

(*Plagionotus floralis* Pall.) (Coleoptera: Cerambycidae)

Interrelationship between the variety, distance between rows and age of alfalfa plants and the damage from the larvae of alfalfa longhorn beetle (*Plagionotus floralis* Pall.) (Coleoptera: Cerambycidae)

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SUMMARY

(*Medicago sativa* L.)
K ú
Plagionotus floralis (Pallas 1776)
(Coleoptera: Cerambycidae).
2010-2015 .
" " -
: " 6",
" 3" " 1"
12,5 70 cm.
(%)

Alfalfa (*Medicago sativa* L.) is a major grass-fodder crop in Bulgaria. Its root system is the host of a large number of insect pests, among which economically important is the alfalfa longhorn beetle *Plagionotus floralis* (Pallas 1776) (Coleoptera: Cerambycidae). During the period 2010-2015, field experiments with alfalfa were carried out in the experimental field of the IASS "Obrazcov Chiflik" - Rousse, which included three varieties: „Pleven 6”, „Prista 3” and „Mnogolistna 1” at ages one to four years at distance between rows of 12,5 and 70 cm. The number of plants with healthy and damaged roots is determined in the laboratory after their cutting.
The aim of the study is to determine the amount (%) of the damaged plants depending on the alfalfa variety, the plant age and the distance between rows. It

„ 3” „ („ 6” „ 1”) - (12,5 m 70 m),
Plagionotus floralis,
 :

Plagionotus floralis -
 (Sakharov, 1923; Koval et al., 1998)

(Mitruskin, 1940)
 (uki and Eri , 1995).

Popova (1968) 20-30%
 – 60-80%.
 100% *Pl. floralis* -

()
 10 30% Chorbadjiev
 (1930, 1932). 70-80%
 6-7 - Makarov
 (1968).

Nikolova and Kertikova (2008)
 „ 2” (28,6%). „ ” (17,4%)
 -
 62,0%,

was found that alfalfa variety („Pleven 6”, „Prista 3” and „Mnogolistna 1”) and distance between rows (12,5 m and 70 m) factors did not have a significant effect on the relative amount of the damaged plants from the alfalfa longhorn beetle larvae. The amount of damaged plants in all three varieties increases with increasing their age.

Key words: alfalfa, *Plagionotus floralis*, damage, variety, age, distance between rows

INTRODUCTION

Plagionotus floralis is one of the most important insect pests in alfalfa in the areas of Saratov and Krasnodar in Russia (Sakharov, 1923; Koval et al., 1998) and is largely responsible for the dilution of the old alfalfa crops in Romania (Mitruskin, 1940) and Serbia (uki and Eri , 1995).

Similar is the opinion of other authors, according to which older and weedy alfalfa crops are preferred because the flowering plants in them are a source of food for adult insects. In the village of Dulovo (Silistra region) in two-three years old alfalfa Popova (1968) observed 20-30% damage from the alfalfa longhorn beetle, and after the third year – 60-80%. For damage up to 100% of the larvae of *Pl. floralis* in the older alfalfa in the villages of Markovo and Opalchenets (Chirpan district) and for average losses in the attacked areas between 10 and 30% reported also Chorbadjiev (1930, 1932). Information about 70-80% damaged plants in 6-7 - years of alfalfa in Kubrat also gives Makarov (1968).

Nikolova and Kertikova (2008) found that with a low percentage of damage from the larvae of the alfalfa longhorn beetle were the varieties Dara (17,4%) and Prista 2 (28,6%). More strongly attacked are the individual plants grown in the larger food area, in which the attack is on average 62.0%, while the

– 27,3%.
2”

Pl. floralis.

(*Plagionotus floralis* Pall.)

- crop is significantly lower – 27.3%. The "Dara" and "Prista 2" varieties have a low degree of mixed damage from soil pests and may be suitable for plasma tolerance to *Pl. floralis*.

- The aim of this study is to determine the quantity of plants damaged from the larvae of alfalfa longhorn beetle (*Plagionotus floralis* Pall.) in three alfalfa varieties depending on the age and distance between rows of crops.

MATERIAL AND METHODS

The experimental work was carried out during the period 2010-2015 in the experimental field and laboratory of entomology of IASS "Obraztsov Chiflik" - Rousse. The field parcel trials were conducted with three alfalfa varieties: „Pleven 6” (standard), „Prista 3” (one of the most common varieties in production) and „Mnogolistna 1” (multifoliolate type of alfalfa variety) aged one to four years (Table 1).

2010-2015

6” (), „ 3” (: „ 1” () (1).

1.

Plagionotus floralis Pall.

Table 1. Scheme of the field trials to study the effect of the distance between rows and the age of alfalfa varieties on the harmful activity of *Plagionotus floralis* Pall.

Year	/Distance between rows					
	12,5 cm			70 cm		
	1 /1 variant	2 /2 variant	3 /3 variant			
2010	/sowing					
2011	2- /2-year old	/sowing			/sowing	
2012	3- /3-year old	2- /2-year old	/sowing		2- /2-year old	
2013	4- /4-year old	3- /3-year old	2- /2-year old	3- /3-year old		
2014		4- /4-year old	3- /3-year old	4- /4-year old		
2015			4- /4-year old			

cm (12,5)

4

2 m² (1,25 m x 1,6 m).

- In a trial with thick sowing (12.5 cm distance between rows), each variant involves sowing the three varieties in 4 replications. An experimental plot has an area of 2 m² (1,25 m x 1,6 m). The plots are situated in the perpendicular method with randomization (Shanin, 1977;

(Shanin, 1977; Dimova and Marinkov, 1999) and are separated from each other by security strip of 1 m wide (Figure 1).

Dimova and Marinkov, 1999) and are separated from each other by security strip of 1 m wide (Figure 1).

III / replication			IV / V replication		
„Prista 3” 3”	” - 1”	”Pleven 6” 6”	” - 1”	”Prista 3” 3”	”Pleven 6” 6”
”Pleven 6” 6”			”Prista 3” 3”		
I / replication			II / replication		

Fig. 1. Scheme of field trial with 12,5 cm distance between rows (2010-2015)

15-20 m, 0,25 m², 18 cm, 23,8 m² (11,9 m x 2 m), 18 (6), 2 m, 15-20 cm, SPSS 16.0 for Windows P 0,05.

In October-November, of the four replications of each variety during the study period, the plants of 0.25 m² were removed at a depth of 15-20 cm, the roots of all plants were dissected in a laboratory conditions, and the number of plants with healthy and damaged roots was counted.

The trial with wide sowing (70 cm distance between rows) have a total area of 23,8 m² (11,9 m x 2 m) and include 18 rows (6 rows of the three tested varieties arranged in triples) each with a length of 2 linear m. In October-November of each variety were removed plants from 1 linear m at a depth of 15-20 cm in triplicate and dissected in a laboratory conditions. The number of plants with healthy and damaged roots was counted.

Statistical processing of the results was performed using the program SPSS 16.0 for Windows (www.spss.com) at a level of confidence P 0,05.

RESULTS AND DISCUSSION

The obtained results show that there is no proven difference in the quantity of the damaged plants depending on the variety. The degree of damage is increasing from one-year old to four-year

1. („Pleven 6", „Prista 3" and „Mnogolistna 1") (12,5 m and 70 m)

2.

CONCLUSIONS

1. The factors alfalfa variety („Pleven 6", „Prista 3" and „Mnogolistna 1") and distance between rows (12,5 cm and 70 cm) during the study period did not have a significant influence on the relative amount of the damaged plants from the larvae of the alfalfa longhorn beetle.
2. The plant age factor during the study period has a proven effect on the larval damage. The quantity of the damaged plants in all three varieties increases with their age.

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/ REFERENCES

1. **Chorbadjiev, P.**, 1930. Notes on some harmful insects on crop plants in Bulgaria in 1928 and 1929. *Izvestiya na bulgarskoto entomologichno druzhestvo*, book V, 63-106 (Bg).
2. **Chorbadjiev, P.**, 1932. Pests on cultivated plants in Bulgaria in 1928 and 1929 years, *Svedeniya po zemedelioto*, XIII(1-2), 3-63 (Bg).
3. **Dimova, D. and E. Marinkov**, 2005. Experimental design and biometrics. Academic Publishing House of the Agrarian University, Plovdiv (Bg).
4. **uki, D. and P. Eri**, 1995. Lucerka, Novi Sad, 150 (Sr).
5. **Dytham, C.**, 2003. Choosing and Using Statistics. A Biologist'Guide, Second Edition. Blackwell Publishing company, London, pp. 248.
6. **val, ., S. Russkih, . Sinitsina and . Devyatkin**, 1998. Features of ecological protection against phytophagous and use of pollinator insects in the production of alfalfa seeds [in the Krasnodar territory][Agrotechnical and chemical methods for controlling insect pests]. *Production of environmentally safe crop products*, 4, 141-150 (Ru).
7. **Makarov, .**, 1968. Alfalfa longhorn beetle. *Rastitelna zashtita*. 2, 13-18 (Bg).
8. **Mitrjuskin K.**, 1940. *Plagionotus floralis* Pall. and the protection of Lucerne sowings. *Sotsialisticheskoe Zernovoe Khozyaistvo*, 5, 117-120 (Ru).
9. **Nikolova, I. and D. Kertikova**, 2000. Comparative evaluation of Lucerne accessions according to degree of attack by some soil insect pests. *Journal of Mountain Agriculture on the Balkans*, 11(1), 48-59.
10. **Popova, V.**, 1968. Entomofauna of alfalfa. BAN, Sofia (Bg)
11. **Sakharov, N.**, 1923. Report of the Entomological Department for the Years 1916-1923. *Trans. Saratov Agric. Expt. Sia.*, 2 (Ru).
12. **Shanin, Y.**, 1977. Methodology of field trial. Bulgarian Academy of Sciences, Sofia, pp. 96-97 (Bg).

(*Lotus corniculatus* L.)

5800 , .“ “ 89,

Comparative testing of varieties and local populations birdsfoot trefoil (*Lotus corniculatus* L.)

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SUMMARY

2016-2017 .
- .
16
(*Lotus corniculatus* L.),
- .
Leo, -
-
1,
Stamm GH01 Stamm
GH02,
1 2,
- .

The study was conducted in the period 2016-2017 at the Institute for Forage Crops - Pleven. The phenotypic variability in productivity of fresh biomass and the determinant components of 16 genotypes of Birdsfoot trefoil (*Lotus corniculatus* L.), originating from different ecological geographic regions of Europe and Bulgaria are evaluated for the purpose of selection of elite plants and their inclusion in selection programs. They are established unproven differences in quantitative parameters determining the productivity of fresh biomass in variety Leo, but reported lower survival.

The greatest yield of fresh biomass and survival of the tested varieties birdsfoot trefoil is Bulgarian variety used for standard Targovishte 1, and in Austrian populations Stamm GH01 and Stamm GH02, as well as the Bulgarian Local populations 1 and 2 elite plants of which will be included as parental components of synthetic population.

The greatest influence in the formation of yield in genotypes birdsfoot trefoil,

regardless of year of study proves the height of the stems rather than their number.

Key words: *Lotus corniculatus*, breeding, variety, populations

INTRODUCTION

The legumes have important role such as components of mixtures and grassland ecosystem for sustainable agriculture (Naydenova et al., 2013; Vasileva, 2011; Vasileva and Vasilev, 2012; Bozhanska et al., 2016; Vasileva, 2017a). The birdsfoot trefoil (*Lotus corniculatus* L.) is a perennial legume grass species, which which is a valuable component in the establishment of artificial pastures and for forage production (Churkova, 2006b, 2008, 2009, 2012, 2013; Valkov and Chiurazzi, 2016; Vasileva, 2017b). Productivity, quality and resilience of the weeds depend on the choice of species and variety (Naydenova and Mitev, 2015).

The birdsfoot trefoil (*Lotus corniculatus* L.) is widespread in the Mediterranean region (Grant and Niizeki, 1999). The local populations are with great number of agronomically important traits that are well adapted to local ecological conditions. The fact is that acclimated ecotypes provide better production results than the introduced populations and cultivars (Churkova, 2006a; Radi et al., 2013).

Variability for the great number of agronomically traits is very important in order to identify the most productive genotypes, and to introduce them into the further breeding process (Gatari et al., 2013). Understanding the relationships among birdsfoot trefoil morphological, ecogeographic and chemical characteristics may be useful for effective utilizing of different germplasms (Vuckovic et al., 2007). Collection and characterization of germplasm and the original genetic variation represent the basis of plant breeding programs (Prosperi et al., 1999). Therefore, the

regardless of year of study proves the height of the stems rather than their number.

Key words: *Lotus corniculatus*, breeding, variety, populations

INTRODUCTION

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al, 1999).

(Rumbaugh et al., 1988).

- selection of germplasm that will be included in the breeding program is a very sensitive stage of breeding process (Rumbaugh et al., 1988).
- The aim of the investigation was to
- study the potential of varieties and local populations of different origins and
- selection of elite plants based on feeding productivity and durability and inclusion in future breeding programs.

MATERIAL AND METHODS

2016-2017

16

(*Lotus corniculatus*)

(1).

- At the Institute of Forage Crops -
- Pleven in 2016-2017, under field conditions, a 16 variety and populations of birdsfoot trefoil (*Lotus corniculatus* L.) of different origins were investigated (Table 1).

1.

***corniculatus* L.)**

(*Lotus*

Table 1. Name and origin of variety and population from birdsfoot trefoil (*Lotus corniculatus* L.)

	Name	Variety/ Population	Origin
1	1/Targovishte 1	/Variety	/Bulgaria
2	Leo	/Variety	/Canada
3	Taborac	/Variety	/Hungary
4	Gran San Gabriele	/Variety	/Italy
5	Bosna Lotus	/Variety	Bosnia and Herzegovina
6	Natural population	Local population	Bosnia and Herzegovina
7	Stamm GH01	Local population	Austria
8	Stamm GH02	Local population	Austria
9	1 Local population 1	Local population	Central Northern Bulgaria
10	2 Local population 2	Local population	Central Northern Bulgaria
11	3 Local population 3	Local population	Central Northern Bulgaria
12	4 Local population 4	Local population	Central Northern Bulgaria
13	5 Local population 5	Local population	Central Northern Bulgaria
14	8 Local population 8	Local population	Central Northern Bulgaria
15	9 Local population 9	Local population	Central Northern Bulgaria
16	10 Local population 10	Local population	Central Northern Bulgaria

2015 (0.17 m) (0.12 m) 1 L⁻¹.

20-25

, cm;
, cm;

STATGRAPHICS Plus for Windows Version 2.1 Statistica 10.

The varieties and populations of birdsfoot trefoil were sown in pots (0,12 m in diameter and 0,17 m in height) with a volume of 1 L⁻¹ in 2015. In autumn, plants with pronounced productivity of each genotype were planted under field conditions. The only registered variety in Bulgaria, Targovishte 1, is used for control. Each variant consists of five replications.

At the beginning of the growing season, the survival of the plants after the wintering was taken into account. During the vegetation, three cuts were carried out in growth stage beginning of flowering at interval of 20-25 days. In all variants of the experiment for each cutting, the following indicators were taken into account: Height and width of the tuft, cm; Mean height per stem, cm; Fresh and dry biomass, g; Number of stems; Ratio of leaves:stems:flowers in fresh and dry biomass; Relative dry matter ratio in fresh biomass, %.

All experimental data were statistically processed using the STATGRAPHICS Plus for Windows Version 2.1 software and Statistica Ver. 10.

RESULTS AND DISCUSSION

100%

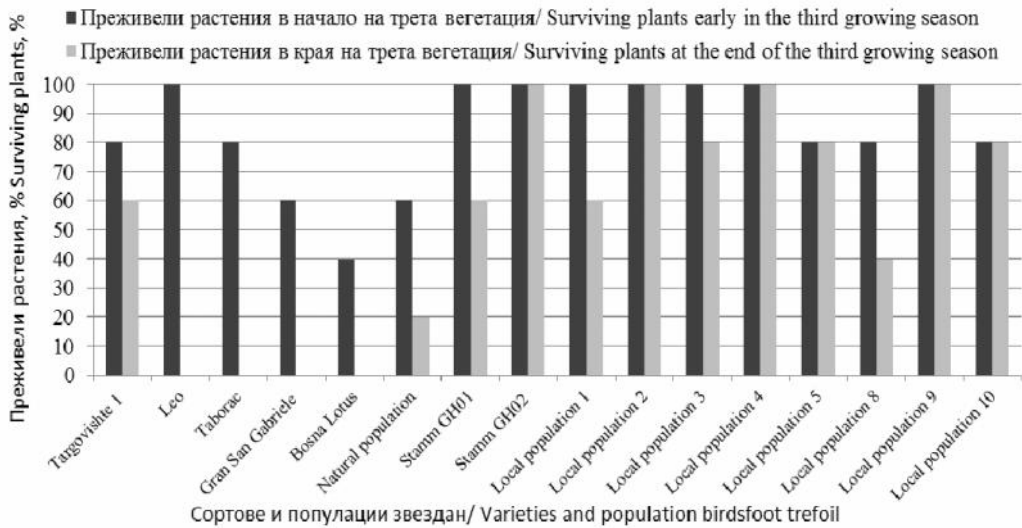
0 100 % (

1). Leo, Taborac, Gran San Gabriele Bosna Lotus

100%

1.

At the beginning of the second vegetation period, 100% survival of the plants was detected in all birdsfoot trefoil genotypes. At the beginning of a third progeny, the survival rate of individual star genotypes varies from 0 to 100% (Figure 1). Plants in intensive varieties Leo, Taborac, Gran San Gabriele and Bosna Lotus died 100% after the first cut. With good resistance and survival by the end of the third growing season there are almost all the populations that are equalizing or have a higher survival rate than the Targovishte 1 variety.



. 1.

Fig. 1. Survival of varieties and populations birdsfoot trefoil at the beginning and end of third vegetation period

(2).
 4 2016 , 2017
 3
 20-25 .
 2016 .
 Taborac, 28,0%
 (48,8 %)
 -
 8,8% ,
 (8,9
 10) -
 , 32,7 59,4
 % -
 1.

The phenotypic variability of the yields of fresh biomass on slopes and on average for plant vegetation is clearly expressed (Table 2). In the tested genotypes birdsfoot trefoil, 4 cuts were made in 2016 and 3 cuts in 2017 at an interval of 20-25 days.

In 2016, in most of the variants of the trial, the intensity of biomass accumulation is maximum in a second cutting. The highest amount of fresh biomass for the vegetation period in 2016 is Taborac variety, which exceeds 28.0% of the control. In this variety, the biomass formed with a maximum in the second cutting (48.8% of the total for vegetation period).

At the end of vegetation, the population of erected habitus - Local population 2 exceeds the control by 8.8%, and in the case of the straining type (Local populations 8, 9 and 10), a lesser amount of fresh biomass for the vegetation period is from 32.7 to 59.4% lower than Targovishte 1 variety.

2.

(2016-2017 .)

Table 2. Fresh biomass in varieties and populations birdsfoot trefoil, by cuts and average for vegetation (for 2016-2017)

Genotypes	I	II	III	IV	Average for vegetation
	I cutting	II cutting	III cutting	IV cutting	
2016					
Targovishte 1	0,221abc	0,373bcd	0,159def	0,055bcde	0,807bcd
Leo	0,198abc	0,376bcd	0,194fg	0,064cde	0,823bcd
Taborac	0,257bcde	0,505cd	0,223g	0,043abc	1,033d
Gran San Gabriele	0,261bcde	0,358bcd	0,133bcde	0,031ab	0,783bcd
Bosna Lotus	0,354def	0,261abc	0,126bcde	0,022a	0,692bc
Natural population	0,258bcde	0,378cd	0,180efg	0,041abc	0,866cd
Stamm GH01	0,346bcde	0,461d	0,140cdef	0,054bcde	0,901cd
Stamm GH02	0,238abcd	0,442d	0,147cdef	0,036	0,863cd
Local population 1	0,256bcde	0,308bcd	0,134bcde	0,072e	0,771bcd
Local population 2	0,367ef	0,357bcd	0,108bcd	0,041abc	0,878cd
Local population 3	0,357def	0,314bcd	0,123bcde	0,043abc	0,765bcd
Local population 4	0,386f	0,272abc	0,096abc	0,038ab	0,792bcd
Local population 5	0,297cdef	0,317bcd	0,122bcde	0,070de	0,646bc
Local population 8	0,112a	0,211ab	0,044a	0,043abc	0,328a
Local population 9	0,148ab	0,285abc	0,078ab	0,032ab	0,543ab
Local population 10	0,112a	0,138a	0,040a	0,037ab	0,261a
/ Average	0,261	0,335	0,128	0,045	
2017					
Targovishte 1	0,487i	0,261e	0,101def		0,849e
Leo	0,141ab	0,000a	0,000a		0,141a
Taborac	0,128ab	0,000a	0,000a		0,128a
Gran San Gabriele	0,102a	0,000a	0,000a		0,102a
Bosna Lotus	0,217bc	0,000a	0,000a		0,217a
Natural population	0,306cde	0,161cd	0,086cde		0,553cd
Stamm GH01	0,294cde	0,349g	0,169h		0,812ef
Stamm GH02	0,320def	0,355g	0,126fg		0,800de
Local population 1	0,343efg	0,140bc	0,060bc		0,543c
Local population 2	0,425hi	0,308f	0,139g		0,872ef
Local population 3	0,641j	0,272e	0,075cd		0,987f
Local population 4	0,581j	0,202d	0,039b		0,823de
Local population 5	0,390fgh	0,290ef	0,117efg		0,796de
Local population 8	0,310de	0,123bc	0,032b		0,464bc
Local population 9	0,391gh	0,101b	0,085cd		0,577c
Local population 10	0,249cd	0,106b	0,041b		0,396b
/ Average	0,333	0,167	0,067		

Stamm GH01 Stamm

GH02

Unlike the second vegetation period, in the third year the largest proportion of the formed biomass for vegetation period is in the first cut. The exception is the Austrian populations Stamm GH01 and Stamm GH02 with pronounced summer productivity and a larger share of the second cutting of the total biomass for vegetation. In these, the biomass accumulation intensity is maximum in second cutting (respectively

43,0 (44,3%), -
 -
 -
 2017 2 5, 35,3 36,4%.
 3 4
 (64,9 70,7%).
 -
 2 3
 2,8 16,3%
 1.
 (3).
 -
 ,
 ,
 ,
 Lotus, Bosna
 21,8%
 .
 Gran
 San Gabriele, Bosna Lotus
 1, 2, 3, 4, 5, Natural
 population Stamm GH02
 ,
 1.
 -
 1,
 2, Stamm GH01 Stamm GH02,
 Stamm GH02.
 -
 Leo Stamm GH01,
 Stamm GH02 1
 -
 1 2,7
 11,5% (P=0.05). 2, 3, 4, 5
 (
 8, 9 10)
 -
 1.
 1 -
 -

43.0 and 44.3% of the total biomass for the vegetation) and the total biomass for vegetation does not differ significantly from the control variant.

Of the Bulgarian populations with pronounced summer productivity in 2017 are Local population 2 and 5 with a share of second cutting, respectively 35,3 and 36,4%. In the Bulgarian populations 3 and 4 there is the largest share of the first cutting (64.9 and 70.7% respectively). With the greatest amount of fresh biomass aggregated for the vegetation period, the Bulgarian local populations 2 and 3 exceed respectively 2,8 and 16,3% Targovishte 1 variety.

The average length of the stems in the varieties and populations birdsfoot trefoil by cuts and by genotype (Table 3). Depending on the number of cuts, there is a tendency for the formation of the longest stems in the second cut, followed by the first, third and fourth cuts. An exception was observed with Bosna Lotus, where the stems formed in the first cutting exceeded by 21.8% the length reached in a second cut.

In the first cutting, Gran San Gabriele, Bosna Lotus and Local populations 1, 2, 3, 4, 5, Natural populations and Stamm GH02 form long stems compared to Targovishte 1 variety. In the second cutting, the trend is maintained for Local population 1, 2, Stamm GH01 and Stamm GH02, and in a third cutting only at Stamm GH02.

On average for the vegetation in Leo varieties and Stamm GH01, Stamm GH02 and 1 stems, the length of the stems proved to exceed the established in Targovishte variety 1 of 2.7 to 11.5% (P = 0.05). Local population 2, 3, 4, 5 and those with semi-prostrate habitus (populations 8, 9 and 10) have statistically proven shorter stems compared to Targovishte 1 variety.

In Targovishte 1 variety the length of the stems in the first cutting is highest

23,2% . 12,8
 Bosna Lotus, Natural population, Stamm
 GH01 2 3 -
 (8,8 28,1 %)
 . -

and decreases in the second with 12,8
 and 23,2% in the third cutting. Unlike the
 control variant, for the populations of
 Bosna Lotus, Natural population, Stamm
 GH01 and No 2 and 3, a greater length of
 the stems in a second cutting (from 8.8 to
 28.1%) was reported compared to the
 first cutting. In the third cutting, all
 populations form the shortest stems
 compared to the previous cuttings.

3.

(2016-2017)

Table 3. Height of the stem in varieties and populations of birdsfoot trefoil, by cuts and average for vegetation (for 2016-2017)

Genotypes	I	II	III	IV	Average for vegetation
	I cut	II cut	III cut	IV cut	
2016					
Targovishte 1	37,05cd	44,12bcd	31,80fgh	17,97ef	32,73d
Leo	38,38def	49,55f	33,00h	15,53c	34,13ef
Taborac	34,83bc	49,13f	32,10gh	15,93cd	33,00de
Gran San Gabriele	40,63fgi	42,30bc	28,20e	13,20b	31,08c
Bosna Lotus	50,71g	41,63b	30,37f	13,70b	34,10ef
Natural population	41,90gh	45,60de	31,00fg	17,80ef	34,08ef
Stamm GH01	34,98bc	52,67g	32,40gh	17,43de	34,37f
Stamm GH02	39,58efg	54,52g	35,37i	16,53cde	36,50g
Local population 1	44,45i	46,43ef	31,60fgh	13,03b	33,88def
Local population 2	42,70hi	47,20ef	22,97cd	12,63b	31,38c
Local population 3	42,40hi	41,22b	21,27c	13,07b	29,49b
Local population 4	39,60efg	37,07a	19,25b	10,67a	26,65a
Local population 5	41,83gh	41,18b	21,37cd	17,20de	30,40bc
Local population 8	25,83a	43,68bcd	16,63a	20,05g	26,55a
Local population 9	38,20de	45,00cde	20,17bc	16,27cd	29,91b
Local population 10	32,82b	36,87a	15,97a	19,07fg	26,18a
/ Average	39,12	44,89	26,47	15,63	
2017					
Targovishte 1	37,63 fgh	32,83 bc	28,90 f		33,12 de
Leo	29,97 c	0,00 a	0,00 a		9,99 b
Taborac	18,53	0,00 a	0,00 a		6,18 a
Gran San Gabriele	24,87 b	0,00 a	0,00 a		8,29 ab
Bosna Lotus	33,57 cdef	0,00 a	0,00 a		11,19 b
Natural population	32,87 cde	40,28 f	28,70 f		33,95 efg
Stamm GH01	35,13 defg	45,00 g	30,90 f		37,01 fgh
Stamm GH02	38,63 ghi	47,53 g	30,53 f		38,90 h
Local population 1	32,17 cde	33,68 bcd	21,43 e		29,09 cde
Local population 2	46,83 j	38,67 ef	30,00 f		38,50 gh
Local population 3	51,57 k	38,93 ef	18,53 cd		36,34 fg
Local population 4	48,90 jk	36,77 def	16,10 c		33,92 ef
Local population 5	32,57 cde	35,43 cde	19,40 de		29,13 c
Local population 8	36,70 efgh	31,50 b	12,65 b		26,95 c
Local population 9	40,5 hi	31,63 b	18,20 cd		30,11 cd
Local population 10	32,27 cd	35,40 cde	20,63 e		29,43 c
/ Average	35,79	27,98	17,25		

2016	2017		
Stamm GH01, Stamm GH02		2	3
	1.		
4).		(
2016	-		
2,			
		80,5	65,0%,
			26,9%
	Leo		
	14,4%		
	1.		
8, 9	10 ()	-
		5	2
			3.
			29,4%
	1.		8, 9
10			
28,4%			22,3

Average for vegetation period in 2016 and 2017, stem lengths in the Stamm GH01, Stamm GH02, and 2 and 3 populations proved to exceed the length at variety Targovishte 1.

The yield generated for vegetation is a sum of the stems formed from one plant through the individual cuttings (Table 4). The number of stems that produce the yield of fresh biomass per plant shows a distinction between studied varieties and local populations starved and the cuttings made during vegetation.

In 2016, with the highest number of stems for vegetation, there is a Local population 2, where in the first and second cuttings statistically proved exceed the control by 80,5 and 65,0% respectively and the average for the vegetation with 26,9% more stems. Leo variety stands out with 14.4% more stems than Targovishte 1. Accordingly, in Local populations 8, 9 and 10 (straining type) the number of stems forming the average yield for the vegetation is the smallest.

Unlike the second vegetation and the lack of survival in varieties, the star determines the low number of stems at the first decay. In the Austrian populations the relatively high number of supernatants averaged for vegetation, as well as for Bulgarian populations 2 and 3. In the local population 5, 29,4% higher number of stems was observed compared to Targovishte 1. In populations 8, 9 and 10 from the tendency to reduce the number of stems in the subsequent slopes is confirmed, and the total for the vegetation is from 22.3 to 28.4% of the control variant.

4.

2 (2016 .)

Table 4. Number of stems per plant in varieties and populations starved in slopes and average for vegetation Year 2 (for 2016)

/ Genotypes	2016				Average for vegetation
	I cut	II cut	III cut	IV cut	
Targovishte 1	62,8bc	118,8abc	181,2bcde	117,2bcd	120,0bcdef
Leo	66,2bcd	139,0bcd	191,4cde	152,6d	137,3def
Taborac	95,75de	119,0abc	227,75e	112,8abcd	121,9bcdef
Gran San Gabriele	63,2bc	150,4cde	136,4abc	90,6abc	110,2abcde
Bosna Lotus	56,75abc	99,4ab	130,2ab	72,4ab	86,9ab
Natural population	76,4bcd	144,6bcd	163,8bcd	111,4abcd	124,1bcdef
Stamm GH01	76,0bcd	173,2de	146,6abcd	111,4abcd	126,8cdef
Stamm GH02	63,4bc	135,2bcd	132,2ab	84,4abc	103,8abcd
Local population 1	78,4bcd	146,8bcd	161,8bcd	100,8abc	122,0bcdef
Local population 2	113,4e	196,0e	169,0bcde	130,6cd	152,3f
Local population 3	70,25bcd	116,2abc	165,2bcd	92,4abc	107,5abcd
Local population 4	71,8bcd	133,4bcd	182,2bcde	105,8abcd	123,3bcdef
Local population 5	85,75cde	179,0de	199,5de	135,8cd	150,0ef
Local population 8	24,5a	77,75 a	98,0a	100,3abc	75,1a
Local population 9	53,6ab	139,0bcd	135,0abc	84,2abc	91,6abc
Local population 10	61,0bc	145,5bcd	88,75a	64,5a	89,9abc
/ Average	70,0	136,0	156,8	104,2	
2017					
Targovishte 1	150,3 def	173,3 ef	142 bcd		155,1 e
Leo	79,8 ab	0,0 a	0,0 a		26,6 a
Taborac	76,8 ab	0,0 a	0,0 a		25,6 a
Gran San Gabriele	63,3 a	0,0 a	0,0 a		21,1 a
Bosna Lotus	94,0 abcd	0,0 a	0,0 a		31,3 a
Natural population	109,3 abcde	112,5 bc	106,0 bcd		109,3 bc
Stamm GH01	141,6 def	144,8 cde	158,0 cd		148,1 cde
Stamm GH02	85,8 abc	103,0 b	108,0 bcd		98,8 b
Local population 1	97,4 abcde	121,0 bcde	80,7 b		99,7 b
Local population 2	145,4 def	138,4 bcde	161,0 d		148,1 de
Local population 3	151,4 ef	143,4 bcde	135,0 bcd		143,1 cde
Local population 4	144,0 def	160,2 de	87,6 b		130,6 bcd
Local population 5	168,0 f	208,5 f	225,0 e		200,5 f
Local population 8	140,8 cdef	133,3 bcde	69,5 ab		114,5 bcd
Local population 9	166,8 f	114,5 bc	80,2 b		120,5 bcd
Local population 10	122,5 bcdef	118,3 bcd	92,3 bc		111,0 bc
/ Average	121,1	104,5	90,3		

5). -

(r=0.703).

(

Correlations between the biomass of a plant and the height and number of stems from one plant are established (Table 5). The greatest influence in the formation of the yield is the height of the stems than the number of stems - regardless of the year of study ($r=0.703$).

5.

(2016-2017 .)

Table 5. Correlation between the parameters determining the productivity of fresh biomass in varieties and and populations of birdsfoot trefoil (2016-2017)

Parameters	Fresh biomass, g	Height stems, cm	Stem, count
2016			
Fresh biomass, g	0		
Height stems, cm	0,703	0	
Stem, count	0,627	0,311	0
2017			
Fresh biomass, g	0		
Height stems, cm	0,918	0	
Stem, count	0,887	0,844	0

CONCLUSIONS

100%
 Leo, Taborac,
 Gran San Gabriele Bosna Lotus.
 -
 (r=0.703 r=0,918).
 8, 9 10 ()
 -
 1, -
 Natural
 population, Stamm GH01, Stamm GH02,
 2 3.
 -
 (r=0.311
 r=0,844). 5
 -
 1 25,00% (2016
) 29,27% (2017).
 -

In the third year after the first crop of varieties Leo, Taborac, Gran San Gabriele and Bosna Lotus 100% plant death was detected.

The greatest influence of the yield has the height of the stems than their number regardless of the year of study ($r = 0.703$ and $r = 0.918$). In local populations 8, 9 and 10 (straining type), stems are proven shorter than the control variant variety Targovishte 1, and longer stems on average for period of study are Natural population, Stamm GH01, Stamm GH02, Bulgarian Local populations 1 and 2.

A positive correlation between biomass from one plant and the number of stems from one plant was established in both study years ($r = 0.311$ and $r = 0.844$ respectively). In the case of local population 5, a higher number of stems compared to Targovishte 1 was recorded with 25,00% (2016 years) and 29,27% (in 2017).

The greatest yield of fresh biomass and survival of the tested varieties trefoil is Bulgarian variety used for standard

1, Stamm GH01	Stamm GH02,	-	Targovishte 1, and in Austrian populations Stamm GH01 and Stamm GH02, as well as the Bulgarian Local populations 1 and 2 elite plants of which will be included as parental components of synthetic population.
1	2,		

/ REFERENCES

1. **Bozhanska, T., Ts. Mihovski, G. Naydenova, D. Knotová and J. Pelikán**, 2016. Comparative studies of annual legumes. *Biotechnology in Animal Husbandry*, 32(3), 311-320.
2. **Churkova, B.**, 2006a. Investigation of birdsfoot trefoil (*Lotus corniculatus* L.) populations and varieties grown in Central Northern Bulgaria. *Bulgarian Journal of Agricultural Science*, 12(3), 455-460.
3. **Churkova, B.**, 2006b. Evaluation test on local population of birdsfoot trefoil in mixtures with meadow grass. In: Proceeding of the 21st General Meeting of the European Grassland Federation, Badajoz, Spain, pp. 62-63.
4. **Churkova, B.**, 2008. Productive abilities, botanical composition and biological characteristics of varieties of birdsfoot trefoil. *Journal of Mountain Agricultural on the Balkans*, 11(5), 829-838.
5. **Churkova, B.**, 2009. Main productive characteristics of varieties and populations of birdsfoot trefoil and relations. *Journal of Balkan Ecology*, 12(2), 177-181.
6. **Churkova, B.**, 2012. Biochemical characterization of species and populations birdsfoot trefoil (*Lotus corniculatus* L.) grown in the region of Troyan. *Banat's Journal of Biotechnology*, 3(5), 51-57.
7. **Churkova, B.**, 2013. Content of crude protein, crude fiber and crude ash in dry mass of birdsfoot trefoil varieties and populations. *American Journal of Agricultural Science and Technology*, 1(3), 77-83.
8. **Gatari, D, V. Radi, B. Duri, M. Šari, Z. olovi and B. Petkovi**, 2013. Morphometric characteristics of *Lotus corniculatus* L. genotypes. *African Journal of Biotechnology*, 12(35), 5423-5426.
9. **Grant, W. F. and M. Niizeki**, 1999. Birdsfoot Trefoil (*Lotus corniculatus* L.), in R. J. Singh, Ed., Genetic Resources, Chromosome Engineering, and Crop Improvement. *Forage Crops* 5:153-204.
10. **Ilieva, A., V. Vasileva and A. Katova**, 2015. The effect of mixed planting of birdsfoot trefoil, sainfoin, subterranean clover, and tall fescue on nodulation, and nitrate reductase activity in shoots. *Journal of Global Agriculture and Ecology*, 3(4), 222-228.
11. **Naydenova, G. and D. Mitev**, 2015. Study of breeding populations of birdsfoot trefoil and red clover in mixtures. *Plant science*, 52(5), 30-36 (Bg).
12. **Naydenova, G., V. Vasileva and D. Mitev**, 2013. Objectives and approaches in the breeding of perennial legumes for use in temporary pasturelands. *Journal of Mountain Agriculture on the Balkans*, 18(6), 972-982.
13. **Proserpi, J. M., J. Ronfort and G. Genier**, 1999. Constraints to the introduction of Medics in French Mediterranean farming's systems. In: Proceedings of the XIII EUCARPIA Medicago spp. Group Meeting, Istituto di Miglioramento Genetico Vegetale. Perugia, Italy. Pp. 154-161.

14. **Radi , V., S. Vu kovi , . Gatari , S. Prodanovic, M. Drini , A. Kralj and D. Paj in**, 2014. Characterization of birdsfoot trefoil (*Lotus corniculatus* L.) genotypes from the local population in Bosnia and Herzegovina. *African Biodiversity and Conservation*, 44(6), 98-105.
15. **Rumbaugh, M. D., J. L. Caddel and D. E. Rowe**, 1988. Breeding and Quantitative Genetics. In: Hanson A.A., Barnes D.K., and Hill R.R. (1988). Alfalfa and alfalfa improvement. *Agronomy Monograph*, 29: 777-808.
16. **Valkov, V. T. and M. Chiurazzi**, 2016. An In Vitro Procedure for Phenotypic Screening of Growth Parameters and Symbiotic Performances in *Lotus corniculatus* cultivars Maintained in Different Nutritional Conditions. *Plants (Basel)*, ISSN 2223-7747; CODEN: PLANCD, 5 (4), 40.
17. **Vasileva, V.**, 2011. Study on productivity of perennial legume crops in mixtures. *Journal of Mountain Agriculture on the Balkans*, 14 (2), 296-307.
18. **Vasileva, V. and E. Vasilev**, 2012. Study on Productivity of some Legume Crops in Pure Cultivation and Mixtures. *Agriculturae Conspectus Scientificus*, 77 (2), 91-94.
19. **Vasileva, V.**, 2017a. Parameters Related to Nodulating Ability of Some Legumes. *International Journal of Agriculture Food Science & Technology*, 3(1), 005-008.
20. **Vasileva, V.**, 2017b. Quality Characteristics of Forage Biomass from Birdsfoot trefoil and Mixtures with Subterranean clover. In: Proceedings of Scientific Conference with International Participation "Animal Science – Challenges and Innovations", 1-3 November, 2017, Sofia, pp. 92-103 (Bg).
21. **Vuckovic, S., I. Stojanovic, S. Prodanovic, B. Cupina, T. Zivanovic, S. Vojin and S. Jelacic**, 2007. Morphological and Nutritional Properties of Birdsfoot Trefoil (*Lotus corniculatus* L.) Autochthonous Populations in Serbia and Bosnia and Herzegovina. *Genetic Resources and Crop Evolution*, 54(2), 421-428.

Antinutritional components in the forage of perennial legumes in Bulgaria

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SUMMARY

The inclusion of perennial legumes in the composition of hay and pasture grasslands contributes to a significant increase in their quality. It is well known that these plant species synthesize and contain secondary metabolites that may affect the palatability, consumption, digestibility and nutritional value of forage as well as causing metabolic disturbances and toxicosis in farm animals.

The paper reviews Bulgarian and foreign studies on content of saponins, cyanoglycosides, tannins and phytoestrogens in forage from alfalfa, red clover, white clover and bird's-foot-trefoil.

According to published data, the concentrations of these substances correspond strongly with the species and genotype affinities of the legumes, the harvesting phase and the abiotic factors of the environment. This confirms the need for a preliminary assessment of the effect of feeding with these forage species in the form of grazing or freshly cut green

mass.

Key words: perennial legumes, secondary plant metabolites

The share of perennial legumes in the composition of hay and pasture grasses contributes to a significant increase in their quality. It is well known that these plant species synthesize and contain secondary metabolites that can affect the palatability, consumption, digestibility and nutritional value of the feed, and cause metabolic disturbances and toxicosis in farm animals.

These substances are saponins, cyanoglycosides, tannins, phytoestrogens (Ilieva et al., 2001), whose synthesis can be considered for an adaptive mechanism and their concentration corresponds to the species and genotypic reaction to the environmental factors.

Saponins

Alfalfa is the legume forage crop with the highest significance and distribution in Bulgaria. It is characterized by relatively high levels of saponin content (Kertikova et al., 2000).

12 types of nitrogen free glycosides are considered as saponins, which are found in different parts of the plant (leaves, roots, seeds) and differ in their biological activity (Voutquenne et al., 2002). Some of the alfalfa saponins are detritus, which inhibits nutrition, copulation and egg production of phytophags (Golawska, 2012).

Some of them have also an allelopathic (Oleszek et al., 1992), antimicrobial (Zehavi and Polacheck, 1996), fungistatic (Czaban et al., 2013) and estrogenic activity. The synthesis of saponins in alfalfa plant varies under the influence of genetic factors (Vetesi, 1986; Rotili, 1989; Small et al., 1990; Macuha, 1991), and also depends on the growing season

mass.

Key words: perennial legumes, secondary plant metabolites

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(Pecetti et al., 2006) (Ilieva and Blazhev, 1991).

(Plhak, 1983; Price et al., 1987; Oleszek, 1996; Timbekova et al., 1996).

(Ueda et al., 1996), Gee et al. (1997) *Zanhic acid tridesmoside*,

(Lu and Jorgensen, 1987; Klita et al., 1996).

(Malik and Singhal, 2009; Morgavi et al., 2011).

(Jurzysta and Waller, 1996; Tava and Pecetti, 1998; Simeonov et al., 1999; Munro, 2009).

(Oleszek, 1990; Massiot et al., 1991; Tava et al., 2000; Voutquenne et al., 2002),

(respectively on temperatures and solar radiation), age of grassland (Pecetti et al., 2006) and degree of disease attack (Ilieva and Blazhev, 1991).

The anti-nutritional and toxic effects of alfalfa saponins on farm animals have been identified in a number of studies (Plhak, 1983, Price et al., 1987, Oleszek, 1996, Timbekova et al., 1996). In poultry and monogastric animals fed on alfalfa flour, leaf alfalfa saponins adversely affect growth rates.

They suppress forage consumption (Ueda et al., 1996) as they give it a bitter taste. According to Gee et al. (1997), the effect is mainly due to *Zanhic acid tridesmoside*, saponin that causes irritation of mucous membrane of the mouth and the digestive tract.

For ruminants, alfalfa saponins don't have a repulsive taste but inhibit microbial fermentation and synthesis in the rumen, thus changing digestive sites (Lu and Jorgensen, 1987; Klita et al., 1996).

This biological activity of alfalfa saponins in the latest studies is investigated as an opportunity to influence fermentation processes in the rumen of ruminants and as a potential way to reduce their methane emissions (Malik and Singhal, 2009; Morgavi et al., 2011).

Alfalfa saponins can cause intravascular haemolysis of erythrocytes with appropriate clinical and paraclinical changes (Jurzysta and Waller, 1996; Tava and Pecetti 1998; Simeonov et al., 1999; Munro, 2009). The haemolytic activity of saponins varies both according to their structure (Oleszek, 1990; Massiot et al., 1991; Tava et al., 2000; Voutquenne et al., 2002) and the species and category of host animals.

Pigs, horses and rabbits are significantly more sensitive than ruminants – sheep

(*HCN* – 19.4 mg/100g DM) (28.4 29.2 mg/100g DM), (Michovski and Goranova, 2006) – 7mg *HCN*/100g DM. 50-75 mg/100g DM. (Churkova, 2000; 2008). (Majak and Cheng, 1984; Carlsen and Fomsgaard, 2008). (*thiocyanate* - *SCN*-) (Crush and Caradus, 1995; Majak et al., 1987; 1990). T (Crush and Caradus, 1995).

- small-sized leaf morphotype is less cyanogenic (with *HCN* content – 19.4 mg/100g DM) compared to medium and large-sized leaf varieties (respectively 28.4 and 29.2 mg/100g DM) as well as the fact that the large-sized leaf gene plasm of foot-hill mountain regions of Bulgaria is very low in cyanoglycosides (Michovski and Goranova, 2006) - 7mg *HCN*/100g DM.

Hydrogen cyanide has a direct toxic effect on animals when it is in a concentration above 50-75 mg/100g of DM. Poisoning of the animals is fast, with cramps, difficulty breathing, rapid heartbeat and paralysis of breathing. Taking fresh forage of white clover and bird's-foot-trefoil rarely can cause such direct poisoning because the cyanoglycoside content is below critical levels and because these grasses are not normally grown independently, but in grass mixtures (Churkova, 2000; 2008).

According to some studies, strong abiotic stress during vegetation as well as high doses of mineral fertilization can lead to an increase in the concentration of cyanoglycosides in these species. Most studies have reported a toxic effect of metabolites of hydrogen cyanid dissolution (Majak and Cheng, 1984; Carlsen and Fomsgaard, 2008).

Detoxification of hydrogen cyanid occurs in the rumen or liver, depending on the animal species that take it, forming thiocyanate (*SCN*) (Crush and Caradus, 1995; Majak et al., 1987; 1990). Thiocyanate is a potential inhibitor of iodine binding in the thyroid gland, and when these legumes are taken during grazing they can cause an increase in sheep size (Crush and Caradus, 1995). In addition, cyanide metabolites may also have an effect on selenium metabolism in animals.

(*proanthocyanidins*) (Berard et al., 2011) (Burggraaf, 2003; 2008). 50-60 g/kg DM, (Aerts et al., 1999; Wen et al., 2003; Min et al., 2003). 20-40 g/kg DM, (Athanasiadou Kyriazakis, 2004; Arrigo et al, 2009; Escaray et al, 2011). (25 g CT/kg DM). (45 g CT/kg DM) (Berard et al, 2011).

coumestrol, apigenin, luteolin and quercetin (Seguin et al., 2004); *daidzein, genistein, formononetin biochanin A* (Steinshamn, 2010).

luteolin quercetin *apigenin,*

Tannins

Condensed tannins - CT (*proanthocyanidins*) are a class of flavonoid polymers contained in bird's-foot-trefoil, sainfoin (Berard et al., 2011) and in lower concentrations in red and white clover racemes (Burggraaf, 2003; 2008). The biological effect of condensed tannins depends on their concentration in fresh forage. When their content is above 50-60 g/kg DM, they are antinutritional factors because they reduce the consumption and digestibility of forage and reduce the growth of body mass and depress wool growth in sheep (Aerts et al., 1999; Wen et al. et al., 2003; Min et al., 2003).

When their content is in the range of 20-40 g/kg DM, condensed tannins reduce protein absorption in ruminants' rumen and prevent swelling and verminousness of animals (Athanasiadou and Kyriazakis, 2004, Arrigo et al, 2009, Escaray et al, 2011). The ordinary bird's-foot-trefoil cultivated in Bulgaria has optimal levels of tannin content (25 g CT/kg DM). Higher concentrations are characteristic for sainfoin (45 g CT/kg DM) and in the pre-flowering phases of plants (Berard et al, 2011).

Phytoestrogens

Perennial legumes also contain isoflavone compounds that have estrogenic activity in the animal organism and can significantly disrupt its reproductive functions. Such compounds are *coumestrol, apigenin, luteolin and quercetin* in alfalfa (Seguin et al., 2004); *daidzein, genistein, formononetin* and *biochanin A* in red clover (Steinshamn, 2010). The content of phytoestrogens in alfalfa differs in different parts of the plant, with the highest concentrations of *apigenin, luteolin* and *quercetin* being found in the flowers.

It varies also according to developmental phases, as the lowest is in the beginning

<p>al., 2004).</p> <p>(Adams, 1995),</p> <p>(Mustonen et al., 2009).</p>	<p>(Seguin et al., 2004).</p> <p>(Boller, 1994).</p> <p>(Hall, 2007).</p>	<p>of flowering phenophase (Seguin et al., 2004). Formononetin, which occurs at the highest concentrations in red clover, has a high inheritance and is not affected by environmental conditions and developmental stages. Based on this, red clover cultivars were created with low concentration of formononetin (Boller, 1994).</p> <p>Alfalfa and clover isoflavones have been shown to impair ovarian function in cows and sheep, reduce fertility rates and increase fetal mortality (Adams, 1995), and may also pass into milk of lactating cows (Mustonen et al., 2009).</p> <p>Clover phytoestrogens can cause an increase in the frequency of urate stones in sheep (Hall, 2007).</p>
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CONCLUSIONS

- The survey of studies on the
- content of saponins, cyanoglycosides,
- tannins and phytoestrogens in grass
- forage from perennial legumes shows that
- the concentrations of these substances
- are strongly correlated with the species
- and genotypic affinities of the grasses, the
- harvesting phase and the abiotic factors
- of the environment. This confirms the
- need for a preliminary assessment of the
- effect of feeding with these forage species
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/ REFERENCES

1. **Abberton, M. T., J. H. MacDuff, A. H. Marshall and M. W. Humphreys,** 2007. The genetic improvement of forage grasses and legumes to reduce greenhouse gas emissions. Plant Breeding and Genetics Programme, Institute of Grassland and Environmental Research, Aberystwyth, United Kingdom in collaboration with the Plant Production and Protection Division, Crop and Grassland Service, of the Food and Agriculture Organization of the United Nations. FAO.
2. **Adams, N. R.,** 1983. Sexual behaviour of ewes with clover disease treated repeatedly with oestradiol benzoate or testosterone propionate after ovariectomy. *Journal of Reproduction and Fertility*, 68(1), 113-117.
3. **Adams, N. R.,** 1995. Detection of the effects of phytoestrogens on sheep and cattle. *Journal of Animal Science*, 73(5), 1509-1515.

4. **Adams, N. R. and B. Y. Tang**, 1986. Changed control of cervical secretion from infertile ewes previously exposed to oestrogenic clover pasture. *Journal of Reproduction and Fertility*, 76(1), 147-152.
5. **Aerts, R. J., T. N. Barry and W. C. McNabb**, 1999. Polyphenols and agriculture: beneficial effects of proanthocyanidins in forages. *Agriculture, Ecosystems & Environment*, 75(1), 1-12.
6. **Applebaum, S. and Y. Birk**, 1979. Saponins. In *Herbivores. Their Interactions with Secondary Plant Metabolites*, pp. 539–566 (GA Rosental and TH Janzen, editors). London: Academic Press.
7. **Arrigo, Y. and F. Dohme**, 2009. Sainfoin versus alfalfa as supplements for grazing dairy cows. *Revue suisse d'agriculture*, 41(5), 283-288.
8. **Athanasiadou, S. and I. Kyriazakis**, 2004. Plant secondary metabolites: antiparasitic effects and their role in ruminant production systems. *Proceedings of the Nutrition Society*, 63(04), 631-639.
9. **Berard, N. C., Y. Wang, K. M. Wittenberg, D. O. Krause, B. E. Coulman, T. A. McAllister and K. H. Ominski**, 2011. Condensed tannin concentrations found in vegetative and mature forage legumes grown in western Canada. *Canadian Journal of Plant Science*, 91(4), 669-675.
10. **Burggraaf, V. T., P. D. Kemp, E. R. Thom, G. C. Waghorn, D. R. Woodfield and S. L. Woodward**, 2003. Agronomic evaluation of white clover selected for increased floral condensed tannin. In: *Proceedings of the New Zealand Grassland Association*, 65, 139-145.
11. **Burggraaf, V. T., P. D. Kemp, E. R. Thom, G. C. Waghorn, D. R. Woodfield and S. L. Woodward**, 2004. Performance of dairy cows grazing white clover with increased condensed tannin. In: *Proc. NZ Grassl. Assoc*, 66, 221-226.
12. **Burggraaf, V., G. Waghorn, S. Woodward and E. Thom**, 2008. Effects of condensed tannins in white clover flowers on their digestion in vitro. *Animal Feed Science and Technology*, 142(1), 44-58.
13. **Caradus, J. R., W. McNabb, D. R. Woodfield, G. C. Waghorn and R. Keogh**, 1995. Improving quality characteristics of white clover. In *Proc. Agron. Soc. NZ*, 25, 7-12.
14. **Carlsen, S. C. and I. S. Fomsgaard**, 2008. Biologically active secondary metabolites in white clover (*Trifolium repens* L.)—a review focusing on contents in the plant, plant–pest interactions and transformation. *Chemoecology*, 18(3), 129-170.
15. **Cheeke, P. R.**, 1971. Nutritional and physiological implications of saponins: a review. *Canadian Journal of Animal Science*, 51(3), 621-632.
16. **Cheeke, P. R.**, 1996. Biological effects of feed and forage saponins and their impacts on animal production. In *Saponins used in Food and Agriculture*, 377-385. Springer US.
17. **Churkova, B.**, 2000. A comparative study of varieties trefoil (*Lotus corniculatus* L.) in the foothill at North Central Bulgaria. *Plant Science*, (4), 234-237 (Bg).
18. **Churkova, B.**, 2008. Productive abilities, botanical composition and biological characteristics of varieties of birdsfoot trefoil. *Journal of Mountain Agricultural on the Balkans*, 11(5), 829-838.
19. **Churkova, B.**, 2012. Biochemical characterization of species and populations birdsfoot trefoil (*Lotus corniculatus* L.) grown in the region of Troyan. *Banat's Journal of Biotechnology*, 3(5), 51-57.
20. **Churkova, B.**, 2014. Productivity and Botanical Composition of a Mixed Sward of Birdsfoot Trefoil and Red Fescue Depending on the Term of Sowing and Proportion

of Components. *International Journal of Agriculture Innovations and Research*, 3(1), 276-280.

21. **Dewhurst, R. J., L. Delaby, A. Moloney, T. Boland and E. Lewis**, 2009. Nutritive value of forage legumes used for grazing and silage. *Irish Journal of Agricultural and Food Research*, 167-187.
22. **Escaray, F. J., A. B. Menendez, A. Gárriz, F. L. Pieckenstain, M. J. Estrella, L. N. Castagno and O. A. Ruiz**, 2012. Ecological and agronomic importance of the plant genus *Lotus*. Its application in grassland sustainability and the amelioration of constrained and contaminated soils. *Plant Science*, 182, 121-133.
23. **Fievez, V., C. Dragomir, L. Mbanzamihigo and D. Demeyer**, 2001. Clover saponins as methane inhibitors and their effect on rumen N utilisation efficiency as studied in vitro and in vivo. *Mededelingen Van De Faculteit Landbouwkundige En Toegepaste Biologische Wetenschappen, Universiteit Gent*, 66(4), 299-304.
24. **Gee, J. M., J. M. Wal, K. Miller, H. Atkinson, F. Grigoriadou, M. V. W. Wijnands and I. T. Johnson**, 1997. Effect of saponin on the transmucosal passage of -lactoglobulin across the proximal small intestine of normal and -lactoglobulin-sensitised rats. *Toxicology*, 117(2), 219-228.
25. **Goławska, S., I. Łukasik, A. Wójcicka and H. Sytykiewicz**, 2012. Relationship between saponin content in alfalfa and aphid development. *Acta Biologica Cracoviensia Series Botanica*, 54(2), 39-46.
26. **Hall, E.**, 2007. Red Clover. Pastures Australia, a collaboration between AWI, GRDC, MLA, RIRDS.
27. **Ilieva, A. and V. Blazhev**, 1991. Changes in the content of total phenols and saponins in case of alfalfa fusarium wilt. *Fiziologiya na rasteniyata*, 17(1), 49-54.
28. **Jurzysta, M., and G. R. Waller**, 1996. Antifungal and hemolytic activity of aerial parts of alfalfa (*Medicago*) species in relation to saponin composition. In *Saponins Used in Traditional and Modern Medicine*, pp. 565-574. Springer US.
29. **Kertikova, D. V., M. G. Vlahova, A. V. Ilieva and A. I. Atanassov**, 2000. Increase of yield and quality in lucerne by applying classical methods and biotechnological approaches. *Bulgarian Journal of Agricultural Science*, 6(5), 507-512.
30. **Kooyers, N. J., L. R. Gage, A. Al Lozi and K. M. Olsen**, 2014. Aridity shapes cyanogenesis cline evolution in white clover (*Trifolium repens* L.). *Molecular ecology*, 23(5), 1053-1070.
31. **Lane, L. A., J. F. Ayres and J. V. Lovett**, 2000. The pastoral significance, adaptive characteristics, and grazing value of white clover (*Trifolium repens* L.) in dryland environments in Australia: a review. *Animal Production Science*, 40(7), 1033-1046.
32. **Lu, C. D. and N. A. Jorgensen**, 1987. Alfalfa saponins affect site and extent of nutrient digestion in ruminants. *The Journal of nutrition*, 117(5), 919-927.
33. **Macuha, P.**, 1991. Contents of saponins in various lucerne genotypes. *Vedecke Prace Vyskumneho Ustavu Rastlinnej Vyroby v Piestanoch. Krmoviny (CSFR)*, 23, 59-69.
34. **Majak, W. and K. J. Cheng**, 1984. Cyanogenesis in bovine rumen fluid and pure cultures of rumen bacteria. *Journal of Animal Science*, 59(3), 784-790.
35. **Majak, W., R. E. McDiarmid, J. W. Hall and K. J. Cheng**, 1990. Factors that determine rates of cyanogenesis in bovine ruminal fluid in vitro. *Journal of Animal Science*, 68(6), 1648-1655.
36. **Majak, W., R. E. McDiarmid, K. Jakober and K. J. Cheng**, 1989. Diurnal changes in rates of degradation of cyanogenic glycosides in bovine rumen fluid. *Toxicon* (UK).

37. **Malik, P. K. and K. K. Singhal**, 2009. Effect of lucerne (*Medicago sativa*) fodder supplementation on nutrient utilization and enteric methane emission in male buffalo calves fed on wheat straw based total mixed ration. *Indian Journal of Animal Sciences*, 79(4), 416-421.
38. **Massiot, G., C. Lavaud, V. Besson, L. Le Men-Olivier and G. Van Binst**, 1991. Saponins from aerial parts of alfalfa (*Medicago sativa*). *Journal of Agricultural and Food Chemistry*, 39(1), 78-82.
39. **Min, B. R., T. N. Barry, G. T. Attwood and W. C. McNabb**, 2003. The effect of condensed tannins on the nutrition and health of ruminants fed fresh temperate forages: a review. *Animal Feed Science and Technology*, 106(1), 3-19.
40. **Morgavi, D.P., M. Eugène, C. Martin and M. Doreau**, 2011. Reducing methane emissions in ruminants: is it an achievable goal? In: Ranilla M.J. (ed.), Carro M.D. (ed.), Ben Salem H. (ed.), Morand-Fehr P. (ed.). Challenging strategies to promote the sheep and goat sector in the current global context. Zaragoza: CIHEAM / CSIC / Universidad de León / FAO, 2011. p. 65-73. (Options Méditerranéennes : Série A. Séminaires Méditerranéens; n. 99). 13. International Seminar of the Sub-Network on Nutrition of the FAO-CIHEAM Inter-Regional Cooperative Research and Development Network on Sheep and Goats, 2009/10/14-16, León (Spain). <http://om.ciheam.org/om/pdf/a99/00801537.pdf>
41. **Mustonen, E. A., M. Tuori, I. Saastamoinen, J. Taponen, K. Wahala, H. Saloniemi and A. Vanhatalo**, 2009. Equol in milk of dairy cows is derived from forage legumes such as red clover. *Br. J. Nutr.*, 102(11), 1552-1556.
42. **Oleszek, W.**, 1990. Structural specificity of alfalfa (*Medicago sativa*) Saponin Haemolysis and Its Impact on Two Haemolysis based Quantification Methods. *J. Sci. Food Agric.*, 53, 477-485.
43. **Oleszek, W.**, 1993. Allelopathic potentials of alfalfa (*Medicago sativa*) saponins: Their relation to antifungal and hemolytic activities. *Journal of Chemical Ecology* 19(6), 1063-1074.
44. **Oleszek, W.**, 1996. Alfalfa saponins: structure, biological activity and chemotaxonomy. In *Saponins Used in Food and Agriculture*; Waller, G. R., Yamasaki, K., Eds.; Plenum Press: New York, pp 155-170.
45. **Pecetti, L., A. Tava, M. Romani, M.G. De Benedetto and P. Corsi**, 2006. Variety and environment effects on the dynamics of saponins in lucerne (*Medicago sativa* L.). *European Journal of Agronomy*, 25(3), 187-192.
46. **Pihak, F.**, 1983. Determination of content of Haemolytic saponins in lucerne and its changes during growth and cuts. *Sb. Ved. Pr. Vyzk. Slechtitelsky Ustav Picninarsky Troubku Brna*, pp. 5-13.
47. **Price K., I. Johnson and G. Fenwick**, 1987. The chemistry and biological significance of saponins food and feedingstuffs. *CRC Crit. Rev. Food Sci. Nutr.*, 26, 127-135.
48. **Price, K. R. and G. R. Fenwick**, 1992. Zanhic acid tridesmoside and other dominant saponins from alfalfa (*Medicago sativa* L.) aerial parts. *J. Agric. Food Chem.*, 40, 191-196.
49. **Price, K. R., I. T. Johnson and G. R. Fenwick**, 1987. The chemistry and biological significance of saponins in food and feedstuffs. *CRC Crit. Rev. Food Sci. Nutr.*, 26, 27-135.
50. **Retana-Márquez, S., H. Hernández, J. Flores, M. Muñoz-Gutiérrez, G. Duarte, J. Vielma, G. Fitz-Rodríguez, I. Fernández, M. Keller and J. Delgado**, 2012. Effects of phytoestrogens on mammalian reproductive physiology. *Tropical and Subtropical Agroecosystems*, 15(1), 129-145.

51. **Rotili, P.**, 1989. Methodological and technical aspects of the constitution of lucerne. Varieties adapted to drying and to protein extraction. In: Proceedings of Dry Crops 89, Fourth International Green Crop Drying Congress, held at Churchill College, University of Cambridge, UK, 3-5 April 1989, pp. 22-29.
52. **Sakamoto, S., S. Kofuji, M. Kuroyanagi, A. Ueno and S. Sekita**, 1992. Saponins from *Trifolium Repens*. *Phytochemistry*, 31, 1773-1777.
53. **Saviranta, N. M. M., M. J. Anttonen, A. Von Wrightand and R. O. Karjalainen**, 2008. Red clover (*Trifolium pratense* L.) isoflavones: determination of concentrations by plant stage, flower colour, plant part and cultivar. *J. Sci. Food Agric.*, 88(1), 125-132.
54. **Seguin, P., W. Zheng and A. Souleimanov**, 2004. Alfalfa Phytoestrogen Content: Impact of Plant Maturity and Herbage Components. *Journal of Agronomy and Crop Science*, 190(3), 211-217.
55. **Sen, S., H. Makkar, and K. Becker**, 1998. Alfalfa Saponins and Their Implication in Animal Nutrition. *J. Agric. Food Chem.*, 46, 131-140.
56. **Simeonov, S. and Y. Nikolov**, 1999. Clinical veterinary toxicology. Stara Zagora, pp. 3-354 (Bg).
57. **Small, E., M. Jurzysta and C. Nozzolillo**, 1990. The evolution of hemolytic saponin content in wild and cultivated alfalfa (*Medicago sativa*, Fabaceae). *Econ. Bot.*, 44(2), 226-235.
58. **Smit, J. and K. Urbanska**, 1986. Rhodanese activity in *Lotus corniculatus* s.l. *Journal of Natural History*, 20(6), 1467-1476.
59. **Steinshamn, H.**, 2010. Effect of forage legumes on feed intake, milk production and milk quality - a review. *Anim. Sci. Pap. Rep.*, 28(3), 195-206.
60. **Tava A, M. Chiari and W. Oleszek**, 2000. Separation of alfalfa (*Medicago sativa* L.) saponins as their borate complexes by capillary electrophoresis. In: Oleszek W, Marston A (eds) Saponins in food, feedstuffs and medicinal plants. In: Proceedings of the phytochemical society of Europe, Kluwer, 45, 40-45
61. **Tava, A. and L. Pecetti**, 1998. Hemolytic activity and saponin content in lucerne *Medicago sativa* complex genotypes. *Journal of Genetics and Breeding*, 52(1), 33-37.
62. **Timbekova, A. E., M. I. Isaev and N. K. Abubakirov**, 1996. Chemistry and biological activity of triterpenoid glycosides from *Medicagosativa*. In Saponins Used in Food and Agriculture; Waller, G. R., Yamasaki, Y., Eds.; Plenum Press: New York, pp. 171-182.
63. **Ueda, H., Y. Kakutov and M. Chshima**, 1996. Growth-depressing effect of alfalfa saponin in chicks. *Anim. Sci. Technol.* 67(9), 772-779.
64. **Vetesi, M.**, 1986. Breeding of a low saponin lucerne variety for replacing the protein requirement of monogastric animals. *Enytermeles*, 287-292.
65. **Voutquenne, L., C. Lavaud, G. Massiot and L. Le Men-Olivier**, 2002. Structure-Activity Relationships of Hemolytic Saponins. *Pharmaceutical Biology*, 40, 253.
66. **Wen, L., C. Roberts, J. Williams, R. Kallenbach, P. Beuselinck and R. McGraw**, 2003. Condensed Tannin Concentration of Rhizomatous and Nonrhizomatous Birdsfoot Trefoil in Grazed Mixtures and Monocultures. *Crop Science*, 43, 302-306.
67. **Wilson, K.**, 2008. Cyanogenesis: A Chemical Defense in Plants. *Chemical Ecology*.
68. **Zehavi, U. and I. Polacheck**, 1996. Saponins as Antimycotic Agents: *Glycosides of Medicagenic Acid.*, 404, 535-546.

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Optimization of the method for the extraction of biologically active substances from fresh vegetable raw materials by enzymatic hydrolysis and cryogenic concentration

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SUMMARY

An extraction method has been optimized by incorporating in the technological order steps of cell wall enzyme grinding, ultrafiltration of the separated supernatant fluid and cryogenic concentration to obtain a liquid or dry extract with a high content of biologically active substances. An optimal algorithm has been developed to implement the different approaches for enzymatic grinding, ultrafiltration, cryogenic concentration and drying at the temperature range -50°C to 30°C. The ultimate goal of applying this algorithm is to keep thermolabile compounds to achieve maximum biological activity, and a frequency with no residual organic solvents in the final extract.

Laboratory-grown micelle culture of higher medical mushrooms of the species *Ganoderma lucidum* grown in Potato Dextrose Agar (PDA) medium supplemented with yeast extract was used as a starting material. The mycelial

Ganoderma lucidum,
Potato Dextrose

Agar (PDA)

() ,
 HPLC
 Acharya et al., 2015).
 (Krishnendu

- culture is subjected to fresh (live) processing without prior drying, by running the shock freezing process line to maximally preserve the functional stability of the biologically active substances.
-
- Proven degradation of the cell wall by microscopic observation and HPLC analysis to detect the products obtained after hydrolysis of the chitin layer (Krishnendu Acharya et al., 2015).

Key words: medicinal mushroom, enzymes, lyophilization, Ganoderma lucidum, Reishi, extract, cell wall, glucosamine, enzymatic hydrolysis, chromatographic analysis, microscopic observation

INTRODUCTION

One of the main problems in the production of biologically active products for use in medicine, functional nutrition and cosmetics, is to provide the substances isolated from the raw material with maximum biological activity.

In the different extraction methods, a different level of this indicator is observed. In the commonly used methods using solvents other than water (alcohols, glycols, ethers, gasoline, hexane, oils, paraffins, etc.) contaminations of residual amounts of solvent are observed.

Removal of the solvent usually involves raising the temperature at which a process is reported to destroy some thermally labile biologically active substances.

Solving this problem requires processes involved in the processing method to be carried out at a lower temperature than would destroy the target biologically active substances present in the feedstock.

G. lucidum is a widely used medical mushroom for the production of various products for prophylaxis and functional

G. lucidum

nutrition. This is the most commonly used mushroom for medical purposes with different therapeutic properties: anticancer activity, antioxidant, anti-inflammatory, antiallergic and neuroprotective properties, has a beneficial effect on the liver and kidneys, shows antiviral and antibacterial activity.

Extracts with a high content of biologically active substances are used to strengthen the immune system, to prevent cardiovascular diseases, hypertension, diabetes, respiratory and gastrointestinal disorders, immunodeficiency states, etc. (Solomon P. Wasser, 2014).

18-20°C (Johanna Rytioja, 2016).

2-3 %

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For the release of biologically active substances from the cell and the cell wall without the need of organic solvents, an enzymatic method for degradation of the cell structures is applied which runs in the temperature range of 18-20°C (Johanna Rytioja, 2016).

In this method the solvent is water, the necessary acidity of the solution is achieved by natural acid and the processing conditions retain the unchanged biologically active substances as naturally occurring processes in the natural systems. Removal of the solvent in this approach is carried out under vacuum and low temperature by a lyophilization process in which the conservation of volatile and thermolabile molecules is guaranteed.

During lyophilization, the water passes to a solid state during freezing, then sublimates in vacuum conditions by passing directly to the gaseous state without passing through a liquid state, thus not being able to drag with it dissolved substances that remain in the dry state fraction. The resulting dried substance remains with 2-3% humidity, which prevents the development of pathogenic processes and if the resulting product is packed with material that stops passing of water, gas and ionizing

(Pál, 2017).

radiation, a very long shelf life is achieved (Pál, 2017).

MATERIAL AND METHODS

In the present work, a method for producing a biologically active extract is provided wherein the process algorithm includes processes responsive to the temperature regime according to the preset conditions for the thermally-labile compounds as shown in Figure 1.

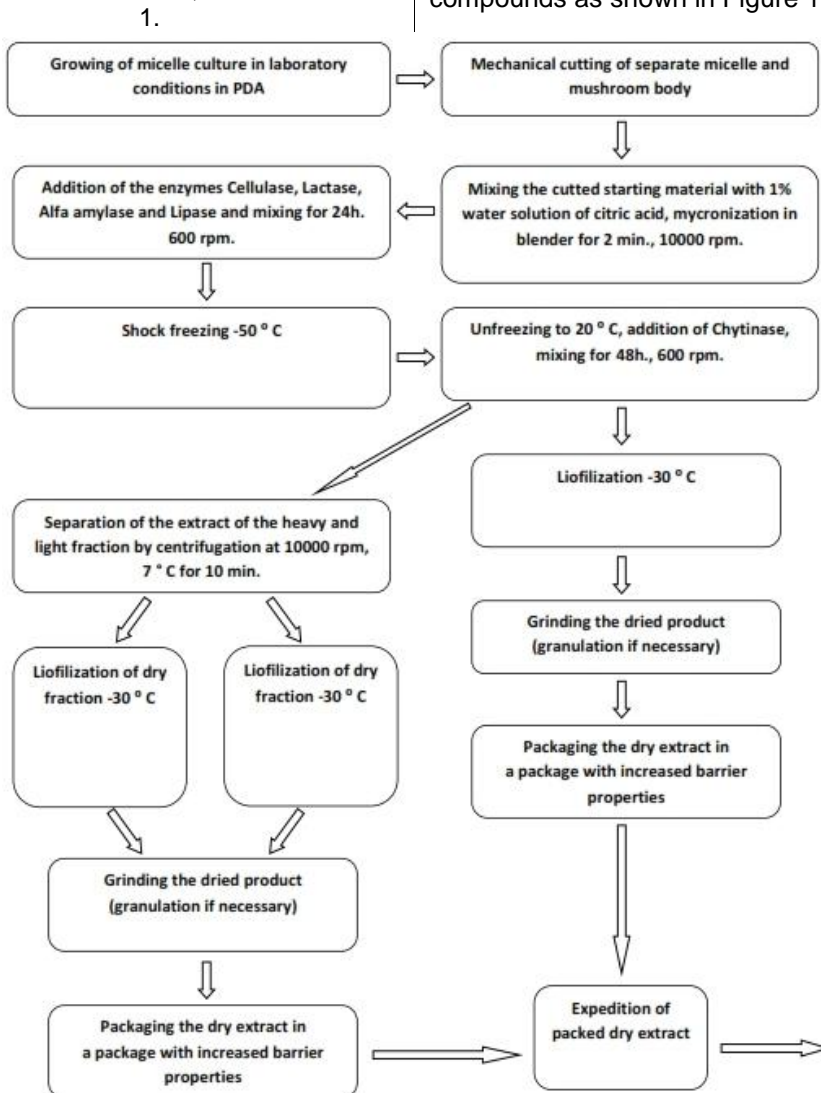


Fig. 1. Block scheme of the stages of treatment to obtain dry enzyme treated substance

<p><i>Ganoderma lucidum.</i></p>	<p>- The starting material used in this work is a live mycelium and a body from <i>Ganoderma lucidum</i>. It is selected because it is very rich in biologically active substances, containing over 400 kinds of them. The other indicator for selecting this starting material is the presence of a dense and difficult to degrade chitin-containing cell wall. The following enzymes were used to break down the cell wall: DeniLight (Lipase), Lipopan (Lipase), Fungamyl 4000 SG (Celiac Amylase), Celluclast 1.5 L (Cellulose), Chitinase in 0.5 g quantities each.</p>
<p>400</p>	
<p>: DeniLight (), Lipopan (), Fungamyl 4000 SG (), Celluclast 1,5 L (), Chitinase 0,5 g</p>	
<p><i>G. Lucidum</i></p> <p>(PDA)</p>	<p>The raw mycelia and body of <i>G. lucidum</i> was grown for six months in laboratory conditions at ICFT in Sofia. Potato Dextrose Agar (PDA) broth was used with the addition of extract from autoclaved yeast <i>Saccharomyces cerevisiae</i>. The starting material for the experiment was separated in the initial stage of the development of the body.</p>
<p><i>Saccharomyces cerevisiae.</i></p>	
<p>1% - H₂O.</p>	<p>- The initially released mushroom was mechanically cut and placed in 1% citric acid in H₂O. The solution thus obtained was micronized by mechanical grinding in a blender equipped with cutting bits at 20 000 rpm. in 2 minutes.</p>
<p>20000 ./ . 2</p>	
<p>DeniLight (), Lipopan (), Fungamyl 4000 SG (), Celluclast 1,5 L () 0,5 g</p>	<p>The enzymes DeniLight (Lipase), Lipopan (Lipase), Fungamyl 4000 SG (Alpha amylase), Celluclast 1.5 L (Cellulose) were added to the resulting micronised solution in amounts of 0.5 g each and for 24 hours was stirred with a magnetic stirrer at 600 rpm. at a temperature of 20 ° C. In the next step, the solution was shock-frozen to -50°C, then unfrozen, and at the temperature of 20°C was added the Chitinase enzyme at 0.5 g and stirred with a magnetic stirrer for 48 h at 600 rpm.</p>
<p>600 ./ . 20°C.</p>	
<p>-50°C,</p> <p>Chitinase 20°C</p>	
<p>0,5 g. 48 . 600 ./ .</p>	<p>Upon expiration of the process time, a sample was separated for microscope</p>

10000 /
10

7°C

MeOH

1:5,

100 µm,
40 µm,
1 µm,,
0,45 µm
0,2 µm.

:
HP Agilent 1260 Infinity Quaternary LC, DAD [=270 nm], RP-C18 [250 mm],

observation. The resulting extract was divided into two parts. The complex extract of the first part was directly dried by a lyophilization process, the powder product thus obtained being ready for packing and dispatching.

The second part of the solution was separated into a heavy and light fraction by centrifugation at 10000 rpm. and a temperature of 7°C for 10 minutes. A sample for chromatographic analysis was removed from the obtained supernatant. The light and heavy fraction of the resulting complex extract was dried by a lyophilization process. The dried products thus obtained are ready for packing and shipping.

Two studies were carried out to determine cell wall degradation. The first study was done by microscopic observation of samples taken from the solution at the beginning of the technological order and from a sample separated from the resulting complex extract prior to separation of the heavy and light fraction. Chromatography was done using a sample separated from the supernatant obtained after separation of the complex extract.

The sample was diluted with MeOH in a ratio of 1:5 then treated with an ultrasonic bath for complete dissolution.

The resulting pH was purified by multi-stage filtration using a 100 µm pore size filter paper, a 40 µm pore size glass filter, a 1 µm pore size glass filter, a pore size syringe cartridge 0.45 µm and a syringe cartridge with a pore size of 0.2 µm.

Chromatographic analysis was performed under the following conditions: HP Agilent 1260 Infinity Quaternary LC, DAD [= 270 nm], RP-C18 [250 mm], isocratic mode and liquid phase with composition: (A) 1% : ACN [70:30], (B)

: () 1% (H₂O) :
 ACN [70:30], (B) MeOH, A:B (90:10).
 -
 , Caffein (1000 mg/ml) Glucosamin
 HCL (100 mg/ml), N-Acetil-D-Glucosamin
 (200 mg./ml)
 ()
 0,2 µm. -
 -
 MeOH 1:10.
 Accu-scope 3012
 Micrometrics 5MP.

MeOH, A: B (90:10). A solution
 containing Caffein (1000 mg/ml)
 Glucosamine HCL (100 mg/ml), N-Acetyl-
 D-Glucosamine (200 mg/ml) dissolved in
 the mobile phase (A) of pores of 0,2 µm.
 An 1:10 dilution of MeOH was performed
 to perform the assay.

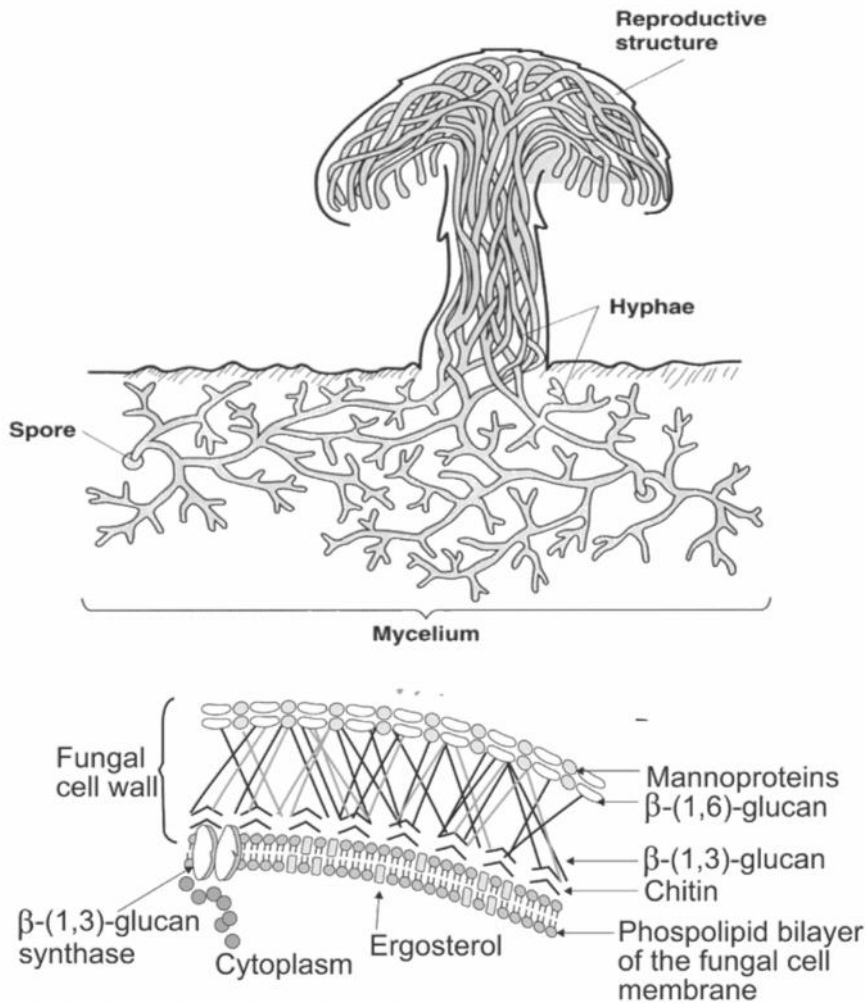
Microscopic observation of a
 sample separated from the micronised
 solution in the initial step and of the
 complex extract in the final step of the
 method was done with an Accu-scope
 3012 microscope with a Micrometrics
 5MP capture camera mounted thereon.

RESULTS AND DISCUSSION

The applied methodology proved
 the possibility of the fungal cell wall being
 destroyed without the use of organic
 solvents and high temperature. This is
 due to the enzymes DeniLight (Lipase),
 Lipopan (Lipase), Fungamyl 4000 SG
 (Cellulase), Celluclast 1.5 L (Cellulose)
 and Chitinase in combination with
 mechanical micronization and shock
 freeze.

Figure 2 shows the structure of the
 fungus and the cell wall of the fungal cell.
 The image shows that the whole structure
 is covered by filamentary formations,
 called the hyphi (Carmen Sanchez, 2017).

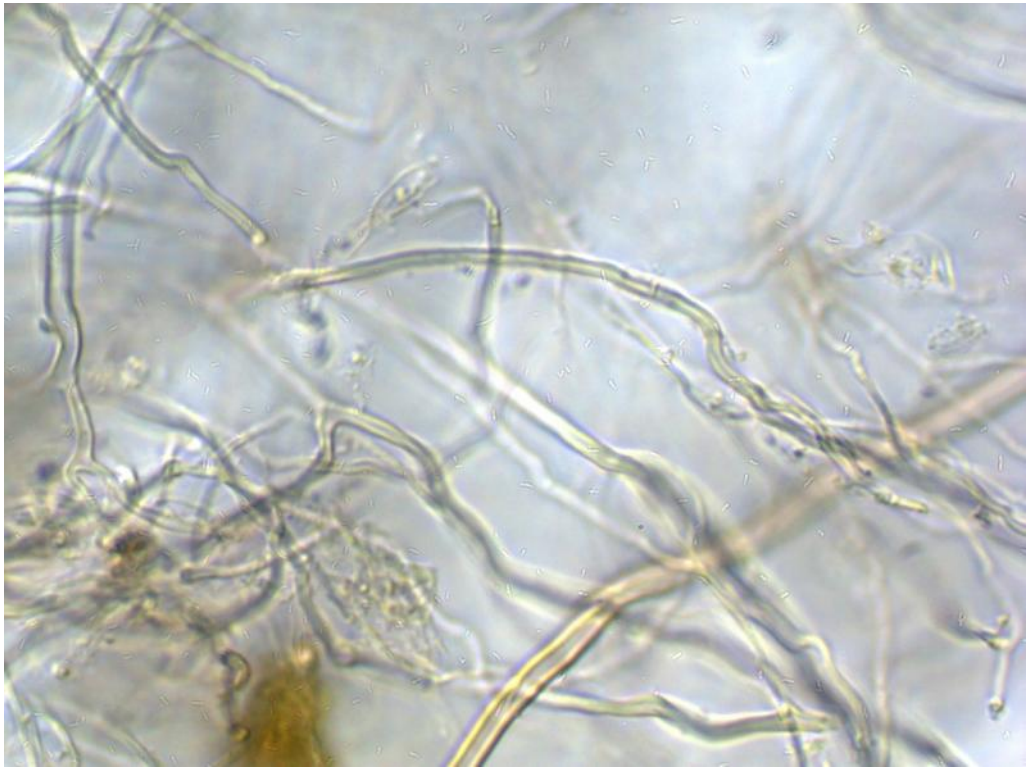
DeniLight
 (), Lipopan (), Fungamyl
 4000 SG (), Celluclast 1,5
 L () Chitinase
 -
 2 -
 -
 ,
 ,
 (Carmen Sanchez, 2017).



2.
Fig. 2. Structure of mushroom body, micelle and cell wall

- The stable chitin layer directly on the dyslipidic layer is clearly visible.
- Additional structures over the chitin layer are also seen, further protecting the cell wall.
- In order to decompose all the layers of the cell wall in the process developed, an approach is chosen in which the individual layers are broken down in several stages, in which the following steps are performed:

- Mechanical micronization – at that stage partial tear of the fibrous structure was observed.

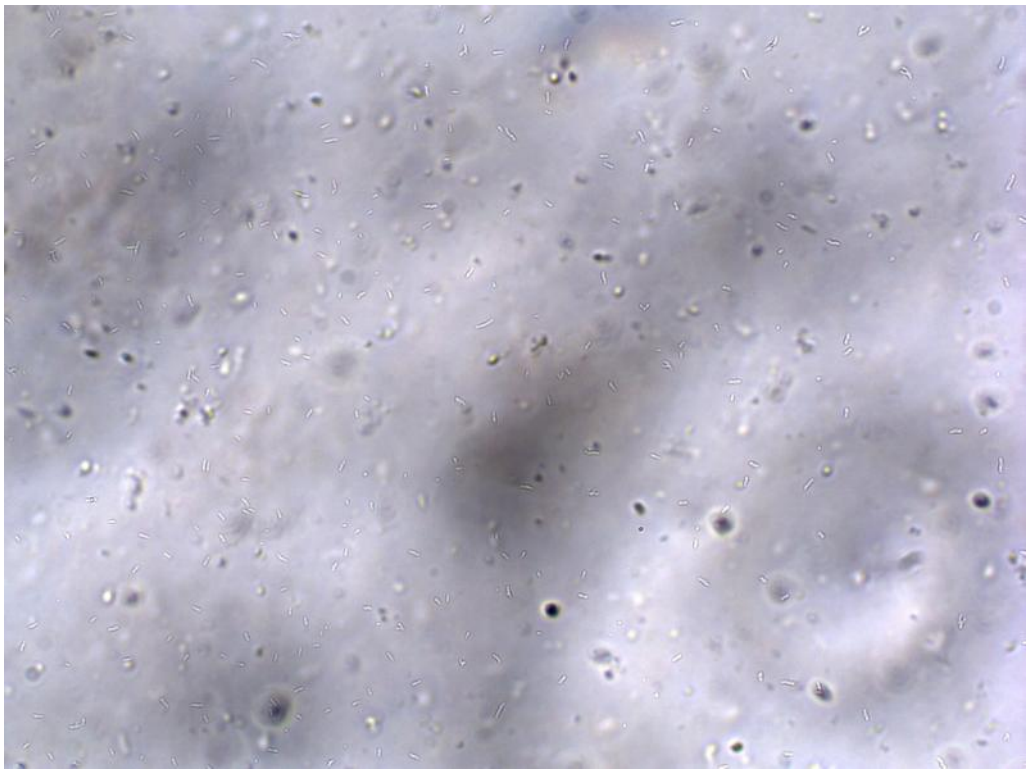


. 3.

Fig 3. Coined field of view from a microscopic observation of a preparation from the raw material after the mechanical micrization step

4

- Figure 4 shows a captured sample of the microscopic observation of the resulting complex extract in the final step after the enzymatic cell wall degradation.
- The image clearly shows the absence of whole cell structures with a non-degraded cell wall. This indicates that the cell walls are degraded and the intracellular substances have passed into the solution, thereby obtaining a biologically active extract with preserved biologically active substances as the ultimate target of the method.
- The activity is available due to the low-temperature mode in which the experiment was conducted.



. 4.

Fig. 4. Captured field of view by microscopic observation of a complex extract preparation at the final stage after enzymatic cell wall degradation

5
 -
 .
 Glucosamin HCL
 N-Acetil-D-Glucosamin.
 -
 Glucosamin
 ,
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 ;
 .

Figure 5 shows the chromatograms of the standard and the separated sample of the supernatant at the stage after separation of the complex extract. The peak of the first chromatogram is the Glucosamine HCL and N-Acetyl-D-Glucosamine peaks. On the chromatogram of the sample with the light fraction of the extract, the Glucosamine fractions are again read in the times of the standards, which is further evidence of degradation of the chitin layer from the cell wall. The chromatographic study was conducted with a view to qualitative determination without quantitative calculations not covered by this project. The purpose of this study is the only demonstration of the degradation of the chitin layer of the fungal cells.

CONCLUSIONS

From the experiments made, it can be concluded that using this method produces extracts in which all the active substances remain preserved because the process algorithm performance takes place at low temperatures which suppress the biological activity of the thermolabile compounds. The method can be adjusted depending on the need to maximally preserve target biologically active substances by changing the steps involving an enzymatic reaction as well as the use of different enzymes depending on the vegetable matrix treated of the raw material used.

/ REFERENCES

1. **Carmen Sanchez**, 2017. Reactive oxygen species and antioxidant properties from mushrooms. *Synthetic and Systems Biotechnology*, 2(1), 13-22.
2. **Johanna Rytioja**, 2016. Enzymatic plant cell wall degradation by the white rot fungus *Dichomitus squalens*.
3. **Krishnendu, Acharya, M. Bera, E. Tarafder and A. Dasgupta**, 2015. Pharmacognostic standardization of *Ganoderma lucidum*: A commercially explored medicinal mushroom. *Scholars Research Library Der Pharmacia Lettre*, 7(7), 175-181.
4. **Pál, S.**, 2017. Pharmaceutical Excipients of solid dosage forms. Institute of Pharmaceutical Technology and Biopharmacy.
5. **Solomon P. Wasser**, 2014. Medicinal Mushroom Science: Current Perspectives, Advances, Evidences, and Challenges. *Biomed J.*, 37(6), 345-356 doi: 10.4103/2319-4170.138318.

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Reinstatement of management on abandoned grassland in a mountainous area of Slovakia

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SUMMARY

350 000 ha ()

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(1.005 . .)

Picea abies (L.) H.Karst, *Alnus glutinosa*
(L.) Gaertn. *Rosa canina*
L. ()

Approximately one third of permanent grassland area of Slovakia is not utilized. These 350,000 ha (abandoned) grassland are underwent succession changes from this secondary vegetation cover to the primary one – forests (coniferous forest, especially). From agricultural or forage production point of view it is not desirable. So the aim of the experiment is to reinstate forage production on abandoned grassland in mountainous area of Slovakia. We have reintroduced different forms of management on abandoned grassland.

Experiment is located in Liptovská Teplička village (1,005 m a.s.l.) on alluvial meadow (Ľubovický Váh River) with *Picea abies* (L.) H.Karst, *Alnus glutinosa* (L.) Gaertn. and *Rosa canina* L. encroachment. Reinstatement of different (from extensive to intensive) forms of management have suppressed tree and shrub species. But, when we used some metrics to characterizes a degree of

and Vostal, 2009). (Terek and Vostal, 2009). (Peltzer et al., 2010). *Picea abies* (L.) H.Karst, *Alnus glutinosa* (L.) Gaertn. *Rosa canina* L., (Maloch, 1952).

physiognomic section within succession characterized by shifts in species composition.

The development of biocoenosis ends either climatic or edaphic climax, which is represented by the greatest biodiversity and the greatest amount of interspecific relationships (Terek and Vostal, 2009). And succession is accompanied by an increase in productivity of an ecosystem. After many years (thousands or millions) it can undergo substantial declines in productivity and retrogression will happen (Peltzer et al., 2010). But, in the earlier phases, encroachment of pioneer woody species, like *Picea abies* (L.) H.Karst, *Alnus glutinosa* (L.) Gaertn. and *Rosa canina* L., has occurred on grassland in mountainous areas, especially (Maloch, 1952).

In order to reverse the negative trend, or to stop the process and to bring it back to forage production, it is necessary to provide an extra energy (in form of cutting, mulching, or fertilization) to the grassland. Since permanent grassland are represented by both production and non-production function, it is also necessary to perform pratotechnical intervention that will lead to its rapid restoration.

The aim of the contribution is to quantify succession rate after reinstatement of management on abandoned grassland in mountain area of Slovakia. We hypothesise that reinstatement of management will promote grasses and legumes at the expense of tree and shrub species, and this abandoned grassland will restore its composition showing signs of the younger stages of succession.

MATERIAL AND METHODS

The field experiment with reinstatement of different management on abandoned grassland was established in

() 2013 . a mountain area (Low Tatras Mts.) of Slovakia in 2013. At the site (1005 m a.s.l.) in cadastre of Liptovská Teplá ka village is long-term sum of rainfall 925 mm annually (525 mm over growing period) and long-term average of temperature is 4.5°C (9.5°C in growing period). On previously 15 years non-utilized, alluvial grassland with dominance of herbs (*Alchemilla xanthochlora* Rothm., *Cruciata leavipes* Opiz, *Hypericum perforatum* L.), there was encroachment of woody species: *Picea abies* (L.), H.Karst, *Rosa canina* L. on drier microhabitats and *Salix* sp. L. and *Alnus glutinosa* (L.) Gaertn. on wetter one. In the springtime of 2013 the grassland consisted by 41 vascular plant species (excluding trees and shrubs).

() . Table 1 presents management treatments, so it represents all types of utilization from extensive to intensive form in this mountainous area (grazing was not included).

1.

Table 1. Description of pratotechnical treatments on abandoned semi-natural grassland

Treatment	/Description
1	- /None
2	abandoned grassland left to gradual succession
3	/One cut a year
4	/Two cuts a year with fertilization of P ₃₀ and K ₆₀ kg.ha ⁻¹
5	/Two cuts a year
6	/Mulching once a year
7	/Two cuts a year with fertilization of N ₄₅ kg.ha ⁻¹ + PK
8	/Two cuts a year with fertilization of N ₉₀ kg.ha ⁻¹ + PK
	/Three cuts a year with fertilization of N ₉₀ kg.ha ⁻¹ + PK

Note: numbers in subscript represent doses of nutrients

(Maloch, 1952). We mapped the vegetation before each treatment to record some changes in botanical composition by of reduced projective dominance (Maloch, 1952). Species were aggregated to main botanical group of grasses, legumes and other herbs, subsequently.

TLA (time-lag analysis) – We applied TLA method (time-lag analysis) (Collins et al., 2000), it is a

(Collins et al., 2000),

distance- based approach used to study temporal dynamics of ecological communities by regressing community dissimilarity over increasing time lags (one-year lags, two-year lags, and three-year lags – used in this paper).

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It measures the rate and origin of the change of the plant community, population or species group (Collins et al., 2000, Kampichler et al., 2012).

(Collins et al., 2000, Kampichler et al., 2012).

Time-lags are square-root transformed. Results, the slope of the regression line, have three state: positive – dissimilarity on lag implies that the community is undergoing directional change; negative – the community returns to an earlier state of the time series; and zero – community is stable (Kampichler et al., 2012).

lag

(Kampichler et al., 2012).

All data were transformed by the Hellinger transformation (Euclidean distance) $N'_{ij} = \sqrt{N_{ij} / N_{+j}}$, where N_{ij} is the population size of species i in year j , and N_{+j} is the sum of individuals across all species in year j . (Legendre and Gallagher, 2001).

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It has the advantage of making assemblages directly comparable independent of their species richness and abundance (Kampichler et al., 2012).

(Kampichler et al., 2012).

Data was processed by Statit Custom QC.

Statit Custom QC.

RESULTS AND DISCUSSION

Here, we present the results of four-year period from 2013 to 2016. Botanical composition of the main floristic group of grassland at the beginning and end of experimental period are given in Table 2. The first usage by cutting or mulching has suppressed the occurrence of woody plants in the grassland (data not shown). Frequency of utilization (including mineral fertilization) has influenced botanical composition. Non-parametric

2013

2016

2.

ANOVA (Kruskal-Wallis test: $\chi^2 = 7.02$, Df = 3, P = 0.07). Spearman correlation: $r = 0.47$, P = 0.007). ANOVA: $\chi^2 = 12.20$, Df = 3, P = 0.007 and rank correlation: $r = 0.54$, P = 0.001). (Kruskal-Wallis test: $\chi^2 = 17.67$, Df = 3, P < 0.001). Spearman correlation: $r = -0.54$, P < 0.001). (Maloch, 1952; Frame, 1992; Hopkins, 2000; Novák, 2008). Holúbek (2000), (50-70%, 10-30%, 10-20%)

Kruskal-Wallis ANOVA revealed statistically marginal differences among group of frequency on grasses abundances ($\chi^2 = 7.02$, Df = 3, P = 0.07). There was significant increment of grass presences in grassland with increasing frequency of utilization (Spearman correlation: $r = 0.47$, P = 0.007). Data about legumes have provided similar results as grasses, but with more statistical power (non-parametric ANOVA: $\chi^2 = 12.20$, Df = 3, P = 0.007 and rank correlation: $r = 0.54$, P = 0.001). Increased shares of grasses and legumes under higher frequency of utilization were at expense of other herbs. Differentiations among frequency of utilization were statistically significant (Kruskal-Wallis test: $\chi^2 = 17.67$, Df = 3, P < 0.001). Increased utilization frequency diminished the group of other herbs (Spearman correlation: $r = -0.54$, P < 0.001). Development in botanical composition of grassland was expected and this trend is generally known from literature (e.g. Maloch, 1952; Frame, 1992; Hopkins, 2000; Novák, 2008). But, optimal ratio among these groups on semi-natural grassland (grasses 50-70%, legumes 10-30%, other herbs 10-20%) according to Holúbek (2000) was not satisfied.

2.

(D%).

Table 2. Botanical composition influenced by different frequency of utilization and mineral fertilization at the beginning and ending of the experiment (D %). Data are averages of frequency of mowing or mulching

/Group	/Treatments							
	2013							
	1 (0)	2 (1)	3 (2)	4 (2)	5 (1)	6 (2)	7 (2)	8 (3)
/Grasses	32.2	53.8	37.8	48.0	47.6	51.4	61.5	58.7
/Legumes	7.8	4.8	13.3	5.1	7.8	9.1	5.3	5.2
/Herbs	60.0	41.4	45.6	45.6	44.6	38.6	31.2	34.9
	2016							
/Grasses	38.3	31.8	33.7	47.5	33.1	47.9	63.3	53.2
/Legumes	3.3	6.9	45.8	18.3	8.5	29.6	13.3	15.9
/Herbs	55.1	56.4	18.3	30.1	54.8	23.0	22.3	29.9

Note: In the parenthesis are shown frequency of utilization per annum.

<p>Kampichler et al. (2012)</p> <p>TLA</p> <p>0,02 0,25.</p> <p>3,</p> <p>(2 5)</p>	<p>Kampichler et al. (2012) mentioned the range of slope of TLA is generally between 0.02 and 0.25. Based on results shown in Table 3, treatment with one frequency of utilization (treatments 2 and 5) stabilized grassland communities by the same way, differing with biomass manipulation only. These two treatments represent a primeval systems <i>sensu</i> Kampichler et al. (2014).</p>
<p>Kampichler et al. (2014).</p> <p>(-</p> <p>8),</p> <p>(</p> <p>Peltzer et al., 2010).</p>	<p>- Three times mown semi-natural grassland (the most intensive treatment 8) shows unique sings of backward succession (not retrogression <i>sensu</i> Peltzer et al., 2010). In other words, grassland under this manipulation gets to the desired younger phase of succession.</p>
<p>Kampichler et al. (2014),</p> <p>b = 0.04</p> <p>(b <0.04)</p> <p>(b> 0.04).</p>	<p>- The most dramatic change displayed treatment with mown twice a year. Other treatments, according to Kampichler et al. (2014), show managed or disturbed systems. The authors use the slope of regression line $b = 0.04$ as a threshold between natural turnover ($b < 0.04$) and communities subjected to anthropogenic impact ($b > 0.04$).</p> <p>- Here, botanical group of grasses are the most stable constituent of alluvial grassland community with marks of younger phase of succession on treatments influenced by the highest dose of nitrogen application. Legumes on treatment without utilization undergo directional change. On other treatments this functional plant group is either stabilized or shows the younger phases of succession.</p>
<p>Brita ák et al. (2012)</p> <p>7).</p>	<p>- Group of other herbs is not eliminated enough and it is presented by the most dramatic species turnover resulting in the highest slopes of regression line (it undergoes directional change, especially under management on treatment 7).</p> <p>- Brita ák et al. (2012) found more drastic directional changes in three types of grassland: semi-natural, abandoned and ruderal. All slopes of regression line were</p>

b > 0,1.

Kampichler et al., 2014).

b > 0.1.

Overall, the reinstatement of utilization on previously abandoned grassland shows some stability. Therefore more intensive approach (like more frequent utilization, higher dose of mineral fertilizers, organic fertilizers use, direct drilling or ploughing and reseeding by productive plant species) or longer-time period of monitoring are needed.

In our experiment a shortage of this approach is a lower time periods because many ecological processes take place over longer one, including rare events, episodic phenomena, processes with high annual variability, subtle processes, and complex phenomena, leave signals at different temporal frequencies (Kampichler et al., 2014). It means that we need much longer time period.

3. TLA

Table 3. Results of TLA method. Slope of regression line (unitless)

Group/Treatment	1	2	3	4	5	6	7	8
/Grasses	0.028	0.010	0.034	0.007	0.017	0.007	-0.024	-0.014
/Legumes	0.064	-0.015	0.022	0.039	-0.017	0.024	-0.005	0.011
/Herbs	-0.030	0.009	0.032	0.057	0.007	0.052	0.087	0.000
/Community	0.062	0.003	0.087	0.103	0.007	0.083	0.058	-0.003

CONCLUSIONS

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NPK

Reinstatement of different (from extensive to intensive) forms of management have suppressed tree and shrub species immediately. But, when we used time-lag analysis to characterize a degree of succession, we were surprised that grassland has shown stable composition. Only the most intensively treatment (application of NPK fertilizers and cutting three times a year) is shifting to younger phase of succession – desired grassland with dominance of grasses and legumes, with little proportion of herbs. Our results suggest the reinstatement of management is time-consuming in regard to obtaining high-performing grassland consisting of grasses and legumes.

/ REFERENCES

1. **Blüthgen, N., C.F. Dormann, D. Prati, V.H. Klaus, T. Kleinebecker, N. Hölzel, F. Alt and S. Boch et al.**, 2012. A quantitative index of land-use intensity in grasslands: Integrating mowing, grazing and fertilization. *Basic and Applied Ecology*, 13, 207-220.
2. **Britaák, N., Hanzes and I. Ilavská**, 2012. Quantification of succession in three types of grassland. *Acta Universitatis Prešovensis: Folia Oecologica*, 55(8), 89-97 (Sk).
3. **Britaák, N., Hanzes and I. Ilavská**, 2015. Botanical composition, succession, qualitative and quantitative similarities of not-managed grassland in comparison with one-mown grassland annually. *Lúkarstvo a pasienkarstvo na Slovensku*, 9, 15-20 (Sk).
4. **Collins, S.L., F. Micheli and L. Hartt**, 2000. A method to determine rates and patterns of variability in ecological communities. *OIKOS*, 91, 285-293.
5. **Fiala, J.**, 2002. Current grassland management systems. *Úroda*, 50, 9-11 (Cz).
6. **Frame, J.**, 1992. Improved grassland management. Farming Press Books, Ipswich.
7. **Holúbek, R.**, 2000. Plants of meadows and pastures: Diversity of plants. Slovak University of Agriculture, Nitra (Sk).
8. **Hopkins, A.**, 2000. Grass: its production and utilization. Blackwell Science, Oxford.
9. **Janovi, J. and Vozár**, 2004. What's about semi-natural grassland that are not use for feed purpose. *Naše Pole*, 8, 24-25 (Sk).
10. **Kamplichler, C., C.A.M. van Turnhout, V. Devictor H.P. van der Jeugh**, 2012. Large-scale changes in community composition: determining land use and climate change signals. *PLoS ONE*, 7, doi.org/10.1371/journal.pone.0035272.
11. **Kampichler, C., D.G. Angeler, R.T. Holmes, A. Leito, S. Svensson, H.P. van der Jeugd and T. Wesolowski**, 2014. Temporal dynamics of bird community composition: an analysis of baseline conditions from long-term data. *Oecologia*, 175, 1301-1313.
12. **Legendre, P. and E.D. Gallagher**, 2001. Ecologically meaningful transformation for ordination of species data. *OIKOS*, 129, 271-280.
13. **Maloch, M.**, 1952. Forage production (Management of meadows and pastures basics of growing meadows and pastures). Orá, Bratislava (Sk).
14. **Novák, J.**, 2008. Pasture, meadows and lawns. Patria, Prievidza (Sk).
15. **Peltzer, D.A., D.A. Wardle, V.J. Allison, W.T. Baisden, R.D. Bardgett, O.A. Chadwick et al.**, 2010. Understanding ecosystem retrogression. *Ecological Monographs*, 80(4), 509-529.
16. **Ružiková, H. and H. Kalivoda**, 2007. Flower meadows: Natural wealth of Slovakia. Veda, Bratislava (Sk).
17. **Slávkiková, D. and V. Krajovi**, 1998. Biodiversity conservation and management of semi-natural grassland in Po ana – Protected Landscape Area. IUCN, Bratislava (Sk).
18. **Terek, J. and Z. Vostal**, 2009. Basics of ecology and environmental science. Prešov University, Prešov (Sk).
19. **Zaplata, M.K., S. Winter, A. Fischer, J. Kollmann and W. Ulrich**, 2013. Species-driven phases and increasing structure in early-successional plant communities. *American Naturalist*, 181, e17-e27.

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Nardus stricta L.

, 5600 ,

Impact of foliar application of organic fertilizer on the bioproductive indicators of forage of natural grassland of *Nardus stricta* L. type in the region of the Central Balkan Mountain

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SUMMARY

During the period 2011-2013, the impact of organic foliar fertilizing with Biostim was studied on *Nardus stricta* natural pasture in the area of the Central Balkan Mountain in the conditions of mountain meadow soils and 1400 m above sea level.

The yield and botanical composition of natural grassland of *Nardus stricta* type were established. The following variants were studied: 1. Nontreated variant without any application of foliar fertilizing (Control); 2. Biostim 100 ml/da; 3. Biostim 200 ml/da; 4. Biostim 300 ml/da; 5. Biostim 400 ml/da.

In all variants of foliar treatment, an increase in the yield of dry matter was found in the third year. The highest effect average for the experimental period was reported in variant 2 (100ml/da) – an excess over the control with 28.75%.

2011-2013 .
-
1400 ,
(*Nardus stricta*)
.
-
Nardus stricta.
: 1.
(); 2. 100
ml/da; 3. 200 ml/da; 4.
300 ml/da; 5. 400 ml/da.
.
-
2 (100ml/da) –
28.75%.

Nardus stricta
grostis alba, *grostis capilaris*, *Festuca ovina*, *Festuca rubra*.

Foliar fertilizing with Biostim had an impact on the botanical composition of the grassland, as it significantly increased the share of grasses and motley grasses, and decreased legumes.

The major changes in grass composition occur in the third year when *Nardus stricta*, which is specied that determine the grassland, was replaced from *Agrostis alba*, *Aggrostis capilaris*, *Festuca ovina*, *Festuca rubra*.

Key words: natural grassland, foliar fertilizing, yields, botanical composition

INTRODUCTION

The type of fertilizers and the methods of their introduction are an important factor influencing the process of optimization of the fertilization of natural lawns.

Several decay and grazing of the grass during the period of active vegetation also requires the corresponding feeding with mineral or organic fertilizers (Yakimova, 1977). It is well known that with well-regulated and balanced organic and mineral fertilization conditions are created conditions for favorable changes in the phytocenological and qualitative profile of natural grass communities (Stoeva and Vateva, 2008; Tenz et al., 2010; Samuil et al., 2013). Depending on the fertilizers used, the participation of typical species that define the grassland or grassland eco-types may be increased or decreased by Maru ca et al. (2014).

It is necessary to apply both ecofriendly fertilizing norms that are environmentally friendly and on the other hand that have a positive impact on yield and forage quality obtained from these natural resources (Totev et al., 1998; Kolczarek et al, 2008), as well as to find the necessary balance in the implementation of the whole range of technologies for upgrading in order to preserve biodiversity and ecosystem

(Stoeva and Vateva, 2008; Tenz et al., 2010; Samuil et al., 2013).

Maru ca et al. (2014).

(Totev et al., 1998; Kolczarek et al, 2008),

equilibrium.

In addition to traditional organic fertilizing, foliar organic fertilizers can also be applied as a new alternative by treating directly on the photosynthetic apparatus of plant species inhabiting the natural lawns (Jankowski et al., 1999; Jankowska-Huflejt, 2012). This fertilizing method contributes to increased yields and leads to corresponding economic benefits (Bebawi et al., 2011).

Foliar treatment of natural grasslands gives a direction for changing the floristic composition in a desired direction. Depending on the fertilizers used, it is possible to support the development of the valuable species of grass and legumes (Jankowska-Huflejt, 2006), as well as to suppress a certain group of undesirable low-productive representatives in grasslands.

According to some authors, the introduction of foliar fertilizers based on humic acids stimulates the growth of the root system of plants, increasing the consumption rate of nutrients, including phosphorus that is more difficult to reach (Armstrong, 1999; Lambers et al., 2006; Magani and Kunchida, 2009; Datta et al., 2011). According to other authors, the humic acids, which are the basis of leaf organic fertilizers, actively improve and stimulate plant respiration and photosynthesis, nitrogen and carbohydrate metabolism (Ovcharhenko, 2001; Senn et al., 2003; Sengalevich, 2007).

In a humic acid study on alfalfa, Vasileva and Kostov (2015a, b), found a higher dry root mass, a higher ratio of nitrogen in root/nitrogen in above-surface matter, a higher amount of digestible nitrogen, and higher and more stable yields of forage and seeds.

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and Balezentiene, 2010).

(Naydenova et al., 2013).

- The application of foliar humic fertilizer on natural grassland stimulates the formation of more protein and improves the nutritional value of forage (Klimas and Balezentiene, 2010). The quality of grassland improves with the increasing of share legumes in the grassland (Naydenova et al., 2013).

The lack of studies on the use of leaf humic fertilizers in natural grasslands in Bulgaria, including in the Central Balkan Mountain region, necessitates to conduct them.

The purpose of the study was to determine the impact of the annual treatment with organic fertilizer on the bioproductive indicators of natural pasture of *Nardus stricta* type, located in the region of the Central Balkan Mountain (Troyan region).

MATERIAL AND METHODS

The experiment was conducted over the period 2011-2013 on a natural grassland – a pasture (*Nardus stricta* type). The experimental design was block method in 4 replications with 5 m² plot size.

For 3 years, the following variants were studied: 1. Nontreated variant without any application of foliar fertilizing (Control); 2. Foliar application with Biostim 100 ml/da; 3. Foliar application with Biostim 200 ml/da; 4. Foliar application with Biostim 300 ml/da; 5. Foliar application with Biostim 400 ml/da.

Foliar treatment with organic fertilizer was applied once a year, and the working solution was introduced with a sprayer during active grass vegetation.

The composition of Biostim foliar fertilizer is as follows: reaction (pH) - 6.8, salt concentration 20.15, nitrogen (N) - 2.1%, phosphorus (P) - 1.54%, potassium (K) - 11.2%, magnesium (Mg) - 0.01%, iron (Fe) - 0.024%, zinc (Zn) - 0.037%.

2011-2013
-
(*Nardus stricta*)
5 m².
3
: 1.
() ; 2.
100ml/da; 3.
200ml/da; 4.
300ml/da; 5.
400
ml/da.
e
,
:
(pH) - 6.8,
20.15, (N) -
2.1%, (P) - 1.54%, (K) -
11.2%, (Ca) - 0.15%,
(Mg) - 0.01%, (Fe) - 0.024%,

(Zn) - 0.037%.
 - 2.25,
 14%, - 7%.
 -
 .
 1. :
 (kg/da) -
 105
 1 kg
 1 da
 2. -
 (%) -
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 (Lidanski, 1988)

Organic content - 2.25, humic acids - up to 14%, fulvic acids up to 7%.

The harvesting of the experimental areas was carried out at the stage of ear formation - the beginning of flowering of *Nardus stricta*.

The following indicators were studied in the experiment:

1. Dry matter yield (kg/da) determined by mowing the area of each harvest plot in replications with subsequent drying under laboratory conditions at 105 °C of plant samples of 1 kg and recalculation for an area of 1 da on the basis of dry matter.

2. Botanical changes in grassland (in %) - determined by weight analysis of grass samples taken just prior to mowing, percentage share of the main botanical groups (grasses and legumes), motley grasses (as a whole) and the participation of separate species over the years.

The statistical processing of yield data was carried out by dispersion analysis (Lidanski, 1988).

RESULTS AND DISCUSSION

Temperature and rainfall data are listed in Table 1 and Table 2.

1.
Table 1. Air temperature in °C, meteorological station in Troyan

Month Year	I	II	III	IV	V	VI	VII	VIII	IX	X	XI	XII	I-XII	IV-IX
2011	-0,2	-0,3	4,9	9,7	14,7	18,5	21	20,2	18,2	8,8	1,8	2,6	9,99	17,1
2012	-2,1	-5,4	5,6	12,2	15,1	20,8	24,2	21,7	17,8	13,2	7,6	-0,4	10,86	18,6
2013	1,1	3,8	5,4	12	17,5	18,7	19,4	22,7	15,4	12,1	7,7	0,8	11,38	17,6
Average 2011-2013	-0,4	-0,6	5,3	11,3	15,8	19,3	21,5	21,5	17,1	11,4	5,7	1,0	10,74	17,8

2.

mm,

Table 2. Monthly and annual precipitation rate in mm, meteorological station in Troyan

Month	I	II	III	IV	V	VI	VII	VIII	IX	X	XI	XII	I-XII	IV-IX
2011	22,5	24,8	41,7	68	69,1	98,4	72,9	96,8	6,8	109	2	34,3	646,3	412,0
2012	124,5	45,6	35,5	36	174	51,8	7,2	39,1	30,8	43,6	11,8	79,1	679,0	338,9
2013	43,3	62,6	49,7	92,1	90,3	274,6	61,2	14,9	22,7	53,2	27,7	15	807,3	555,8
Average 2011-2013	63,4	44,3	42,3	65,4	111,1	141,6	47,1	50,3	20,1	68,6	13,8	42,8	710,9	435,6

3.

The dry mass yields per year and average for the experimental period are shown in Table 3.

The impact on dry mass in the foliar fertilizing with Biostim in the first two years is slightly expressed - the fertilized variants do not exceed in productivity the nontreated control (var. 1, Table 3). The cause for the low productivity of *Nardus stricta* grassland is climate conditions.

In 2011, the average rainfall during the vegetation season (April to September) was 412.0 mm. Extreme droughts occurred in 2012 especially during the summer months, with a moisture content of 7.2 mm in July, the average rainfall during the vegetation period was 338.9 mm, which was extremely low (Table 2).

This drought inevitably had a negative impact on the growth and development of grass vegetation, as well as on the absorption of Biostim foliar fertilizer. In the third experimental year the impact of leaf fertilization was high. Variant 2 is characterized by very good yields of dry mass (386.18 kg/da).

Variants 3 and 4 (fertilizing at a dose of 200ml/da and 300ml/da) had also good yields, respectively, exceeding with 85.49% (var. 3) and 50.66% (var. 4). The minimum exceeding of 5.07% (var. 5) was reported for the highest fertilizing

The dry mass yields per year and average for the experimental period are shown in Table 3.

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5,07% (. 5).
 ,
 ,
 (90,3 mm 274,6 mm, 61,2 mm 17,5 19,4).
 2011-2013
 -
 100 ml/da (. 2) - 28,75%.

- dose of 400 ml/da. These results, which
 - characterize the yields, can be explained
 - by the positive impact of Biostim organic
 - fertilizer. We consider this is implemented
 - in combination with the favourable
 - climate conditions in the third year (the
 - abundant rainfall during the active
 - vegetation in May, June and July, from
 - 90.3 mm to 274.6 mm, 61.2 mm and the
 - average monthly temperatures from 17.5
 - to 19.4°C). This shows that the impact of
 - Biostim humic fertilizer on *Nardus stricta*
 - grassland is directly dependent on
 - environmental factors.

On average, for the period 2011-2013, the highest dry matter yield was obtained in the foliar fertilizing with Biostim at a dose of 100 ml/da (var. 2) - an excess of 28.75%. The other foliar fertilizing variants did not exceed the nontreated control yield.

**3.
(2011-2013),**

g/d

Nardus stricta

Table 3. Dry matter yield in kg/da by years and average over a three-year period (2011-2013), from natural grassland of *Nardus stricta* type with foliar fertilizing with Biostim

/Variants	2011			2012			2013			/Average 2011-2013		
	g/d	% compared to C	Proof	g/d	% compared to C	Proof	g/d	% compared to C	Proof	g/d	% compared to C	Proof
1. Non-treated - C	217.94	100.0	-	173.43	100.0	-	140.32	100.0	-	177.23	100.0	-
2. /Biostim (100 ml/da)	165.07	75.74	-	133.32	76.87	-	386.18	275.21	+++	228.19	128.75	-
3. /Biostim (200 ml/da)	134.63	61.77	-	142.23	82.01	-	260.28	185.49	+++	179.04	101.02	-
4. /Biostim (300 ml/da)	129.58	59.45	-	144.37	83.24	-	211.40	150.66	++	161.78	91.28	-
5. /Biostim (400 ml/da)	176.19	80.84	-	164.85	95.05	-	147.44	105.07	-	162.83	91.87	-
GD 5%	137.08	62.74		133.61	77.01		41.41	29.47		87.22	49.14	
GD 1%	192.41	88.06		187.54	108.09		58.13	41.37		122.43	68.98	
GD 0.1%	271.64	124.32		264.76	152.60		82.06	58.41		172.85	97.38	

56,97 % (1).

- Botanical composition of grassland
 - Perennial grasses took the highest
 - average share in *Nardus stricta* pasture
 - for the three-year period with foliar
 - fertilizing with Biostim - 56.97% (Figure
 - 1). The highest share was found in the

- 72,6%.
 - 41,75%.
 - 25,32%,
 3,36%.
 200ml/da (.3),
 400 ml/da (.5),
 ().
 4.
 17,2%,
 (.2) 13,0%,
 26,0%.
 200 ml/da (.3),
 - 38,5%,
 - 19,2%.
 300 ml/da (.4),
 - 25,0%,
 50,0%.
 (.5),
 6,3% 36,8% 15,8%,
 5,3%.

- second year, when they reached 72.6%.
 - In the third year, the share of grasses decreased by almost a half compared to the previous year. Motley grasses occupied an average of 41.75%.

Contrary to grass species, their tendency was to grow from the first to the third year. They took the smallest share in the second year - 25.32%, with a stronger presence of grass species. Legumes took an average of 3.36%. They were a bit more in the second year.

Grass species occupied the largest share in the third variant at the second fertilizing dose of 200ml/da (var. 3), motley grasses had the lowest share for the first variant, while they had the highest share in the nontreated first variant (var. 1C). Legume species had the highest share in the treatment with 400 ml/d (var. 5), and the lowest share in the first variant (nontreated).

The share of individual species in the grassland under the impact of foliar fertilization with Biostim is shown in Table 4. Data in the first experimental year show that grass species prevail in all variants: sheep fescue, *Nardus stricta*, bentgrass. In var. 1 (C) sheep fescue is 17.2% and *Nardus stricta* is 13.8%. At the first fertilizing dose of 100 ml/da, *Nardus stricta* is 13.0%, bent grass is 13.0% and sheep fescue reaches 26.0%. When treating the grassland with 200 ml/da (var. 3), sheep fescue has the highest percentage share of grass species - 38.5%, followed by *Nardus stricta* - 19.2%. After treatment of the grassland with a dose of 300 ml/da (var. 4), bent grass and sheep fescue have an equal percentage share - 25.0%, motley grasses amounts to 50.0%. At the highest dose of foliar treatment (lime 5), sheep fescue and *Nardus stricta* were 36.8% and 15.8%, respectively, bent grass was 6.3% and timothy grass was 5.3%.

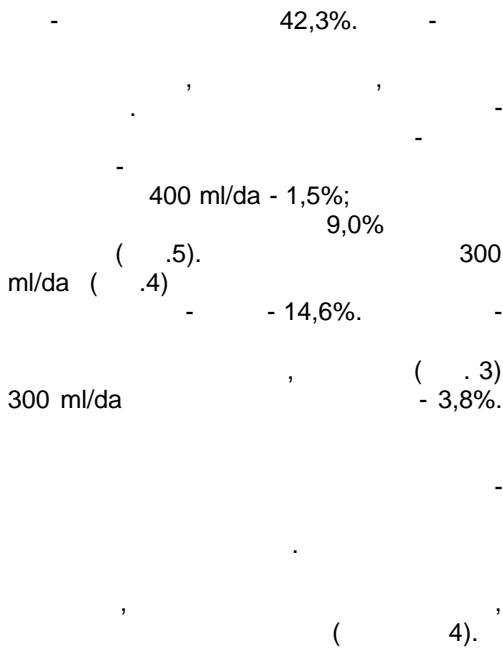
- 58,7%,
 .2 – 2,6%.
 : ,
 ,
 :
 ,
 :
 ,
 -
 .
 -
 200 ml/da
 (.3),
 45,0% ,
 - 31,5% .
 -
 - 13,6%
 400 ml/da,
 (.5) – 41,1%.
 ,
 5,9%
 3,5% (5 4). -
 .1
 () – 6,7%, - .3 – 5,4%,
 -
 1,3%.
 (2011 .)
 -
 .
 65,6%
 56,8% (5 1).
 ,
Nardus stricta L.
 .
 -
 -
 .
 100ml/da (.2),

Motley grasses took the highest share of 58.7% in the control variant, while legumes were found only in variant 2 - bird's-foot-trefoil with 2.6%. The predominant types of grasses in total for all tested variants are: *Nardus stricta*, sheep fescue, common bentgrass. Motley grasses had a high percentage share in all tested variants. During the second year, under the impact of Biostim can be seen that in all variants predominate grass species, such as: white bentgrass, common bentgrass and *Nardus stricta*.

The highest percentage of grasses was recorded in fertilization with Biostim at a dose of 200 ml/d (var. 3), as white bentgrass reached 45.0% and bentgrass - 31.5%. Motley grasses took the lowest share - 13.6% and the highest in fertilization at a dose of 400 ml/da, (var. 5) - 41.1%.

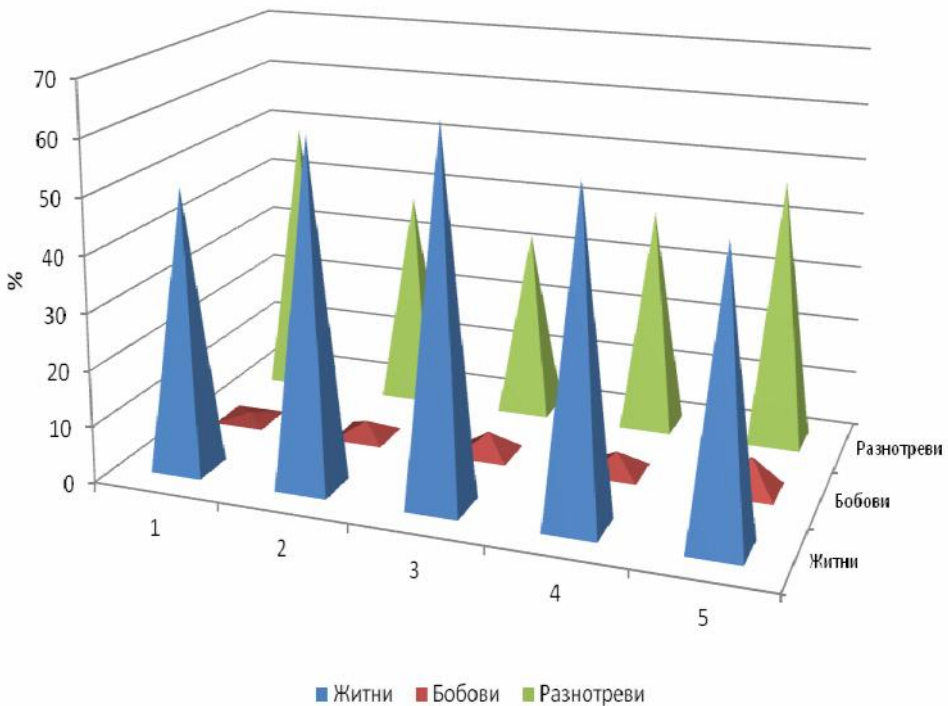
The legumes were presented by bird's-foot-trefoil in two variants with foliar fertilizing, ranging from 5.9% to 3.5% (variants 5 and 4). The highest share of *Nardus stricta* was found in var. 1 (K) - 6.7% and the lowest share in var.3 - 5.4%, as the difference between the highest and the lowest percentage of this type in the variants was 1.3%. Compared to the first year (2011), there was a visible reduction in the population of this species.

The botanical changes occurring in the grass composition during the third experimental year are characterized by the following features. The share of motley grasses ranged from 65.6% to 56.8% (variants 5 and 1). There was an increased share of grasses, as the dominant species of *Nardus stricta* L. was not found in the grassland. This year, common bentgrass was a predominant grass species in all variants. When the grassland was treated at a dose of 100ml/da (var. 2), they had the highest share and reached 42.3%. Cock's foot,



sheep fescue and Alpine timothy-grass took a lower share.

The highest share of Alpine timothy-grass was found in the highest fertilization dose with Biostim from 400 ml/da - 1.5%; while red fescue share was 9.0% in the same variant (var. 5). The highest percentage of red fescue of 14.6% was found at a dose of 300 ml/d (var. 4). The legumes were presented by mountain clover as they reached 3.8% share at a dose of 300 ml/da (var. 3). In this year, it is striking that the main species of *Nardus stricta* declined while some grass species were predominant, such as common bentgrass. Over the years, white bentgrass, common wolves, *Nardus stricta* and sheep fescue dominated in *Nardus stricta* pasture (Table 4).



. 1.

***Nardus stricta*
2011-2013 .**

Fig. 1. Botanical composition of natural grassland of *Nardus stricta* type by weight percentage per group, years and average for the period 2011-2013

4.

Nardus stricta

2011-2013 .

Table 4. Botanical composition of natural grassland of *Nardus stricta* type in weight percentage, by species, for the period 2011-2013

/Variants	/Species	2011	2012	2013
1. /Control	<i>Nardus stricta</i>	13.8	6.7	-
	<i>Agrostis capilaris</i>	10.3	20.0	30.9
	<i>Dactylis glomerata</i>	-	-	1.0
	<i>Trifolium pratense</i>	-	-	1.0
	<i>Festuca ovina</i>	17.2	6.7	-
	<i>Agrostis alba</i>	-	33.3	-
	<i>Festuca rubra</i>	-	-	10.3
	Weeds participate	58.7	33.3	56.8
2.100ml/da	<i>Nardus stricta</i>	13.0	6.3	-
	<i>Festuca ovina</i>	26.0	12.5	-
	<i>Festuca rubra</i>	-	-	1.9
	<i>Agrostis alba</i>	-	37.5	1.9
	<i>Phleum alpinum</i>	-	-	3.8
	<i>Agrostis capilaris</i>	13.0	25.0	42.3
	<i>Lotus corniculatus</i>	2.6	-	-
	Weeds participate	45.4	18.7	50.1
3.200ml/da	<i>Nardus stricta</i>	19.2	5.4	-
	<i>Agrostis capilaris</i>	3.8	31.5	39.0
	<i>Agrostis alba</i>	-	45.0	-
	<i>Festuca ovina</i>	38.5	4.5	9.5
	<i>Trifolium montanum</i>	-	-	3.8
	Weeds participate	38.5	13.6	47.7
4.300ml/da	<i>Nardus stricta</i>	-	5.6	-
	<i>Agrostis capilaris</i>	25.0	21.0	29.2
	<i>Agrostis alba</i>	-	42.0	-
	<i>Festuca ovina</i>	25.0	7.0	-
	<i>Festuca rubra</i>	-	-	14.6
	<i>Phleum alpinum</i>	-	-	3.8
	<i>Lotus corniculatus</i>	-	3.5	-
	Weeds participate	50.0	19.9	50.1
5.400ml/da	<i>Nardus stricta</i>	15.8	5.9	-
	<i>Agrostis capilaris</i>	6.3	11.8	23.9
	<i>Festuca rubra</i>	-	-	9.0
	<i>Festuca ovina</i>	36.8	5.9	-
	<i>Agrostis alba</i>	-	29.4	-
	<i>Phleum alpinum</i>	5.3	-	1.5
	<i>Lotus corniculatus</i>	-	5.9	-
	Weeds participate	36.8	41.1	65.6

Collingwood, Australia, *Rangeland Journal*, 33(3), 277-286.

3. **Datta, S., C.M. Kim, M. Pernas, N. Pires, H. Proust, T. Thomas, P. Vijayakumar and L. Dolan**, 2011. Root hairs: development, growth and evolution at the plant-soil interface. *Plant & Soil*, 346: 1-14.

4. **Jankowska-Huflejt, H.**, 2006. Analysis of yields and botanical composition of meadow sward changes under folareee and soil fertilisation with nitrogen. Wydawnictwo Uniwersytetu Marii Curie-Skłodowskiej, Lublin, Poland, *Annales Universitatis Mariae Curie-Skłodowska. Sectio E, Agricultura*, 61, 397-404.

5. **Jankowska-Huflejt, H.**, 2012. Effect of fertilization and folareee feeding on the yields, persistence of species and chemical composition of meadow sward in production conditions, Wydawnictwo Uniwersytetu Przyrodniczego we Wrocławiu, Wrocław, Poland, *Zeszyty Naukowe Uniwersytetu Przyrodniczego we Wrocławiu - Rolnictwo*, 103, 589, 115-123.

6. **Jankowski, K., J. Jodelka and R. Kolczarek**, 1999. Effectiveness of permanent meadow folareee fertilisation with nitrogen Wydawnictwo Akademii Rolniczej we Wrocławiu, Wrocław, Poland, *Electronic Journal of Polish Agricultural Universities, Agronomy*, 2(2), 1-6.

7. **Klimas, E., L. Balezientiene, H. Schnyder, J. Isselstein, F. Taube, K. Auerswald, J. Schellberg, M. Wachendorf, A. Herrmann, M. Gierus, N. Wrage and A. Hopkins**, 2010. Pastures feeding value response to humic fertilizers. In: Proceedings of the 23rd General Meeting of the European Grassland Federation, Kiel, Germany, 29th August - 2nd September , pp 193-195, 2 ref.

8. **Kolczarek, R., G. A. Ciepiela, J. Jankowska, J. Jodelka**, 2008. Influence of different forms of nitrogen fertilization on the content of macrolements (K, Na) in meadow sward: Part I. Polish Society for Magnesium Research, Olsztyn, Poland, *Journal of Elementology*, 13(3), 341-348.

9. **Lambers, H., M.W. Shane, M.D. Cramer, S.J. Pearse and E.J. Veneklaas**, 2006. Root structure and functioning for efficient acquisition of phosphorus: matching morphological and physiological traits. *Ann. Bot.*, 98: 693-713.

10. **Lidanski, T.**, 1988. Statistical methods in biology and agriculture. Zemizdat, Sofia, pp. 150-187 (Bg).

11. **Magani, I.E. and C. Kunchida**, 2009. Effect of phosphorus fertilizer on growth, yield and crude protein content of cowpea. *J. Appl. Biosciences*, 23: 1387-1393.

12. **Maru ca, T., N. Dragomir, V. Mocanu, A. Blaj, F. Tarjoc, C. Dragomir and S. Constantinescu**, 2014. Effect of some improvement works on the floristic composition of the vegetal cover in *Nardus stricta* grasslands. *Animal Science and Biotechnologies*, 47(1), 144-150.

13. **Naydenova, G., Ts. Hristova and Y. Aleksiev**, 2013. Objectives and approaches in the breeding of perennial legumes for use in temporary pasturelands. *Biotechnology in Animal Husbandry*, 29(2), 233-250.

14. **Ovcharenko, .**, 2001. Humates-activators for crop productivity. *Agronomical Bulletin*, No2, 13-14 (Ru).

15. **Samuil, C., V. Vintu, C. Sirbu and M. Stavarache**, 2013. Influence of fertilizers on the biodiversity of semi-natural grassland in the Eastern Carpathians. University of Agricultural Sciences and Veterinary Medicine, Cluj-Napoca, Romania, *Notulae Botanicae, Horti Agrobotanici, Cluj-Napoca*, 41(1), 195-200.

16. **Sengalevich, G.**, 2007. The natural biostimulating organic fertilizer "Humustim" (potassium humate) and its impact on the plant organism. Humustim - a gift from nature, the fertilizer of the future. "DIMI 99" Publishing house, pp. 29-31 (Bg).

17. **Senn, T. L., A. R. Kingman and W. C. Godley**, 2003. A review of humus and humic acids. Research series N 165, South Carolina Agricultural Experimental Station, Cleruson, South Carolina.
18. **Stoeva, K. and V. Vateva**, 2008. Productivity and variation of the grassland of natural pastures in Strandzha mountain, fertilized with organic and mineral fertilizers. Jubilee National Conference - 1-2.10.2008, Kardzhali (Bg).
19. **Tenz, R., R. Elmer, O. Huguenin-Elie and A. Lüscher**, 2010. Effects of fertilisation on a mat-grass grassland. Agroscope FAL Reckenholz, Zürich, Switzerland, Recherche Agronomique Suisse, 5, 176-183.
20. **Totev, T., V. Lingorski, K. Tankov, Ts. Mihovski, B. Churkova, D. Karadocheva, K. Belperchinov and D. Pavlov**, 1998. Changes in productivity and nitrate content of feed under the influence of mineral fertilization in mountain meadows, *Journal of Mountain Agriculture on the Balkans*, 1(5), 366-373.
21. **Vasileva, V. and O. Kostov**, 2015a. Effect of mineral and organic fertilization on alfalfa forage and soil fertility. *Emirates Journal of Food and Agriculture*, 27(9), 678-686.
22. **Vasileva, V. and O. Kostov**, 2015b. Effect of mineral and organic fertilization of alfalfa on some seed yield characteristics, root biomass accumulation and soil humus content. *Acta Agriculturae Serbica*, XX(39), 51-64.
23. **Yakimova, Y.**, 1977. Improvement and use of natural meadows and pastures. Zemizdat, Sofia (Bg).