rice hulls on germination and seedling growth of barnyard grass. *Agronomy Journal*, 92, 1162-1167.
2. Bibi, A. A. and S. Q. Ali, 2012. Combining ability analysis for green forage associated traits in sorghum-sudangrass hybrids under water stress. *International Journal for Agro Veterinary and Medical Sciences*, 6(2), 115-137.
3. Dimitrova, T. and H. Tsukov, 1996. Study on weeds and their control in sorghum forage, early hybrid "Super sile 20". In: Proceedings of the Union of Scientists - Ruse. Series "Agricultural and veterinary sciences", 5, 57-60 (Bg).
4. Dimitrova, Ts., 2005. Study on weed infestation and their control in sorghum *sudanense* hybrid "Turdan 8". *Plant Science*, 33(8), 67-69 (Bg).
5. Enchev, St. and M. Georgieva-Andreeva, 2013. Study of the influence of some soil herbicides on the composition of weeds in Sudanese crops. *Scientific work, practice, development, innovation*, pp. 4-8 (Bg).
6. Hamilton, M., R. Russo and R. Thurston, 1977. Trimmed Spearman-Karber Method for Estimating Median Lethal Concentrations in Toxicity Bioassays. *Environmental Science and Technology*, 11(77), 14-719.

7. **Hinkelmann, K. and O. Kemphorne**, 1994. Design and analysis of experiments. Vol. I: Introduction to experimental design. New York: John Wiley and Sons. Inc, pp. 495.
8. **Hristova, S.**, 2005. Influence of pendimethalin and nicosulfuron on the growth and reproduction effects of corn for grain. In: Collection of Scientific International Conference Stara Zagora, vol. II, pp. 157-164 (Bg).
9. **Hristova, S.**, 2006. Influence of the Weed Control Agents on the Growth and Reproduction of Kn-613 Hybrids. In: Collection of International Scientific Conference Zagora, vol. I, pp. 200-204 (Bg).
10. **Hristova, S.**, 2007. Influence of some herbicides on the level of entrainment and yield of maize hybrid Kn-611. In: Collection of International Scientific Conference Zagora, vol. I, pp. 366-371 (Bg).
11. **Kikindonov, Tz., G. Kikindonov, K. Slanev and S. Enchev**, 2013a. Dynamics of biomass growth and dry matter accumulation in Sudan grass and Sorghum x Sudan grass hybrids. *Scientific Papers. Series A. Agronomy*, 56, 300-303 (Bg).
12. **Kikindonov, Tz., S. Enchev and K. Slanev**, 2013b. Influence of the variety and sowing rate on the green mass productivity of Sudan grass and Sorghum x Sudan grass hybrids. *Agricultural science and technology*, 5(1), 50-52.
13. **Kikindonov, Tz., G. Kikindonov, K. Slanev and S. Enchev**, 2013c. Analysis of the green mass yield's structure of Sudan grass and Sorghum x Sudan grass hybrids. *Scientific Papers. Series A. Agronomy*, 56, 296-299.
14. **Marinov-Serafimov, Pl. and I. Golubinova**, 2015a. Sudan grass sensitivity to some herbicides: I. Selectivity. *Plant science*, 52(6), 3-12 (Bg).
15. **Marinov-Serafimov, Pl. and I. Golubinova**, 2015b. Sudan grass sensitivity to some herbicides: II. Productivity. *Plant science*, 52(6), 13-20 (Bg).
16. **Martin, A., R. Moomaw and K. Vogel**, 1982. Warm-season grass establishment with atrazine. *Agronomy Journal*, 74, 916-920.
17. **Moyer, J., J. Fritz and J. Higgins**, 2003. Relationships among forage yield and quality factors of hay-type sorghum. Online. *Crops Management*
doi:10.1094/-2003-1209-01-RS.
18. **Pannaccie, S. B. and G. Covarelli**, 2010. Chemical weed control in biomass sorghum [*Sorghum bicolor* (L.) Moench]. *Agricultural Segment*: 1(1) AGS/1516. <http://www.segmentjournals.com>.
19. **Tahir, M. and I. Khan**, 2005. Genetic potential of forage yielding sorghum x sudangrass hybrids for resistance to stem borer (*Chilo partellus*) and shoot fly (*Atherigona soccata*). *Pakistan Entomologist*, 27(1), 57-62.
20. **Tahseen, N. K. and S. Jagannath**, 2015. Assessment of allelopathic efficacy of *Parthenium hysterophorus* L. plant parts on seed germination and seedling growth of *Phaseolus vulgaris* L. *Brazilian Journal of Biological Sciences*, 2(3), 85-90.
21. **Yu, L., L. Van Eerd, I. O'Halloran, P. Sikkema and D. Robinson**, 2015. Response of four spring-seeded cover crops to residues of selected herbicides. *Canadian Journal of Plant Science*, 95: 303-313.