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1407 , .

Assessment of the degree of impact of factors such as temperature and storage period on the qualitative indicators of probiotic goat milk variants

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SUMMARY

On the basis of goat milk, by selecting and introducing additional strains to the classical yoghurt yeast, biofermentation experiments were carried out to obtain probiotic products. The influence of the factors as temperature and duration of storage on the main qualitative indicators of three variants of probiotic products based on goat milk were studied.

The degree of impact of the basic technological parameters on the acidity and the total number of micro-organisms of the lactic acid sample variants was assessed by the two-factor dispersion analysis.

The statistical processing of the data obtained showed that with respect to

- titratable and active acidity, the temperature factor had a greater impact, whereas the degree of impact of the storage time factor had a major influence on the total number of microorganisms.

Key words: probiotic products, goat milk, two-factor dispersion analysis

INTRODUCTION

- Probiotic products have been consumed for centuries as a diet with dietary, prophylactic and healing value.
- The most widespread dairy probiotics are fermented milk (Chandan, 1999).

Probiotic dairy products improve the survival of healthy bacteria and the adhesion of living microorganisms in the gastrointestinal flora, thus improving the microbial balance of the gastrointestinal tract.

- In the industrialized world, probiotic foods have become part of the daily diet and have been proven with potential health benefits beyond widely accepted nutritional effects (López et al., 2014).

- Providing probiotic products is a challenge as various factors influence the viability of probiotic microorganisms in the processing and storage process. Therefore, over the last decades, attempts have been made to produce new, safe and quality products (Tripathi and Giri, 2014). In order to preserve consumer confidence in probiotic products, it is important to ensure a high survival of microbial content during production and storage of the products (Saxelin et al., 1999; Cruz et al., 2010).

- The aim of the present study is to assess the degree of impact of basic technological parameters on acidity and the total number of microorganisms of lactic acid product variants by means of two-factor dispersion analysis.

MATERIAL AND METHODS

The study used fresh goat milk with pH – 6.70 and °T=18, pasteurized at 93-95 °C for 15 minutes and cooled to 45-46 °C. The optimum temperature for the fermentation process is 44±2 °C.

Lyophilized strains of *Lactobacillus bulgaricus* 1381, *Streptococcus thermophilus* 1374, *Lactobacillus casei* 1014 and *Bifidobacterium longum* 1714 provided by the ICFT collections were used to obtain the probiotic products. The microorganisms are combined in three probiotic combinations containing classical yoghurt starter enriched with strains of lactic acid bacteria in the following ratio:

1 – *L.bulgaricus*: *Str. thermophilus*, 1:3;
 2 – *L. bulgaricus*: *Str. thermophilus*: *L. casei*, 1:3:1;
 3 – *L.bulgaricus*: *Str. thermophilus*: *B. bifidum*, 1:3:1.

Variant 1 – *L.bulgaricus*: *Str. thermophilus*, 1:3;
 Variant 2 – *L. bulgaricus*: *Str. thermophilus*: *L. casei*, 1:3:1;
 Variant 3 – *L.bulgaricus*: *Str. thermophilus*: *B. bifidum*, 1:3:1.

Physicochemical analyzes: active acidity (pH) – potentiometric; titratable acidity (°T), (BSS 1111-80).

Microbiological analyzes

• Determination of the total number of lactic acid bacteria (CFU/ml). The analysis will be performed by counting the colonies forming units per ml of typical for strains of selective media. For enumeration of colonies of *S.thermophilus* – M17 agar or ST agar, for *L.bulgaricus* – MRS-agar and for *Lactobacillus casei* – LC-agar. The strains were incubated for 72 hours at temperatures corresponding to their specificity. The number of lactic acid bacteria is calculated according to IDF standard 117B: 1991 by the formula:

$$N = \frac{\sum C}{V \times (n_1 + 0,1 \times n_2) \times d}$$

Where: C is the sum of all the listed colonies in the two successive dilutions, V – seed volume, n₁ is the number of

, n_1

, n_2

d

ml

Minitab 16 Statistical Software.

petri dishes used in the first dilution, n_2 is the number of petri dishes used in the second dilution, and d is the dilution factor.

- Reporting survival of the included strains of lactic acid bacteria during storage, and the ratio between them in each of test samples – it is calculated on the basis of the amount of cells per ml of the different strains.

Statistical processing

The results obtained are statistically processed with the Minitab 16 Statistical Software.

RESULTS AND DISCUSSION

During prolonged storage changes in the composition of the probiotic products are inevitable. By varying and controlling the technological parameters – temperature regime and duration of storage can be created optimal conditions for obtaining probiotic products with good flavour receptivity reserved qualitative indicators.

The samples tested were stored for a different period of time (1, 7, 14 and 21 days) at temperatures of 5, 10 and 15 °C.

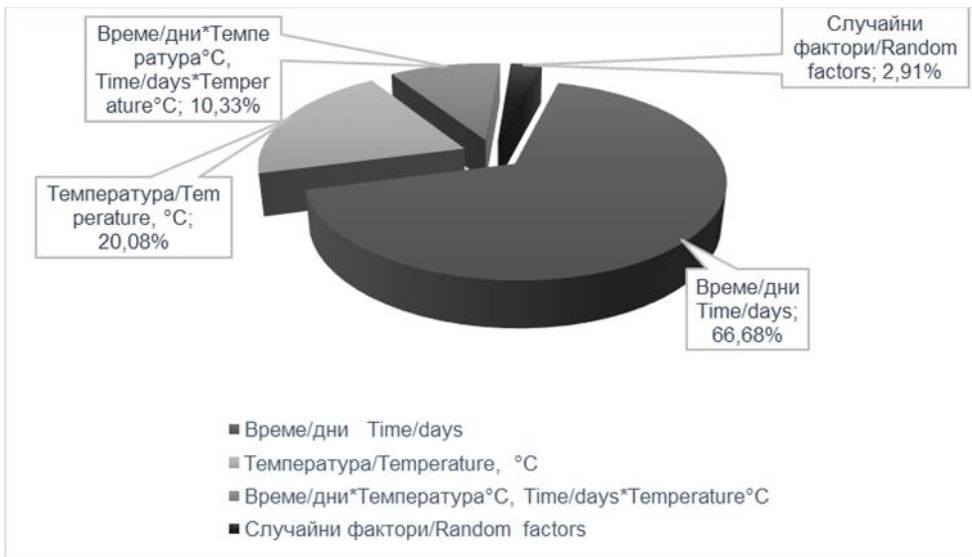
The degree of impact of the basic technological parameters on the acidity and the total number of microorganisms of the lactic acid sample variants was assessed by the two-factor dispersion analysis.

Figures 1, 2 and 3 show statistical data from a two-factor dispersion analysis for the influence of storage time and storage temperature factors on the change in titratable acidity of lactic acid sample variants.

14 21)

(1, 7, 5, 10 15

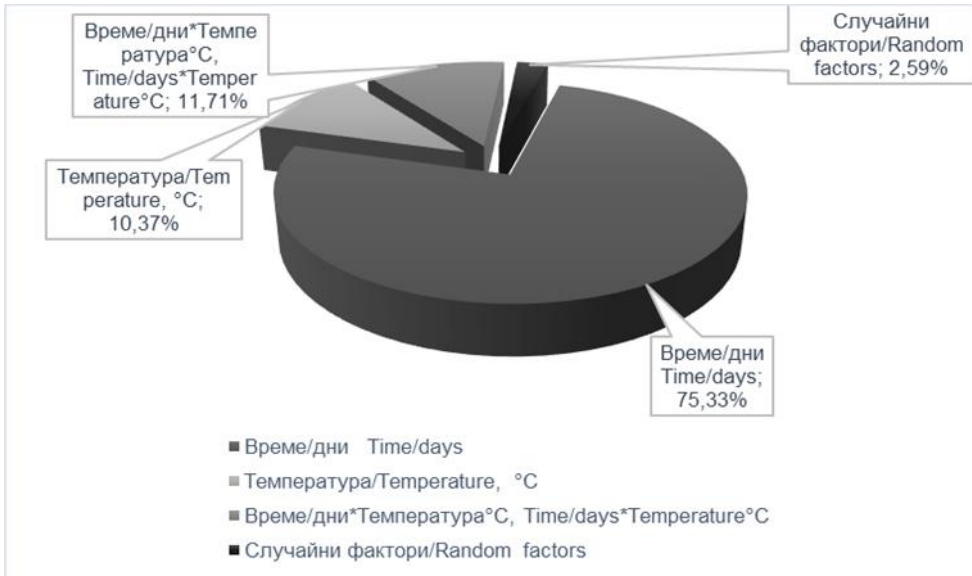
1, 2 3



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1

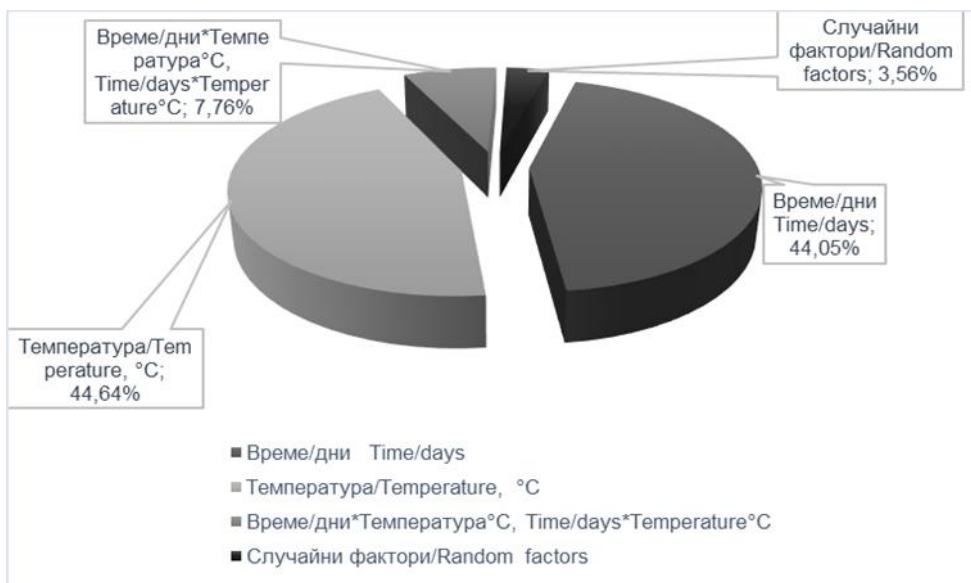
Fig. 1. Impact of temperature and storage time factors on titratable acidity of Variant 1



. 2.

2

Fig. 2. Impact of temperature and storage time factors on titratable acidity of Variant 2

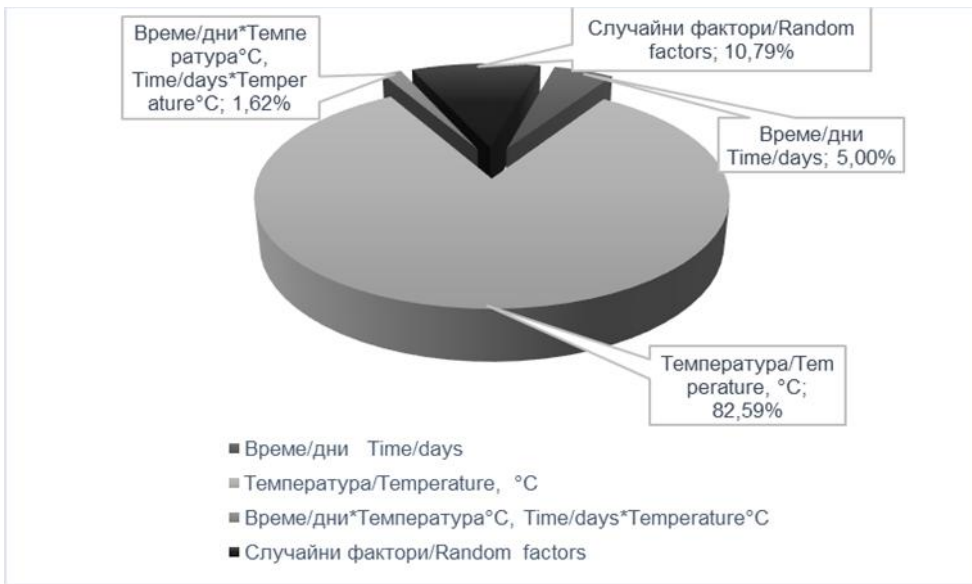


. 3.

3

Fig. 3. Impact of temperature and storage time factors on titratable acidity of Variant 3

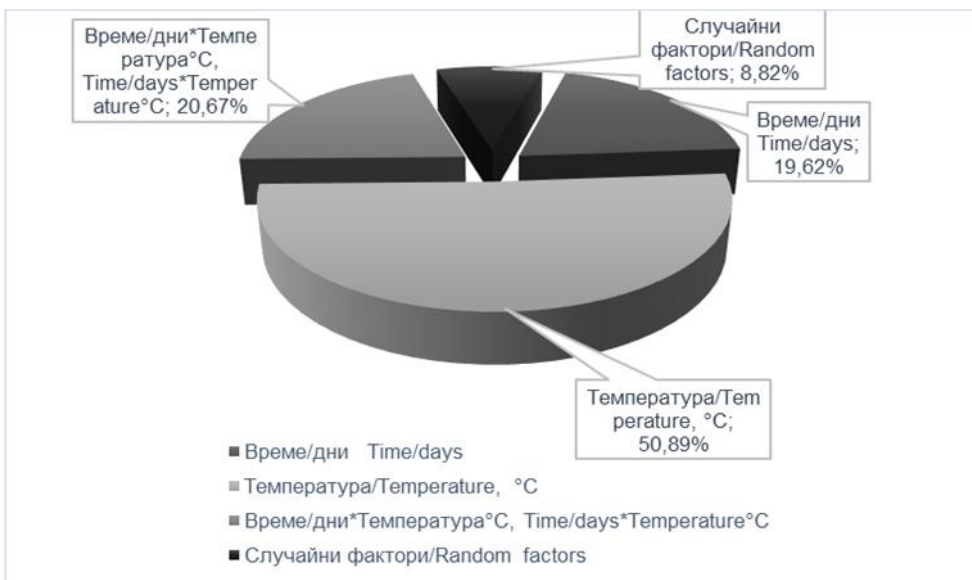
1	2	-	In variants 1 and 2 it was found that titratable acidity was influenced to a greater extent by the storage time factor and to a negligible degree by the temperature. While in variant 3 the influence of the time factor decreases at the expense of an increase of the temperature factor.
		-	
		-	
	3	.	
		.	
		-	In the studies performed the influence of both factors as well as the combination between them was statistically significant at a level of significance of 0.05 (p <0.05).
0,05 (<0,05).		-	
4, 5	6	-	Figures 4, 5 and 6 show statistical data from a two-factor dispersion analysis for the influence of storage time and temperature factors on the change in the active acidity of the probiotic variants.
		-	
		.	



. 4.

1

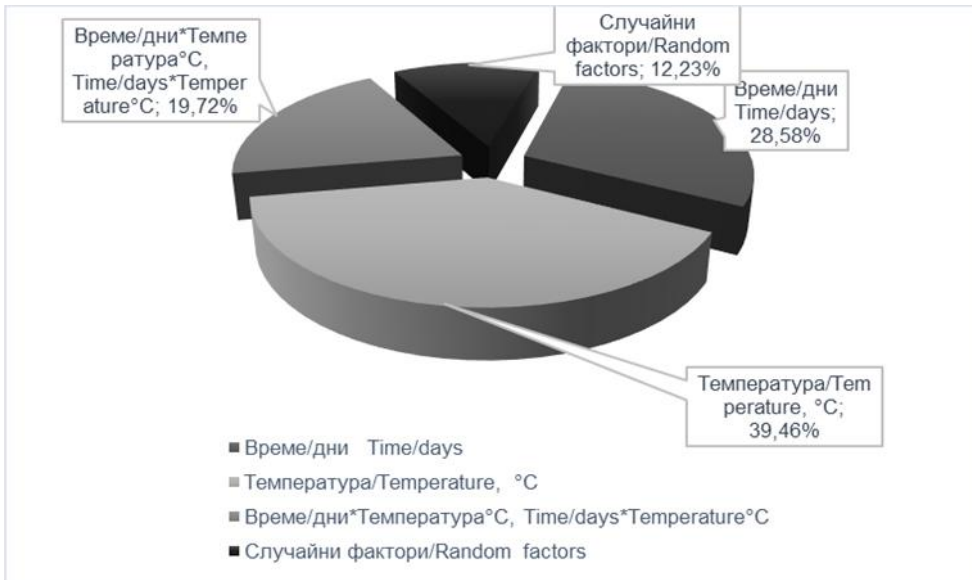
Fig. 4. Impact of temperature and storage time factors on the active acidity of Variant 1



. 5.

2

Fig. 5. Impact of temperature and storage time factors on the active acidity of Variant 2

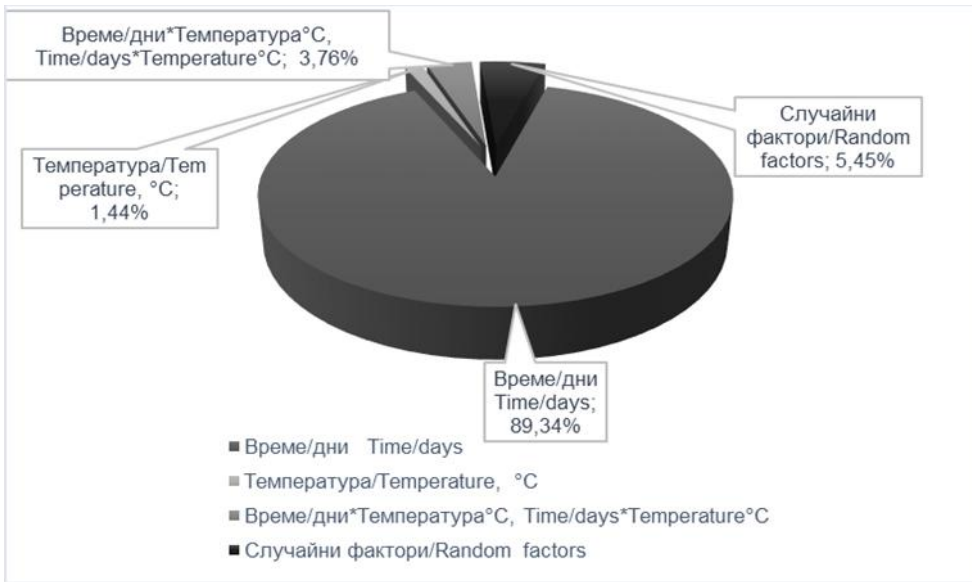


6.

3

Fig. 6. Impact of temperature and storage time factors on the active acidity of Variant 3

1	-	-	In variant 1, the influence of the temperature factor is the highest. It is reported that the interaction between the factors is statistically insignificant at a significance level of 0.05.
2	-	-	The statistical treatment of the results in variant 2 also shows that temperature has a stronger influence on the change in active acidity.
3	-	-	Variant 3 takes into account the influence of the temperature and time factors, but also the mutual influence of the two factors.
7, 9, 11	-	-	Figures 7, 9 and 11 show the data from the statistical processing by two-factor dispersion analysis for the influence of the storage time and temperature factors on the total number of rod-like microorganisms in the lactic acid sample variants.



. 7.

1

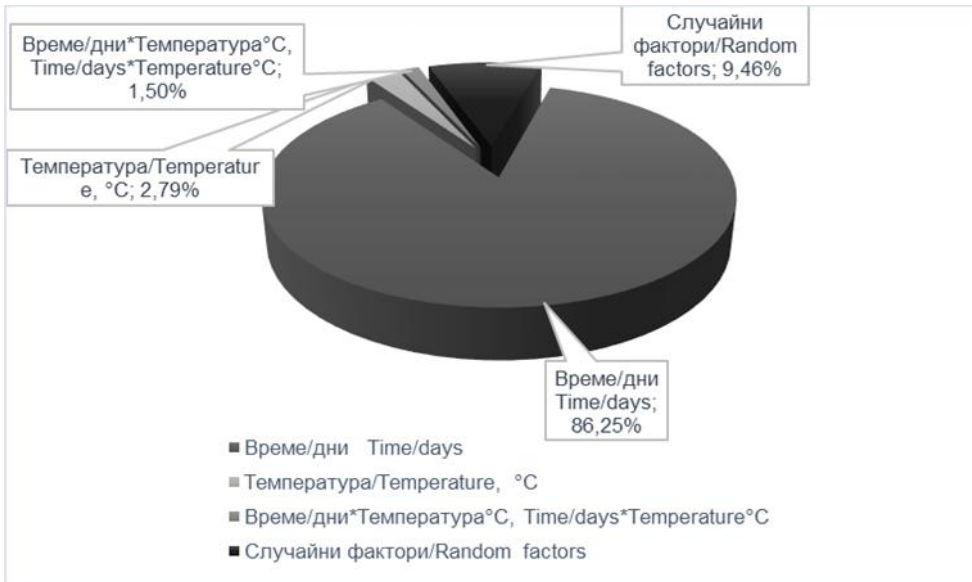
Fig. 7. Impact of temperature and storage time factors on the total number of rod-shaped micro-organisms of Variant 1



. 8.

1

Fig. 8. Impact of temperature and storage time factors on the total number of streptococci of Variant 1



. 9.

2

Fig. 9. Impact of temperature and storage time factors on the total number of rod-shaped micro-organisms of Variant 2



. 10.

2

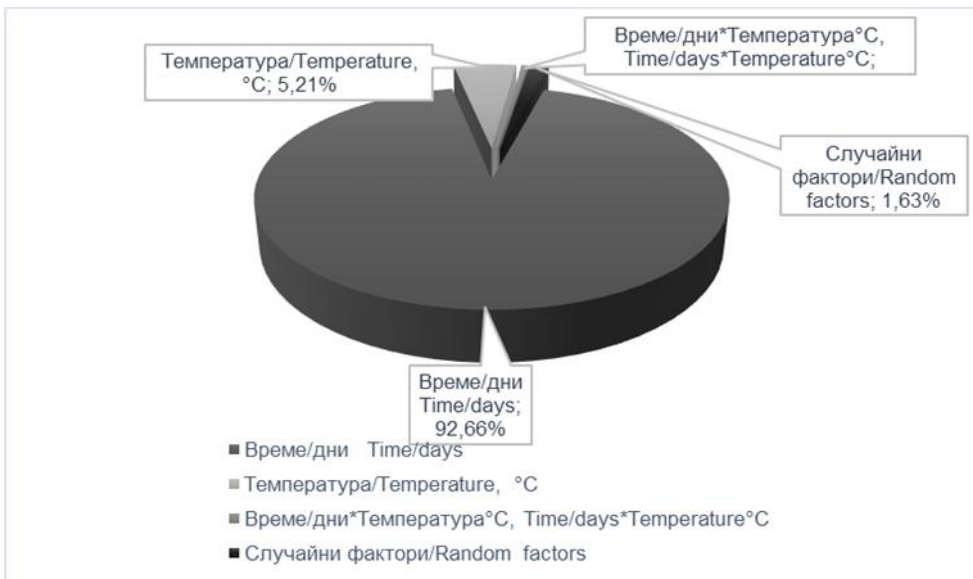
Fig. 10. Impact of temperature and storage time factors on the total number of streptococci of Variant 2



. 11.

3

Fig. 11. Impact of temperature and storage time factors on the total number of rod-shaped micro-organisms of Variant 3



. 12.

3

Fig. 12. Impact of temperature and storage time factors on the total number of streptococci of Variant 3

8, 10 12

0,05.

From the results obtained, we found that the total number of rod-shaped micro-organisms responded only by the storage time factor, and the temperature factor and the interaction between the two factors were statistically insignificant at a significance level of 0.05.

Figures 8, 10 and 12 show statistical data from a two-factor dispersion analysis for the influence of storage time and temperature factors on the total number of streptococci in the lactic acid sample variants.

The results show that the influence of the two factors is statistically significant, but here as well the time factor has a major influence on the number of streptococci in the studied variants.

CONCLUSIONS

The statistical processing of the obtained data by two-factor dispersion analysis indicates that with respect to titratable and active acidity, the temperature factor has a greater impact, whereas the degree of impact of the storage time factor has a major influence on the total number of microorganisms.

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Electrophoretic profile of fermented probiotic products from goat milk

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SUMMARY

The goat milk is a proven healthy product with valuable protein content. Unlike cow milk that can cause allergies and indigestion, goat milk has a higher digestibility and lower allergenic properties. Therefore, due to nutritional and health benefits, the demand for goat milk and goat milk products has increased in recent years.

The purpose of the study is to characterize the protein profiles of different probiotic goat milk products. The products are obtained by selecting and including additional strains to the classic starter and variation and the technological parameters – temperature mode and storage duration.

A series of biochemical tests were performed on the protein spectrum of the test specimens. It was found that the protein profiles of the three probiotic products did not show significant differences. The storage temperatures and period are more important to the

change in the status of protein substances in the test groups studied.

Key words: electrophoretic profile, probiotic products, goat milk

INTRODUCTION

- Goat milk is a proven healthy product with valuable protein content. Recently, the use of goat milk to produce a number of products has become a dynamic and growing industry and is an important part of the economy in many countries (Haenlein, 2005).

(Haenlein, 2005).

- Fermented milk is an extremely appropriate form of absorption of nutrients in milk on the one hand, and of the influence of the health properties of lactic acid bacteria on various functions of the organism, on the other hand (Slacanac et al., 2010).

(Slacanac et al., 2010).

– *Lactobacillus*,
Lactococcus, *Leuconostoc*

- This type of products are the result of fermentation with lactic acid bacteria – *Lactobacillus*, *Lactococcus*, *Leuconostoc* and others, in the process of which transformations of the various milk components are achieved and dairy products with new physiological effects other than the nutritional functions of the raw material are obtained.

- During prolonged storage changes in the composition of probiotic products are inevitable. One of the storage requirements is to preserve to the maximum extent their original chemical and microbiological composition in order not to change their organoleptic qualities and biological value (Champagne, 2005).

(Champagne, 2005).

- The processing of milk and the subsequent storage of fermented probiotic products lead to a number of physicochemical changes in milk proteins. Progressive changes may be: denaturation of whey proteins, interactions between denatured whey proteins and casein micelles. The casein

75-80%

18-20%

(BSA) (Lg), (-La), (Ig)

20%, 10%

50%, 50% (-Lg), 50% (Morr, 1985; Fox and McSweeney, 1998).

- is the main protein component of milk, which represents 75-80% of all milk proteins. Because of its complex composition, casein belongs to the group of phospho-glycoproteins. Also, casein belongs to the group of thermostable proteins because it does not coagulate when subjected to high heat treatment.

The whey protein fraction represents about 18-20% of all milk proteins and contains four major proteins: lactoglobulin (-Lg), lactalbumin (-La), serum albumin (BSA) and immunoglobulin (Ig) and small proteins and enzymes.

These proteins represent respectively 50%, 20%, 10% and 10% whey protein fraction. The degree of whey protein denaturation is often determined by the degree of destruction of -Lg, as it is about 50% of all whey proteins (Morr, 1985; Fox and McSweeney, 1998).

The purpose of this study is to characterize the protein profiles of probiotic products based on goat milk obtained by selecting and including additional strains to classic starter.

MATERIAL AND METHODS

The study used fresh goat milk with pH – 6.70 and °T=18, pasteurized at 93-95 °C for 15 minutes and cooled to 45-46 °C.

The optimal temperature for the fermentation process is 44±2 °C. Lyophilized strains of *Lactobacillus bulgaricus* 1381, *Streptococcus thermophilus* 1374, *Lactobacillus casei* 1014 and *Bifidobacterium longum* 1714 provided by the ICFT collections were used to obtain the probiotic products. The microorganisms are combined in three probiotic combinations containing classical yoghurt starter enriched with strains of lactic acid bacteria in the following ratio:

pH – 6.70 ° =18,
93-95 ° 15 -
45-46 ° . -
44±2 ° . -

Lactobacillus bulgaricus 1381, *Streptococcus thermophilus* 1374, *Lactobacillus casei* 1014
Bifidobacterium longum 1714,

L. bulgaricus: Str.thermophilus, 1:3
(1);
L. bulgaricus: Str. thermophilus: L. casei, 1:3:1 (2);
L.bulgaricus: Str. thermophilus: B. bifidum,1:3:1 (3).

– 1, 2
3
– 5, 10 15°
– 1, 11 21 .

:
1 – 1, 5° ;
2 – 1, 10° ;
3 – 1, 15° ;
4 – 2, 5° ;
5 – 2, 10° ;
6 – 2, 15° ;
7 – 3, 5° ;
8 – 3, 10° ;
9 – 3, 15° .

Laemmli (1970).

Windows 2010. GelAnalyzer
– 6%
10%
: Tris – , 8,5 0,1
% SDS.
– 25 mA.
0,1% Coomassie blue (30-40
min),
24h.
Rf –

L. bulgaricus: Str.thermophilus, 1:3
(Product 1);
L. bulgaricus: Str. thermophilus: L. casei, 1:3:1 (Product 2);
L.bulgaricus: Str. thermophilus: B. bifidum, 1:3:1 (Product 3).

The electrophoretic profiles of the three types of probiotic products – Product 1, Product 2 and Product 3 stored at different temperatures – 5, 10 and 15°C and a storage life of 1, 11 and 21 days are compared.

The following test specimens were studied:

Variant 1 – Product 1, storage temperature 5° ;
Variant 2 – Product 1, storage temperature 10° ;
Variant 3 – Product 1, storage temperature 15° ;
Variant 4 – Product 2, storage temperature 5° ;
Variant 5 – Product 2, storage temperature 10° ;
Variant 6 – Product 2, storage temperature 15° ;
Variant 7 – Product 3, storage temperature 5° ;
Variant 8 – Product 3, storage temperature 10° ;
Variant 9 – Product 3, storage temperature 15° .

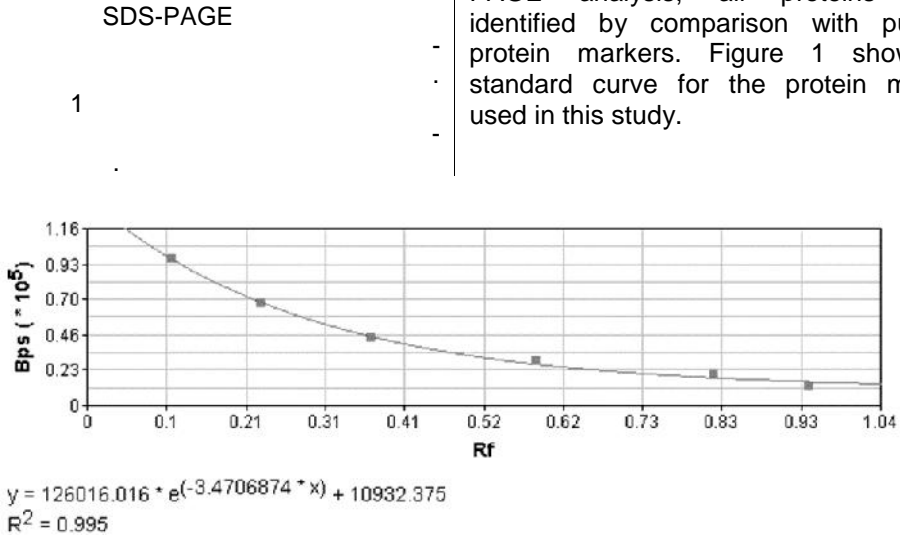
Biochemical tests on the protein spectrum of the test specimens were performed according to the Laemmli method (1970).

The separated gels are analyzed with the GelAnalyzer software for Windows version 2010.

Polyacrylamide gel – 6% stacking and 10% separating. Electrolyte buffer: Tris – glycine, pH 8.5 with 0.1% SDS. Electrophoresis is performed at a current strength of 25 mA. The gel was stained with 0.1% Coomassie blue (30-40 min) and the gel areas lacking protein bands were discolored for 24h. The distances from the start to each strip are measured and the Rf values are determined.

RESULTS AND DISCUSSION

The change in the protein state of the goat yoghurt variants is determined by electrophoretic separation. In the SDS-PAGE analysis, all proteins were identified by comparison with purified protein markers. Figure 1 shows a standard curve for the protein marker used in this study.



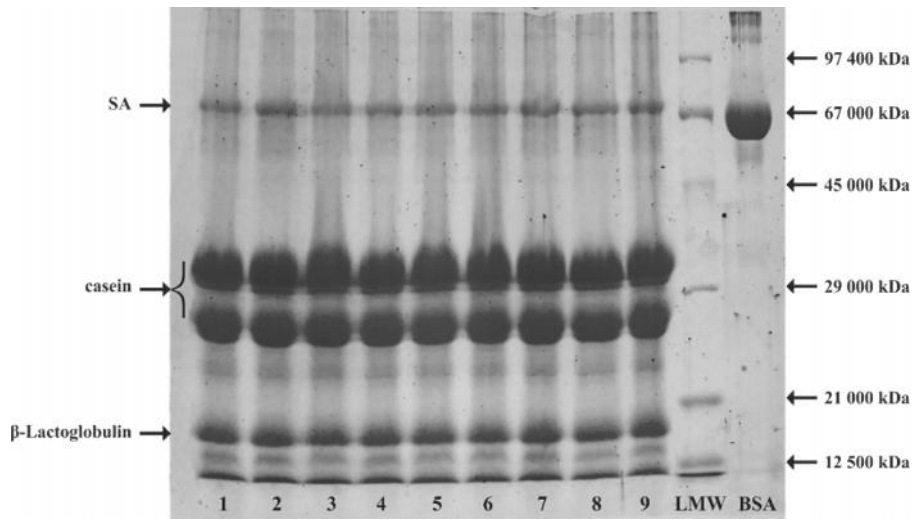
1.
Fig. 1. Standard Protein Marker Curve

– s1, s2,
 ,
 ,
 (Jovanovic et al., 2007).
 2, 3 4

The milk contains four major types of casein proteins – s1, s2, and which differ in composition and amino acid sequence in the polypeptide chain. As mentioned, the milk contains a group of proteins known as whey proteins that can interact and form chemical complexes (Jovanovic et al., 2007).

Figures 2, 3 and 4 show the result of the polyacrylamide gel electrophoresis on goat yoghurt samples after a set storage period at different temperatures of 5, 10 and 15 °C.

5, 10 15 °

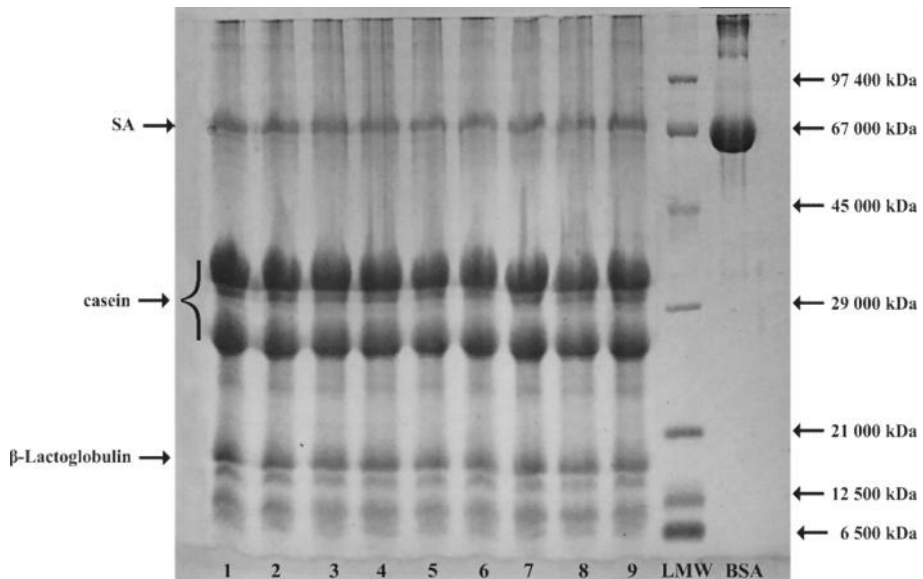


. 2.

- 5, 10 15 °

- 1

Fig. 2. Polyacrylamide gel electrophoresis on goat yoghurt samples stored at 5, 10 and 15 °C for a period of time – 1 day

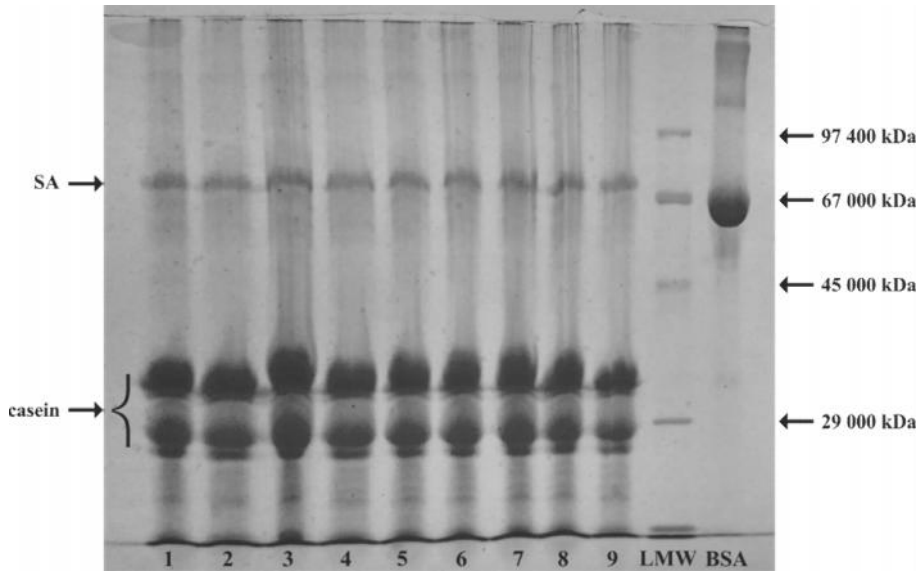


. 3.

- 5, 10 15 °

11

Fig. 3. Polyacrylamide gel electrophoresis on goat yoghurt samples stored at 5, 10 and 15 °C for a period of time – 11 days



. 4.

- 5, 10 15 ° 21

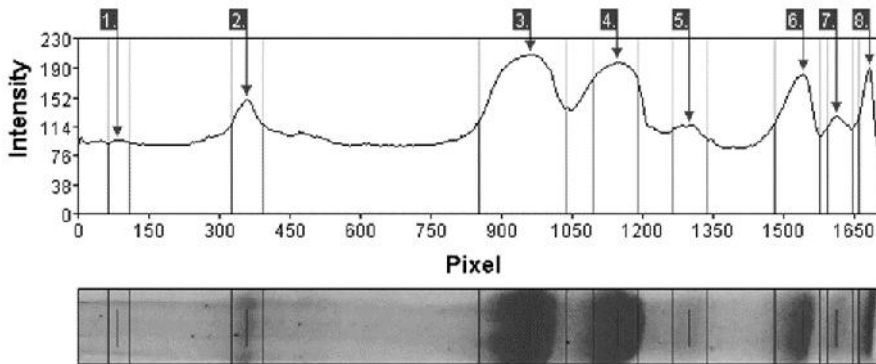
Fig. 4. Polyacrylamide gel electrophoresis on goat yoghurt samples stored at 5, 10 and 15 °C for a period of time – 21 days

	1	1 (1)	Rf
0,985,			
15,2	115 kDa.		0,048
	5		
	5		
-	(29,1 kDa)	-	(23,4
kDa),			
	(16,6 kDa),	-	(14,2
kDa)			
			(71 kDa).

Densitometric analysis determined the electrophoretic motility of the protein fractions of the tested experimental variants.

In Variant 1 (Product 1), after storage for 1 day, Rf values ranging from 0.048 to 0.985, which have molecular weights from 15.2 to 115 kDa, were reported.

Figure 5 shows a densitometric curve of the protein fractions in this variant sample which we assume to be control. From the electrophoretic studies of the protein spectrum it was found that mainly 5 basic bands corresponding to the molecular mass of the following protein fractions were clearly shown: casein fractions -casein (29.1 kDa) and -casein (23.4 kDa) and the whey proteins -lactoglobulin (16.6 kDa), -lactalbumin (14.2 kDa) and serum albumin (71 kDa).



5. **Fig. 5. Densitogram of casein and whey fractions**

Mohd Akmal and Norshafiq (2016) indicate that α -casein and β -casein molecules have determined the following molecular masses, respectively, of 30.2 kDa and 23.9 kDa.

Larson (1985) and Shigeru (1988) have determined the molecular masses of β -lactoglobulin and α -lactalbumin respectively of 18.3 kDa and 14.17 kDa.

(Shauket et al., 1998).

1 11
(2 3).
30 kDa.

SDS-PAGE

Mohd Akmal and Norshafiq (2016) indicate that α -casein and β -casein molecules have determined the following molecular masses, respectively, of 30.2 kDa and 23.9 kDa.

Fractions of analogous molecular mass corresponding to casein proteins were also reported in the present study.

Larson (1985) and Shigeru (1988) have determined the molecular masses of β -lactoglobulin and α -lactalbumin respectively of 18.3 kDa and 14.17 kDa.

Despite the fact that milk proteins have a structural and amino acid similarity, their molecular mass may vary within certain limits even in milk samples of the same species (Shauket et al., 1998).

The results of the electrophoretic examinations of various goat yoghurt variants after 1 and 11 days stay have a similar electrophoretic spectrum as the control one (Figures 2 and 3). For all samples the presence of casein fractions with molecular masses between 20-30 kDa is reported. In the whey proteins are clearly expressed the β -lactoglobulin and serum albumin.

According to SDS-PAGE and the results of the densitometric analysis, the

1 11
 21
 4). 3,
 6 9 (15 °C
 9.
 ()
)

increase in temperature and storage period from 1 to 11 days did not reduce the intensity of the major protein fractions. A lack of co-aggregates and hydrolysates is reported.

Samples stored for 21 days showed a slight change in the type and number of protein bands in respect to the control one (Figure 4). For variants 3, 6 and 9 stored at 15 °C, stretched lines corresponding to casein proteins are reported. It shows a slight decrease in the amount of casein fractions for variant 9.

Also, changes in the protein spectrum of whey proteins are detected (the bands corresponding to α -lactalbumin and β -lactoglobulin are not identified).

CONCLUSIONS

The protein profiles of the three probiotic products based on goat milk obtained by selection and inclusion of additional strains to classical starter did not show significant differences. Factors such as storage and temperature and period are more important for the change in the status of protein substances in the examined test groups

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(IBNA), " " 1, 077015 ,

Some considerations on ruminant feed evaluation by Romanian nutrition system and on diet optimisation

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SUMMARY

- Recent estimations of the feeding value of various Romanian feed resources, based on their chemical analyses, often revealed significant differences from the table feed values. This raised the need to assess these variations and their implications on the formulation of the typical diets for ruminants.

- In this study, 38 bulk feeds were analysed, such as corn, sorghum and alfalfa silage, alfalfa hay (hill and mountain), wheat and barley straws, as well as 30 concentrate feeds (cereal grains): corn, wheat, barley, oats and triticale, and protein feeds, such as soybean meal, peas, bran, etc. The samples were taken from many regions of S, S-E and N-E and also, from the middle of Romania.

Compared to the table references, the assessed nutritive values of the bulk forages ranged between 84% and 101% for energy (NE_m), between 74% and 110% for IDPN and between 79% and 117% for IDPE.

The assessed nutritive values of the concentrate feeds varied from 91% to 102% for the energy, 53% to 125% for IDPN and 96% to 112% for IDPE. These variations may induce biases of up to 30% (such as for IDPN) from the requirements for some categories of ruminant farm animals, with potential negative implications either on the productive levels or feeding efficiencies.

Overall, the study revealed the need to review and update the current table data, in order to align them to the evolution of the nutritive potential of the feedstuffs (e.g. case of corn grains) as well as to the evolution of the methods used for assessing the feed value.

Key words: feed, ruminant, nutritive value

Compared to the table references, the assessed nutritive values of the bulk forages ranged between 84% and 101% for energy (NE_m), between 74% and 110% for IDPN and between 79% and 117% for IDPE.

The assessed nutritive values of the concentrate feeds varied from 91% to 102% for the energy, 53% to 125% for IDPN and 96% to 112% for IDPE. These variations may induce biases of up to 30% (such as for IDPN) from the requirements for some categories of ruminant farm animals, with potential negative implications either on the productive levels or feeding efficiencies.

Overall, the study revealed the need to review and update the current table data, in order to align them to the evolution of the nutritive potential of the feedstuffs (e.g. case of corn grains) as well as to the evolution of the methods used for assessing the feed value.

Key words: feed, ruminant, nutritive value

INTRODUCTION

The Romanian system for assessing the nutritional value of ruminant feeds has emerged as a result of the need to assess their potential for ruminant production as accurate as possible, implicitly their cost, which is reflected considerably in the total cost of animal products.

Initially, in 1983, Burlacu have initiated the Romanian animal nutrition system (including ruminants) taking as a basis a series of data obtained from the experimental determinations in Romania at IBNA Balotesti Institute, but at the same time, having reference to the known nutritional systems at that time – Oscar Kellner Institute in Germany (OKIT) in 1970, Agricultural Research Council (ARC) in UK in 1975, National Research

1983 . Burlacu

(IBNA)

"

(OKIT)

1970 ..

(ARC)		1975	Council (NRC) in USA in 1976 and Institut National de la Recherche Agronomique (INRA) in France in 1978. Within this system Burlacu (1983) took into account the average of the feed nutrition values in the foreign systems, but also their energy value, expressed differently, as for example: net energy for fat (as OKIT), net energy for maintenance and production (as ARC and NRC, 1976) and net energy for milk (INRA, 1978).
„	(NRC)	1976	
(INRA)		1978	
		Burlacu (1983)	
(OKIT),	(ARC NRC, 1976)		
1978).	(INRA,		
	2002		
(Burlacu et al., 2002).			
	Burlacu et al. (2006).		
PDI	INRA (Verité et al., 1987)		
			The Romanian nutrition system was updated in 2002 by adding some equations or coefficients allowing a more accurate assessment (Burlacu et al., 2002). Modern techniques have also been introduced to measure the various components of the system in order to increase accuracy of the feed evaluation. For energy and protein metabolism in cattle, even modelling has been done as presented by Burlacu et al. (2006). Briefly, we mention that the energy of the feeds are reported as net energy, more precisely as nutritive units compared to the energy content of standard barley, and the protein feed value is expressed as in the PDI system of INRA (Verité et al., 1987) by protein digested in small intestine.
			In current animal husbandry practice in Romania, we found differences between the table reference values and those determined for the energy and protein content of many feed ingredients used for ruminants feeding.
			At the same time, it could be noticed that the data of the nutritional systems of different countries are reviewed periodically due to changes caused by climate change, soil, the degree of fodder plants amelioration, the appearance of many by-products for feed use.
			This is the reason that Romanian reference values need to be updated. In this context, this study aims to quantify

deviations of nutritional value from table references, but also to estimate the implications of these deviations for optimizing rations for farm ruminants.

MATERIAL AND METHODS

Presenting the feed system

The system is based on inventory of equations, which allows differentiation of intermediate calculation steps.

a. Indicators for feed energy content are: gross energy (GE), digestible energy (ED), metabolizable energy (EM) and net energy (NE), expressed by feed unit for lactation (UFL) and feed units for meat production (UFV).

The GE was determined by adiabatic calorimetry but in the present it is calculated by equations where the independent variables are the chemical composition nutrients (grams/ kg feed dry matter DM). For concentrates, the equation of Schiemann et al. (1971) is applied:

$$GE \text{ (kJ/kg DM)} = 23.95 \times CP + 39.77 \times EE + 20.05 \times CF + 17.46 \times NFE + h,$$

$$h = \frac{GE - 23.95 \times CP - 39.77 \times EE - 20.05 \times CF - 17.46 \times NFE}{1}$$

Demarquilly et al., (1978),
Andrieu and Demarquilly (1987)

ED
GE
Hoffmann and Schiemann (1980)

where h is a variable coefficient for different feeds and CP=crude protein, EE=ether extract, CF=crude fiber, NFE=nitrogen-free extractives.

For fresh forage and hay the GE is calculated using Demarquilly et al. (1978) equation, and for silage the Andrieu and Demarquilly (1987) equation.

Initially, the ED was determined directly by experiments on digestibility cages as difference between the GE and faeces energy. Other method was the calculation by Hoffmann and Schiemann (1980) formula and using the digestible part of the nutrients:

$$ED \text{ (kJ/kg DM)} = 24.2 \times CPD + 34.1 \times EED + 18.4 \times CFD + 17 \times NFED,$$

CPD = , where CPD= digestible crude protein,

EED = , CFD =
 ; NFED =
 (Sauvant et al., 1987)
 (DOM),
 % ED = % DOM + h,
 DOM
 Tilley and Terry (1963) *in vitro*
 Pepsin-Cellulase (Aufreere, 2007)
 GE
 400 l 24
 50 l (Burlacu, 1985).
 Burlacu
 (1985),
 OKIT:

EED= digestible ether extract, CFD= digestible crude fiber and NFED= digestible nitrogen-free extractives; these values are valid in tables for various categories of feed. Another equation (Sauvant et al., 1987) used the digestible organic matter (DOM) as follows:

% ED = % DOM + h, where h is a variable coefficient for different feeds.

In the present, DOM is evaluated by Tilley and Terry (1963) *in vitro* method and we initiate the application of the Pepsin-Cellulase method (Aufreere, 2007) for highly reproducible results.

The EM is the difference between GE and the sum of the energy lost by faeces, gas fermentation and urine. The production of fermentation gases depends on the composition of the ration, the intake, the age and the production of the animal, reaching 400 L in 24 hours for cows and 50 L for sheep (Burlacu, 1985). The energy for urine varies according to the age of the animal, its physiological state, the level of production and the nitrogen content of the ration. Also, the energy of urine is higher in fattening animals, in gestation and lactation, generally in those with higher production levels. For EM calculation the equation of Burlacu (1985) was applied which was similar with that of the OKIT system:

$$EM \text{ (kJ /kg DM)} = 17.32 \times CPD + 30.9 \times EED + 16.15 \times (CFD + NFED) \pm 1.66 \text{ MJ}$$

Vermorel et al. (1987)

A more rapid calculation is the formula of Vermorel et al. (1987):

$$EM/ED = 0.8417 - 0.000099 \times CFo - 0.000196 \times CPo + 0.0221 \times APL,$$

CFo= g CF/kg OM, CPo= g CP/kg OM, APL = 1.5.

where CFo= g CF/kg OM, CPo= g CP/kg OM, and APL = animal production level settled at 1.5.

NE . Burlacu et al.,

The NE is the difference between EM and caloric energy. Burlacu et al. measured the caloric energy between 1980 and

1980	1985	IBNA	1985 in the IBNA Balotesti Institute by respiratory changes method in special chambers with open flow.
	, NE	-	Today, the NE is calculated in different ways for milk production and for meat production. First, the energy content of the feed is calculated: $q = EM/GE$, and then the efficiency of EM utilisation (K) as net energy for maintenance (NEm), net energy for meat production (NEp) and net energy for lactation (NEI) for a standard APL of 1.5 (Vermorel et al., 1987).
	: $q = EM/GE$	-	
	() (NEm), (NEp) (NEI)	-	
APL 1.5 (Vermorel et al., 1987).		-	
NEm = EM x Km, Km = 0.287 q + 0.554		-	NEm = EM x Km, where Km = 0.287 q + 0.554, is the efficiency for maintenance;
NEp = EM x Kp, Kp = 0,78 q + 0,006		-	NEp = EM x Kp, where Kp = 0.78 q + 0.006, is the efficiency for meat production;
NEI = EM x KI, KI = 0,24 q + 0,463		-	NEI = EM x KI, where KI = 0.24 q + 0.463, is the efficiency for lactation.
	NE	-	In Romania we express NE in feed unit for lactation (UFL) and feed units for meat production (UFV) referring to the energy content of standard barley. The GE of 1 kg of barley is 16.69 MJ (or 3989 kcal) and the ED is 71.5% of GE, that means 11.94 Mj (2853 kcal).
(UFL)	(UFV),	-	
. GE 1 16,69		-	
MJ (3989 kcal), ED 71,5%		-	
GE, . . 11,94 Mj (2853 kcal).		-	
q 0.599, EM 10 Mj		-	The q value is 0.599, so the EM is 10 MJ (2414 kcal). The KI capacity is 0.607, and Kmp is 0.616. Harkins et al. (1974)
(2414 kcal). KI 0.607,		-	calculated the overall efficiency for maintenance and meat production, Kmp. The result is net energy for lactation of 6.07 MJ (1436 kcal) considered 1 UFL, and net energy for maintenance and meat production of 6.16 MJ (1472 kcal) considered 1 UFV.
Kmp 0.616. Harkins et al. (1974)		-	
	, Kmp.	-	
	6.07 Mj (1436 kcal),	-	
	1 UFL,	-	
6.16 Mj (1472 kcal),		-	
b.	1 UFV.	-	
		-	b. The protein value of the feeds expressed in the terms of the PDI system of INRA is the sum of two fractions: IDPN (intestinally digestible protein allowed by nitrogen) and IDPE (intestinally digestible protein allowed by energy).
	INRA,	-	
: IDPN ()		-	
IDPE ()		-	
).	-	
	, PDIA.	-	Both of these energies include the digestible part of the dietary protein, undegraded in the rumen, noted PDIA.

$$PDIA \text{ (g/kg DM)} = PB \text{ (g/kg DM)} \times (1 - Dg) \times dsi$$

Dg

Alderman (1993)
SACCO
:

- Dg is the protein degradability in the rumen and is calculated by Alderman (1993) equation after testing feed by *in sacco* method on fistulized cows:

$$Dg = a + (b \times c) / (c + r)$$

=

(%)

b =

(%)

c =
(h⁻¹)

r =

(ruminal outflow rate) (

, 0,019 APL

() 0,104

APL 4,5 (

)

dsi =

where:

a = rapidly degradable protein fraction (%)

b = potentially degradable protein fraction (%)

c = rate at which b is degraded (h⁻¹)

r = the ruminal outflow rate (is variable; from 0.019 for APL of value 1 (for maintenance) to 0.104 for a maximum APL of 4.5 (for high-productivity dairy cows))

- dsi = true digestibility of undegraded dietary protein in small intestine.

PDIM

PDIM,

PDIMN,

PDIM,

PDIME.

PDIM is the microbial true protein which is truly digestible in the small intestine. The part of the PDIM that could be synthesized in the rumen from the degraded dietary nitrogen is named PDIMN and the other part of the PDIM that could be synthesized by energy available in the rumen is called PDIME.

So, the value of each feed is given directly as the sum of PDIA and PDIM.

PDIM.

PDIA

$$IDPN = PDIA + PDIMN$$

$$PDIMN \text{ (g/kg DM)} = PB \text{ (g/kg DM)} \times (1 - (1 - Dg)) \times 0.9 \times 0.8 \times 0.8 = 0.576 \times CP \times Dg,$$

90%

80%

80%

where 90% of the protein is captured in the rumen and microbial protein has 80% aminoacids with 80% digestibility.

$$IDPE = PDIA + PDIME$$

$$PDIME \text{ (g/kg DM)} = FOM \times 0.145 \times 0.8 \times 0.8 = 0.093 \times FOM,$$

FOM

where FOM is the fermentable organic matter used for 145 g of microbial protein

145 g
FOM
DOM :

synthesis. FOM is calculated from the DOM content by formula:

$$FOM = DOM - [CP \times (1 - Dg) + EE + PF],$$

PF

- where PF are the fermentation products for different silages categories and are presented as table values by Burlacu et al. (2002).
-

Burlacu et al. (2002).

RESULTS AND DISCUSSION

Deviations of the feed nutritive values

- In a recent study 38 bulk feeds were analysed, such as corn, sorghum and alfalfa silage, alfalfa hay (hill and mountain), wheat and barley straws, as well as 30 concentrate feeds (cereal grains): corn, wheat, barley, oats and triticale, and protein feeds, such as soybean meal, peas, bran, etc. The samples were taken from regions of S, S-E and N-E and also, from the middle of Romania.

- First, the chemical composition of the feed samples was determined by the commonly accepted methods (EC 152/2009): dry matter, crude protein, crude fibre, ether extractives, ash and nitrogen-free extractives. The DOM value for each feed resulted from Tilley and Terry (1963) method applied on Ankom Daisy Incubator. The feed Dg values were determined by *in sacco* method.

- The energy and protein feeding values of all the feeds were calculated by the system of Burlacu et al. (2002), presented above. We compared these values with the reference from Burlacu et al. (2002). In Table 1 are presented 4 commonly used feed for ruminants and their nutritive values. For the bulk forages (alfalfa hay and corn silage), the assessed nutritive values ranged between 84% and 101% for energy (NE_m), between 74% and 110% for IDPN and between 79% and 117% for IDPE. For the concentrate

38

; 30

152/2009):

DOM

Tilley and Terry

(1963),

Ankom Daisy.

Dg

SACCO.

(2002),

Burlacu et al.

Burlacu et al. (2002).

1

4

(

84% 101%

(NE_m),

74% 110%

IDPN

79% 117% IDPE.
, (
)
91% 102%
53% 125% IDPN 96% 112%
IDPE.
(
).

feed (corn and soybean meal) the assessed nutritive values varied from 91% to 102 % for the energy, 53% to 125% for IDPN and 96% to 112% for IDPE. We observed also, the same discrepancies for another feeds (not presented).

1.

4

-

Table 1. Examples of nutritive values of the 4 commonly used feeds

	UFL	IDPN (g)	IDPE (g)	Ca (g)	P(g)
/ Concentrates					
- / Corn – reference values	1.43	96	116	0.3	2.9
Corn – 8 samples – obtained values	min. 1.30	51	111	0.2	2.0
	max. 1.46	74	130	1.2	4.1
- / Soya meal – reference values	1.37	326	211	3.6	6.4
Soya meal – 6 samples – obtained values	min. 1.33	309	205	2.0	6.3
	max. 1.39	408	214	3.5	7.8
/ Forages					
- /Alfalfa hay – reference values	0.76	107	68	14.9	2.9
Alfalfa hay – 16 samples – obtained values	min. 0.64	79	54	7.5	2.4
	max. 0.76	118	73	17.4	4.6
- /Corn silage – reference values	1.05	46	68	4.3	2.3
Corn silage – 8 samples – obtained values	min. 0.88	34.6	50	3.7	2.1
	max. 1.06	50.1	79.6	5.4	2.4

1 UFL = 6,07 MJ =

IDPN =

by dietary nitrogen content

IDPE =

allowed by dietary energy content

Ca = / calcium, g

P = / phosphorus, g

/ feed unit for lactation

/ intestinally digestible protein allowed

/ intestinally digestible protein

550 kg
20 l
4%,
15.59 UNL 1325 g PDI.
2,
: 2,75 kg
, 7,01 kg
, 3,08 kg 1,35 kg
IDPN IDPE,
UFL 15.71,

Impact assessment for obtained nutritive feed values

Using the above presented feeds as ingredients we simulated a ration for a 550 kg cow in mid lactation and 20 milk litres daily production, with fat 4%, indoor rearing. The norm is established for 15.59 UFL and 1325 g PDI. As presented in Table 2 the quantities, in DM for each ingredient, are: 2.75 kg alfalfa hay, 7.01 kg corn silage, 3.08 kg corn and 1.35 kg soybean meal. It can be observed that is a stable balance between IDPN and IDPE, their values are aproximately the same and near the established value. The UFL value is 15.71, also near to the established one.

2. 550 , 20 L
4% () -

Table 2. Diet formulation for a 550 kg cow in mid lactation, with 20L/day milk and 4% fat (indoor rearing) – ingredients with normal feeding values

/Ingredients	supply (as is basis), kg	DM (g)	UFL	IDPN (g)	IDPE (g)	Ca (g)	P (g)
/alfalfa hay		850	0.76	107	68	14.9	2.9
/corn silage		279	1.05	46	68	4.3	2.3
/corn grains		873	1.43	96	116	0.3	2.9
/soybean meal		890	1.37	326	211	3.6	6.4
dicalcium phosphate		990	0	0	0	39.7	22.6
/salt		900	0	0	0	0	0
specific minerals (Zoofort)		900	0	0	0	0	0
550 kg/20 L NORM for cow 550 kg/20 L:		16400	15.59	1325	1325	102	65
/Forages							
/Alfalfa hay	3.24	2754	2.093	294.678	187.272	41.035	7.987
/Corn silages	25.14	7014.06	7.365	322.647	476.956	30.160	16.132
/Total forages	28.38	9768.06	9.458	617.325	664.228	71.195	24.119
Compound feeds							
/Corn grains	3.53	3081.69	4.407	295.842	357.476	0.925	8.937
/Soybean meal	1.52	1352.8	1.853	441.013	285.441	4.870	8.658
dicalcium phosphate	0.67	663.3	0.000	0.000	0.000	26.333	14.991
/Salt	0.0705	63.45	0.000	0.000	0.000	0.000	0.000
Specific minerals (Zoofort)	0.0468	42.12	0.000	0.000	0.000	0.000	0.000
Total compounds	5.8373	5203.36	6.260153	736.855	642.9168	32.1276	32.5854
/TOTAL supply		14971.42	15.71796	1354.18	1307.145	103.3227	56.70434
/covering		91.3%	100.8%	102.2%	98.7%	101.3%	87.2%

3 -
,
.
,
78,4% IDPN
84,6% IDPE.
(88,1%. PDI
1080 g) 13,74 UFL

In Table 3 is presented a theoretical ration for the same cow but using ingredients with minimum nutritive values. The deficit of the ingredients nutritive values induces the deficit for ration because only 78.4% of IDPN and 84.6% of IDPE are covered. The energy is covered only 88.1%.
For the resulted PDI (1080 g mean value) and 13.74 UFL, the expected milk

20 l. 15 l
5 l

production of the cow could be only 15 L instead of 20 L. This is a loss of 5 L for animal daily productivity.

3. 550 kg,
, 20 l/ 4% ()-

Table 3. Diet formulation for a 550 kg cow in mid lactation, with 20L/daymilk and 4% fat (indoor rearing) – ingredients with deficient feeding values

/Ingredients	supply (as is basis), kg	DM (g)	UFL	IDPN (g)	IDPE (g)	Ca (g)	P (g)
alfalfa hay		850	0.64	79	55	7.5	2.4
/alfalfa hay		279	0.88	35	50	3.7	2.14
/corn silage		873	1.3	51	111	0.2	2
/corn grains		890	1.33	309	205	2	6.3
/soybean meal		990	0	0	0	39.7	22.6
/dicalcium phosphate		900	0	0	0	0	0
/salt		900	0	0	0	0	0
specific minerals (Zoofort)							
550 kg /20 l NORM for cow 550 kg / 20 l:		16400	15.59	1325	1325	102	65
/Forages							
/Alfalfa hay	3.24	2754	1.763	217.566	151.470	20.655	6.610
/Corn silages	25.14	7014.06	6.172	245.492	350.703	25.952	15.010
/Total forages	28.38	9768.06	7.935	463.058	502.173	46.607	21.620
Compound feeds							
/Corn grains	3.53	3081.69	4.006	157.166	342.068	0.616	6.163
/Soybean meal	1.52	1352.8	1.799	418.015	277.324	2.706	8.523
dicalcium phosphate	1.3	1287	0.000	0.000	0.000	51.094	29.086
/Salt	0.0705	63.45	0.000	0.000	0.000	0.000	0.000
Specific minerals (Zoofort)	0.0468	42.12	0.000	0.000	0.000	0.000	0.000
/Total compounds	6.4673	5827.06	5.805421	575.181	619.3916	54.41584	43.77222
TOTAL supply		15595.12	13.74035	1038.23	1121.565	101.0229	65.39191
/covering		95.1%	88.1%	78.4%	84.6%	99.0%	100.6%

4

15.92 UFL (9.8%
15.59),

The Table 4 is for another theoretical ration for which the same ingredients had maximum nutritive values, mainly higher than the reference ones. Even there are 15.92 UFL (close to required 15.59), the protein is 9.8% excessively. This is a waste of protein and consequently, a waste of cost for the same milk production.

4. 550 kg,
 , 20 l/ 4% () –

Table 4. Diet formulation for a 550 kg cow in mid lactation, with 20L/day milk and 4% fat (indoor rearing) – ingredients with feeding values in excess compared to the reference values

/Ingredients	supply (as is basis), kg	DM (g)	UFL	IDPN (g)	IDPE (g)	Ca (g)	P (g)
/alfalfa hay		850	0.76	118	73	17.4	4.6
/corn silage		279	1.06	50	80	5.42	2.45
/corn grains		873	1.46	74	130	1.2	4.1
/soybean meal		890	1.4	407	215	3.5	7.87
dicalcium phosphate		990	0	0	0	39.7	22.6
/salt		900	0	0	0	0	0
specific minerals (Zoofort)		900	0	0	0	0	0
550 kg/20 l NORM for 550 kg cow/20 l:		16400	15.59	1325	1325	102	65
/Forages							
/Alfalfa hay	3.24	2754	2.093	324.972	201.042	47.920	12.668
/Corn silages	25.14	7014.06	7.435	350.703	561.125	38.016	17.184
/Total forages	28.38	9768.06	9.528	675.675	762.167	85.936	29.853
/Compound feeds							
/Corn grains	3.53	3081.69	4.499	228.045	400.620	3.698	12.635
/Soybean meal	1.52	1352.8	1.894	550.590	290.852	4.735	10.647
dicalcium phosphate	0.35	346.5	0.000	0.000	0.000	13.756	7.831
/Salt	0.0705	63.45	0.000	0.000	0.000	0.000	0.000
Specific minerals (Zoofort)	0.0468	42.12	0.000	0.000	0.000	0.000	0.000
Total compounds	5.5173	4886.56	6.393187	778.634	691.4717	22.18888	31.11237
TOTAL supply		14654.62	15.92113	1454.31	1453.639	108.1247	60.96521
/covering		89.4%	102.1%	109.8%	109.7%	106.0%	93.8%

- If you do not take into account the
 - correct nutritional values, determined
 - each time they are used, we can believe
 - that we have provided the nutritional
 - needs for ration, but in fact they are not
 - ensured because the ingredients are
 - nutritionally deficient and we could expect
 - an under 25 % animal production.

25%

IDPN) 30% (

- The variability of feed could induce
- biases of up to 30% (such as for IDPN)
- from the requirements for some
- categories of ruminant farm animals, with
- potential negative implications either on
- the productive levels or feeding
- efficiencies. If, on the contrary, we have
- ingredients with higher nutritional values
- than references, we can use the
- ingredients in surplus and therefore we
- can have higher costs for the same
- production. The weight of the costs
- depends on the type of ingredients and
- are not negligible for a medium-size or
- big farm.

Dragomir et al. (2011)

At the same time, adequate nutrition of farm animals must also take into account the appearance of many by-products for feed use. It is necessary that they be nutritionally characterized and their use is advisable to occur after their nutritional value is known. For example, in paper of Dragomir et al. (2011) the main corn grain processing by-products are presented, their nutritive value, and their effects on the energy and protein metabolism in the rumen. The paper presents particularities of the rumen protein degradability of the mentioned by-products.

22% IDPN

25%

CONCLUSIONS

In the presented ration a 22% deficit of IDPN could produce a 25% decrease in milk production. If the nutritive feeds values are over the references than a waste of costs could appear. This is a suggestion that the feeds must be evaluated before calculation of the ration. Also, the study revealed the need to review and update the current Romanian reference data, in order to align them to the evolution of the nutritive potential of the feedstuffs as well as to the evolution of the methods used for assessing the feed value.

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